

TURUN YLIOPISTON JULKAISUJA
ANNALES UNIVERSITATIS TURKUENSIS

SARJA - SER. D OSA - TOM. 955

MEDICA - ODONTOLOGICA

**HYPOXIA-ASSOCIATED BIOMARKERS
IN RECTAL CANCER TREATED BY
PREOPERATIVE RADIOTHERAPY
OR CHEMORADIOTHERAPY**

by

Eija Korkeila

TURUN YLIOPISTO
UNIVERSITY OF TURKU
Turku 2011

From the Department of Oncology and Radiotherapy and the Department of Pathology, University of Turku, Turku, Finland

Supervised by:

Professor Seppo Pyrhönen, MD, PhD
Department of Oncology and Radiotherapy
University of Turku, Turku, Finland

Docent Jari Sundström, MD, PhD
Department of Pathology
University of Turku, Turku, Finland

Reviewed by:

Professor Tuomo Karttunen, MD, PhD,
Department of Pathology
University of Oulu, Oulu, Finland

Docent Ilmo Kellokumpu, MD, PhD
Department of Surgery
Central Finland Central Hospital, Jyväskylä, Finland

Dissertation opponent:

Docent Petri Bono, MD, PhD
University of Helsinki, Helsinki, Finland

ISBN 978-951-29-4573-3 (PRINT)

ISBN 978-951-29-4574-0 (PDF)

ISSN 0355-9483

Painosalama Oy – Turku, Finland 2011

To Petri, Karoliina, Petra and Tiltu

ABSTRACT

Eija Korkeila:

Hypoxia-associated biomarkers in rectal cancer treated by preoperative radiotherapy or chemoradiotherapy

Department of Oncology and Radiotherapy and Department of Pathology, Turku University Hospital and University of Turku, Finland
Annales Universitatis Turkuensis 2011

Rectal cancer (RC) is a common malignancy. Preoperative radiotherapy (RT) is usually given to patients with T3-T4 tumours. The aim of the study was to assess hypoxia-inducible factor-1 α (HIF-1 α), carbonic anhydrase IX (CA IX), ezrin and glucose transporter-1 (GLUT-1) expression as predictors of disease-free survival (DFS) and disease-specific survival (DSS) in RC treated by preoperative radiotherapy (RT) or chemoradiotherapy.

Diagnostic biopsies (n=80) and corresponding operative samples from 178 consecutive RC patients, treated by short- (n= 77) or long-course RT with (n=37) or without (n=10) chemotherapy or no treatment preoperatively (n=54), were analysed for HIF-1 α , CA IX, ezrin and GLUT-1 using immunohistochemistry (IHC). Tumour regression grade (TRG) was analysed after long-course RT.

In operative samples, negative/weak (N/W) CA IX staining intensity was associated with favourable DFS ($p=0.003$) and DSS ($p=0.034$). After long-course RT, negative HIF-1 α expression was linked to longer DSS ($p= 0.001$) and N/W GLUT-1 (0.066) with longer DFS than with positive or strong expression of these markers. Moderate/strong (M/S) ezrin expression in biopsies was associated with unfavourable DFS ($p=0.027$) and DSS ($p= 0.002$). In multivariate analysis, M/S CA IX intensity in operative samples was an independent predictor of poor DFS and DSS. Excellent TRG was linked to N/W CA IX ($p=0.057$), ezrin ($p=0.012$) and GLUT-1 ($p=0.013$) in operative samples. In multivariate model with all four markers, CA IX intensity in operative samples independently predicted DSS.

In conclusion, positive HIF-1 α expression and M/S expression of CA IX and GLUT-1 in operative samples as well as M/S ezrin in biopsies were related to unfavourable disease outcome. CA IX intensity in operative samples independently predicted DFS and DSS in multivariate analysis. Moderate/strong CA IX intensity is a powerful predictor of poor disease outcome in RC.

Key words: rectal cancer, chemotherapy, radiotherapy, prognosis, predictive factor, tumour regression, hypoxia-inducible factor-1 α , carbonic anhydrase IX, ezrin, GLUT-1

TIIVISTELMÄ

Eija Korkeila:

Hypoksiaan liittyvät biologiset merkkiaineet leikkausta edeltävällä sädehoidolla tai kemosädehoidolla hoidetussa peräsuolisyövässä

Syöpätautien ja sädehoidon sekä Patologian oppiaine, Turun yliopisto, Turku
Annales Universitatis Turkuensis 2011

Peräsuolensyöpä on yleinen pahanlaatuinen kasvain. Leikkausta edeltävä sädehoito annetaan yleensä T3-T4-kasvaimille. Tutkimuksella pyrittiin selvittämään, voidaanko kasvaimen hapenpuutteeseen liittyvillä biologisilla merkkiaineilla arvioida peräsuolisyövän ennustetta leikkausta edeltävän sädehoidon tai kemosädehoidon jälkeen. Tällaisia merkkiaineita ovat hapenpuutteen vaikutuksesta aktivoituva HIF-1 α , hiilihappoanhydraasi IX (CA IX), sokerin kuljetukseen solussa osallistuva GLUT-1 sekä solun tukirankaproteiini ezrin.

Tutkimukseen otettiin 178 potilasta, jotka olivat saaneet ennen leikkausta lyhyen (n=77) tai pitkän sädehoidon (n=10), pitkän sädehoidon ja solunsalpaajahoidon (n=37) tai ei mitään hoitoa (n=54). Lisäksi osalta leikkausta edeltävää sädehoitoa saaneelta potilaalta tutkittiin hoitoja edeltävät, diagnostiset näytteet (n=80). Tutkimuksessa käytettiin immunehistokemiallisia värjäysmenetelmiä. Kasvaimen regressiota (TRG) arvioitiin pitkän sädehoidon jälkeisistä näytteistä.

Leikkauksnäytteissä negatiivinen/heikko CA IX intensiteetti liittyi sekä pidempään tautispesifiseen ($p=0.034$) että tautivapaaseen elinaikaan ($p=0.003$) ja pitkän sädehoidon jälkeen HIF-1 α -negatiivisuus pidempään tautispesifiseen ($p=0.001$) sekä negatiivinen/heikko GLUT-1 pidempään tautivapaaseen elinaikaan ($p=0.066$). Voimakas ezrin-ilmentymä diagnostisissa näytteissä liittyi lyhyempään tautivapaaseen ja tautispesifiseen ($p=0.027$ ja $p=0.002$) ennusteeseen. Monimuuttuja-analyysissä vahva CA IX intensiteetti leikkauksnäytteissä ennusti itsenäisesti huonompaa tautivapaata ja tautispesifistä selviytymistä. Erinomainen TRG liittyi negatiiviseen/heikkoon CA IX- ($p=0.057$), ezrin- ($p=0.012$) ja GLUT-1 -ilmentymään ($p=0.013$) leikkauksnäytteissä. Kun kaikki neljä merkkiainetta analysoitiin yhdessä monimuuttuja-analyysissä, CA IX intensiteetti leikkauksnäytteissä ennusti itsenäisesti tautispesifistä elinaikaa.

Voimakas CA IX-ilmentymä leikkauksnäytteissä ja positiivinen HIF-1 α - ja vahva GLUT-1-ilmentymä pitkän sädehoidon jälkeisissä leikkauksnäytteissä sekä vahva ezrin-ilmentymä diagnostisissa näytteissä liittyivät epäsuotuisaan ennusteeseen. Monimuuttuja-analyysissä kohtalainen/voimakas CA IX intensiteetti leikkauksnäytteissä ennusti itsenäisesti huonompaa tautivapaata ja tautispesifistä elinaikaa. CA IX on vahva biologinen merkkiaine peräsuolisyövässä.

Avainsanat: peräsuolisyöpä, solunsalpaajahoito, sädehoito, ennuste, ennusteellinen tekijä, kasvaimen regressio, hapenpuutteen vaikutuksesta aktivoituva tekijä 1 α , hiilihappoanhydraasi IX, ezrin, GLUT-1.

TABLE OF CONTENTS

ABSTRACT	4
TIIVISTELMÄ	5
TABLE OF CONTENTS	6
ABBREVIATIONS	10
LIST OF ORIGINAL PUBLICATIONS	13
1. INTRODUCTION	14
2. REVIEW OF THE LITERATURE	16
2.1. GENERAL ASPECTS OF COLORECTAL CANCER	16
2.1.1. Epidemiology of CRC	16
2.1.2. Aetiology of CRC.....	16
2.1.2.1. Sporadic CRC	16
2.1.2.1.1. Carcinogenesis.....	16
2.1.2.1.2. Adenoma-carcinoma sequence	17
2.1.2.1.3. Serrated adenoma pathway	17
2.1.2.1.4. Genomic instability in sporadic CRC	17
2.1.2.2. Familial and hereditary CRC	17
2.1.2.3. Predisposing conditions	18
2.1.2.4. Nutritional and lifestyle factors.....	18
2.1.2.5. Other risk factors.....	19
2.2. RECTAL CANCER.....	19
2.2.1. Signs and symptoms of disease.....	19
2.2.2. Diagnostic and screening procedures	20
2.2.3. Pathology and classification	20
2.2.3.1. Histopathology	20
2.2.3.2. Handling and evaluation of the specimen.....	20
2.2.3.3. Circumferential margin	21
2.2.3.4. Lymph node evaluation	21
2.2.3.5. Tumour differentiation grade.....	21
2.2.3.6. Perineural and vascular invasion.....	22
2.2.4. Tumour response to preoperative treatment.....	22
2.2.4.1. Methods for predicting response to treatment	23
2.2.4.2. Tumour regression grade	23
2.2.4.3. Imaging in predicting response to treatment.....	23
2.2.5. Staging.....	24
2.2.5.1. Preoperative staging	24

Table of Contents

2.2.5.2. Pathology report and postoperative staging.....	24
2.2.6. Surgical treatment of localised rectal cancer	28
2.2.6.1. Anterior resection and abdominoperineal resection	28
2.2.6.2. Total mesorectal excision technique.....	28
2.2.6.3. Laparoscopic surgery.....	28
2.2.6.4. Local excision.....	28
2.2.7. Oncological treatments of rectal cancer	30
2.2.7.1. General aspects of radiotherapy.....	30
2.2.7.2. Preoperative short-course radiotherapy.....	31
2.2.7.3. Preoperative chemoradiotherapy.....	31
2.2.7.4. Time interval between radiotherapy and surgery....	32
2.2.7.5. Postoperative radiotherapy and chemoradiotherapy.	33
2.2.7.6. Novel agents in chemoradiation	33
2.2.7.6.1. Epidermal growth factor receptor inhibitors	33
2.2.7.6.2. Bevacizumab.....	34
2.2.7.7. Adjuvant chemotherapy.....	34
2.2.8. Multidisciplinary approach.....	35
2.2.9. Current treatment recommendations.....	35
2.2.10. Surveillance	35
2.2.10.1. Tumour markers.....	35
2.2.10.2. ASCO and ESMO recommendations for follow-up...	36
2.2.11. Prognosis.....	36
2.2.11.1. Adverse prognostic factors.....	36
2.2.11.2. Survival by stage.....	37
2.3. TUMOUR HYPOXIA	37
2.3.1. General aspects of hypoxia	37
2.3.2. Angiogenesis and hypoxia	38
2.3.3. Tumour vasculature, oxygenation and glycolysis	38
2.3.4. Measuring hypoxia	39
2.3.5. Activation of the hypoxia response element and HIF-1 α	39
2.3.6. Association of HIF-1 α with PI3-AKT and mTOR	40
2.3.7. Down-stream targets of HIF-1 α	41
2.3.7.1. Vascular endothelial growth factor (VEGF)	41
2.3.7.2. Carbonic anhydrase IX (CA IX)	42
2.3.7.3. GLUT-1.....	42
2.3.7.4. Ezrin.....	43
2.3.8. Effect of hypoxia on resistance to treatment.....	43
2.3.8.1. Radiotherapy and hypoxia	44
2.3.8.1.1. Oxygenation and reoxygenation	44

Table of Contents

2.3.8.1.2. Nitroimidazoles and anti-inflammatory drugs in radiosensitisation	45
2.3.8.1.3. Chemotherapeutic agents in radiosensitisation	45
2.3.8.1.4. Oxygen manipulation	46
2.3.8.3. Chemotherapy and hypoxia	46
2.3.8.4. Hypoxia, response to therapy and disease outcome	47
3. AIMS OF THE STUDY	49
4. PATIENTS AND METHODS	50
4. 1. PATIENTS	50
4. 2. METHODS	54
4.2.1. Immunohistochemical (IHC) stainings	54
4.2.1.1. IHC staining procedure	54
4.2.1.2. Evaluation of the IHC staining	54
4.2.1.2.1. CA IX (I)	54
4.2.1.2.2. HIF-1 α (II)	55
4.2.1.2.3. Ezrin (III)	55
4.2.1.2.4. GLUT-1 (IV)	55
4.2.1.2.5. HE-staining	56
4.2.2. Western Blot analysis (WB) (I, III)	56
4.2.2.1. Confirming CA IX expression in WB (I)	56
4.2.2.2. Ezrin in relation to radiation by WB (III)	56
4.2.3. Tumour regression grade evaluation (I-IV)	56
4.2.4. Statistical analysis (I-IV)	58
5. RESULTS	59
5.1. GENERAL ASPECTS OF IHC STAINING (I-IV)	59
5.2. CA IX (I, V)	59
5.3. HIF-1 α analysis (II)	63
5.4. EZRIN (III)	64
5.5. GLUT-1 (IV)	65
5.6. TUMOUR REGRESSION GRADE WITH REGARD TO IHC (I-IV)	66
5.8. COMBINED ANALYSIS OF HIF-1 α , CA IX, EZRIN AND GLUT-1	67
6. DISCUSSION	69
6.1. CA IX (I)	70
6.2. HIF-1 α (II)	72
6.3. EZRIN (III)	74
6.4. GLUT-1 (IV)	76
6.5. TUMOUR REGRESSION GRADE (I-IV)	77

Table of Contents

6.6. COMBINED ANALYSIS OF HIF-1 α , CA IX, EZRIN AND GLUT-1.....	78
7. SUMMARY AND CONCLUSIONS	79
8. ACKNOWLEDGEMENTS.....	80
9. REFERENCES	82
ORIGINAL PUBLICATIONS.....	103

ABBREVIATIONS

[18F]	fluorine-18
Ab	antibody
AIDS	acquired immunodeficiency syndrome
AJCC	American Joint Committee on Cancer
AKT	protein kinase of protein kinase B family
APC	adenomatous polyposis coli gene
APR	abdominoperineal resection
AR	anterior resection
ASCO	American Society for Clinical Oncology
ATP	adenosine triphosphate
BCL-2	B cell lymphoma-2 protein
BRAF	v-raf murine sarcoma viral oncogene homolog B1
CA IX	carbonic anhydrase IX
Ca ²⁺	calcium ion
CA19-9	membrane-associated glycoprotein
CAT	catalogue
CC	colon cancer
CEA	carcinoembryonic antigen
CI	confidence interval
CIN	chromosomal instability
Cl ⁻	chloride ion
COX-2	cyclo-oxygenase-2
CRC	colorectal cancer
CRM	circumferential margin
CT	computerised tomography
DFS	disease-free survival
DNA	deoxyribonucleic acid
DSS	disease- specific survival
E. coli	Escherichia coli
ECM	extracellular matrix
EGFR	epidermal growth factor receptor
ERM	ezrin-radixin-moesin
ESA	erythropoiesis-stimulating agent
ESMO	European Society for Medical Oncology
EUS	endorectal ultrasound
FAP	familial adenomatous polyposis
FOB	faecal occult blood
FULV	fluorouracil-leucovorin combination
G	differentiation grade

Abbreviations

GLUT-1	glucose transporter-1
Gy	Gray
HE	haematoxylin-eosin
HER2	human epidermal growth factor receptor 2
HIF-1	hypoxia-inducible factor-1
HIF-1 α	hypoxia-inducible factor -1 α
HIF-1 β	hypoxia-inducible factor -1 β
HIV	human immunodeficiency virus
hMLH-1	human mutL homolog 1
hMSH-2	human MutS homolog 2, colon cancer, non-polyposis type 1
HNPCC	hereditary non-polyposis colorectal cancer
HR	hazard ratio
HRE	hypoxia-response element
HRP	horseradish peroxidase
ICC	intra-class correlation coefficient
IgG	Immunoglobulin G
IHC	immunohistochemistry
K ⁺	potassium ion
kDa	kilodalton
KM	Kaplan-Meier method
KRAS	V-Ki-ras2 Kirsten rat sarcoma viral oncogene homolog
LDH	lactate dehydrogenase
LR	likelihood ratio
LVFU2	continuous infusion of fluorouracil combined to leucovorin
M	metastasis, metastatic
M/H	moderate/high
mAb	monoclonal antibody
MAP	MYH-associated polyposis
MAPK	mitogen-activated protein kinase
MLH1	mutL homolog 1 colon cancer, nonpolyposis type 2 (E. coli)
mM	millimoles per litre
mmHg	millimetre of mercury
MMR	mismatch repair
MRI	magnetic resonance imaging
MSH2	mutS homolog 2, colon cancer, nonpolyposis type 1 (E. coli)
MSH6	mutS homolog 6 (E. coli)
MSI	microsatellite instable/instability
MSI-H	high level of MSI
MSI-L	low level of MSI
MSS	microsatellite stable
mTOR	mammalian target of rapamycin
MYH	myosin, heavy chain 1, skeletal muscle, adult
N	lymph node
N/W	negative/weak

Abbreviations

Na ⁺	sodium ion
NPV	negative predictive value
NSAID	non-steroidal anti-inflammatory agent
O ₂	oxygen pressure
OR	odds ratio
p53	tumour protein 53
pCR	pathologic complete response
PDGF	platelet-derived growth factor
PET	positron emission tomography
PI3K	phosphoinositide-3-kinase
PMS2	post-meiotic segregation increased 2 (<i>Saccaromyces cerevisiae</i>)
pO ₂	partial pressure of oxygen
PPV	positive predictive value
pRb	retinoblastoma pathway
RAF	v-raf-1 murine leukaemia viral oncogene homolog
RAS	rat sarcoma
RC	rectal cancer
RNA	ribonucleic acid
RT	radiotherapy
SE	sensitivity
SOS	son of sevenless homolog
SP	specificity
Src	v-src sarcoma (Schmidt-Ruppin A-2) viral oncogene homolog (avian)
T	depth of tumour invasion
TCA	tricarboxylic acid cycle
TME	total mesorectal excision
TNM	tumour-node-metastasis classification system
TRG	tumour regression grade
TS	thymidylate synthase
U251	human glioma cell line
U2OS	human osteosarcoma cell line
UICC	Union for International Cancer Control
VEGF	vascular endothelial growth factor
VEGFR	vascular endothelial growth factor receptor
VHL	Von Hippel- Lindau protein
γH2A.X	phosphorylated form of histone H2A.X
κ	kappa

LIST OF ORIGINAL PUBLICATIONS

This thesis is based on the following original articles and unpublished data. The publications are referred to in the text by Roman numerals I-V:

- I Korkeila E, Talvinen K, Jaakkola PM, Minn H, Syrjänen K, Sundström J and Pyrhönen S. Expression of carbonic anhydrase IX suggests poor outcome in rectal cancer. *Br J Cancer* 2009; 100(6):874-80.
- II Korkeila E, Jaakkola PM, Syrjänen K, Sundström J and Pyrhönen S: Preoperative radiotherapy down-regulates the nuclear expression of hypoxia-inducible factor-1 α (HIF-1 α) in rectal cancer. *Scand J Gastroenterol*, 2010; 45(3): 340-348.
- III Korkeila E, Syrjänen K, Bendardaf R, Laulajainen M, Carpén O, Pyrhönen S and Sundström J: Preoperative radiotherapy modulates ezrin expression and its value as a predictive marker in patients with rectal cancer. *Hum Pathol* 2011; 42(3): 384-392.
- IV Korkeila E, Jaakkola PM, Syrjänen K, Pyrhönen S and Sundström J: Pronounced tumour regression after radiotherapy is associated with negative/weak glucose transporter-1 expression in rectal cancer. *Anticancer Res* 2011; 31 (1): 311–315.
- V Korkeila E, Talvinen K, Jaakkola PM, Minn H, Syrjänen K, Sundström J and Pyrhönen, S. Reply: Expression of carbonic anhydrase IX suggests poor response to therapy in rectal cancer. *Br J Cancer* 2009; 101 (2): 373–373 (*letter*).

The original publications have been reprinted with the permission of the copyright holders.

1. INTRODUCTION

Colorectal cancer (CRC) is a common malignancy with rising incidence in the western world. Annually about 1000 patients are diagnosed with rectal cancer (RC) in Finland. The most important prognostic factors of RC include the number of metastatic and examined lymph nodes, the depth of tumour invasion and the involvement of the circumferential margin (CRM) ^{1,2}.

Preoperative short-course radiotherapy (RT), utilised in the treatment of T3-tumours, has been shown to improve local disease control ^{3,4}. Long-course RT, preferably combined with fluorouracil-based chemotherapy, is used to treat locally advanced T4-tumours or tumours with threatened CRM involvement. Long-course RT is recommended preoperatively but it may be administered postoperatively, if the tumour has been under-staged prior to the operation or CRM is found to be involved ⁵.

Large tumours frequently have necrotic regions caused by lack of oxygen. However, tumours have hypoxic areas regardless of their size, stage, differentiation grade and the location ⁶. Hypoxia starts a cascade of events, including the activation of hypoxia-response element (HRE). The key protein in tissue response to oxygen deficiency is hypoxia-inducible factor-1 α (HIF-1 α). HIF-1 α consists of two protein subunits, HIF-1 α and HIF-1 β ⁷⁻⁹. HIF-1 α expression is oxygen dependent ⁷. In hypoxia, HIF-1 α stabilises, followed by dimerisation with the β -unit ⁸. HIF-1 α stabilisation leads to the activation of several genes involved in angiogenesis, glycolysis, invasion, metastasis, apoptosis, pH regulation and growth factor signalling ^{7,8,10-15}.

The downstream targets of HIF-1 α include glucose transporter-1 (GLUT-1) and carbonic anhydrase IX (CA IX) ^{7,10,13}. The mammalian target of rapamycin (mTOR) is a key regulator of HIF-1 α protein synthesis in many cancers ¹⁶. Rapamycin, a protein kinase that inhibits mTOR activity, is shown to act as an up-stream regulator of HIF-1 α in hypoxia ¹⁷. Ezrin belongs to the ezrin-radixin-moesin (ERM) family of proteins ^{18,19}, which function, besides of cross-linking the plasma membrane and the cytoskeleton ²⁰⁻²³, also in signal transduction ²⁴⁻²⁶. Ezrin expression is necessary for metastatic behaviour of cancer cells ²⁷ and ezrin-related metastasis is shown to be linked to mTOR pathway ²⁸. Blocking this pathway with rapamycin is found to inhibit experimental lung metastasis in vivo ²⁸. Thus, the regulation of mTOR and ezrin as well as mTOR, ezrin and HIF-1 α are interrelated.

Hypoxia is associated with resistance to treatment and unfavourable disease outcome in several solid tumours²⁹⁻³². Preoperative radiotherapy is shown to improve local disease outcome and to be better tolerated than postoperative RT^{3, 33}. Chemotherapy is usually combined with long-course RT in the treatment of locally advanced tumours, aiming at enhancing response and diminishing radioresistance^{30, 34, 35}.

2. REVIEW OF THE LITERATURE

2.1. GENERAL ASPECTS OF COLORECTAL CANCER

2.1.1. Epidemiology of CRC

Colorectal cancer (CRC) is a common malignancy in the western world. Annually there are over one million new cases of CRC worldwide, accounting for 9% of all cancers³⁶. In Europe, the United States, and elsewhere in the western world the incidence rates are higher than in other parts of the world, corresponding to 10-13% of all cancers^{37, 38}. In Finland, CRC is the third most common cancer. More than 1500 new cases of colon cancer and about 1000 new cases of RC are diagnosed annually³⁹. Increased survival into advanced age is mainly responsible for the growing incidence of CRC⁴⁰. CRC is rare at young age, albeit patients with genetic alterations are at risk for CRC at a younger age⁴¹. Only about 2% of patients are younger than 40 years, whereas two thirds of patients are at or above age 65 at diagnosis³⁹.

2.1.2. Aetiology of CRC

2.1.2.1. Sporadic CRC

2.1.2.1.1. Carcinogenesis

Carcinogenesis involves growth signalling between cancer cells and non-malignant stromal cells⁴², as well as activation of several pathways, e.g. SOS-RAS-RAF-MAPK⁴³ and disruption of others, e.g. pRb pathway⁴⁴. Tumours also have the ability to generate growth factors⁴². Over-expression of receptors, e.g. epidermal growth factor receptor (EGFR) enables a tumour to hyper-response to growth signals from its environment⁴². Extracellular receptors, integrins, link the cancer cell to the extracellular matrix (ECM)⁴⁵. The integrins and ligand-activated growth receptors have the ability to activate the SOS-RAS-RAF-MAPK cascade⁴⁵. RAS can mediate mitogenic signals without normal up-stream activation and it is shown to be mutated in many cancers, including CRC⁴³. The signalling pathways in cancer are complex, with cross-talking linkages to other pathways⁴². The hallmarks of cancer also include sustained angiogenesis, the ability to resist or inhibit programmed cell death, apoptosis, and the ability to invade and metastasise, features that

separate benign tumours and cancer from each other⁴². Malignant transformation can proceed through the adenoma-carcinoma sequence or the serrated pathway.

2.1.2.1.2. Adenoma-carcinoma sequence

CRC developing through the adenoma-carcinoma sequence^{46, 47} involves a series of events in which a benign adenoma transforms into a malignant tumour via intermediate steps of premalignant states⁴². This multistep process requires oncogene activation and loss of tumour suppressor genes^{46, 48}.

2.1.2.1.3. Serrated adenoma pathway

Serrated colorectal carcinomas are considered to develop from hyperplastic polyps and serrated adenomas through the serrated neoplasia pathway^{49, 50}. Oncogenic BRAF mutations are characteristic to this pathway⁵⁰. Serrated cancers represent microsatellite stable (MSS) or microsatellite instable-low (MSI-L) phenotype^{49, 51}. In IHC analysis, they are more frequently HIF-1 α positive than conventional CRCs⁴⁹. Serrated carcinomas account for about 10-15% of all CRCs⁵⁰ and have a predilection to the caecum and rectum⁵¹.

2.1.2.1.4. Genomic instability in sporadic CRC

CRC carcinogenesis involves two forms of genomic instability: microsatellite instability (MSI) and chromosomal instability (CIN)⁴². The inefficiency of DNA mismatch repair system (MMR) to fix errors in DNA replication probably results to changes in the number of repeats during DNA replication, which leads to the instability of microsatellite loci⁵². High level of MSI (MSI-H) has been identified in about 10% of sporadic CRC⁴⁸. CIN or MSI was found in about 60% of tumours in a study comprising tumour samples from patients with sporadic CRC⁴⁸. A Finnish study of patients with advanced CRC, investigating thymidylate synthase (TS) and two MMR proteins, hMLH1 and hMSH2, showed that patients with low TS and deficient MMR had significantly shorter DFS than patients with high expression of these markers⁵².

2.1.2.2. Familial and hereditary CRC

CRC is most commonly sporadic but about 10-15% of cases are familial⁵³. The hereditary conditions include hereditary non-polyposis colorectal cancer (HNPCC or Lynch disease) and familial adenomatous polyposis (FAP), which are inherited in autosomal dominant manner⁵⁴. People, who have a first-degree relative with CRC, are at two-fold risk of getting the disease⁵⁵. The

risk is more pronounced, if the relative is diagnosed at an early age and if other first-degree relatives are also affected⁵⁵.

FAP accounts for less than 1 % of all CRCs⁴⁰. FAP patients have multiple adenomas and an associated high risk of malignancy at young age⁵⁴ linked to a mutation in APC- tumour suppressor gene⁵⁶. HNPCC accounts for about 2-4% of CRC⁵³. The syndrome has a high penetrance and involves the development of CRC at an early age⁵⁷. HNPCC is associated with an increased lifetime risk of 80% of predominantly right-sided CRC and an increased lifetime risk of 30-60% of endometrial cancer^{54, 58, 59}. Mutation carriers also have an increased risk for small bowel, urothelial and ovarian cancers as well as brain tumours²⁶. The genetic background of HNPCC comprises germ line mutations in the MMR genes (MLH1, MSH2, MSH6 and PMS2)^{56, 58, 60}.

The distinction of sporadic CRC from familial or hereditary cancers is not clear-cut but rather sliding⁵⁶, presumably with less influence of lifestyle, nutritional and environmental factors on hereditary than sporadic forms of cancer⁵⁶. Several workshops have formulated criteria for identifying people at the highest risk of CRC and extending screening methods, genetic testing and counselling at their disposal. These clinical diagnostic tools include the Amsterdam criteria⁶¹ to reveal possible HNPCC (**Table I**) and the Bethesda Guidelines to find families with indications for genetic (MMR) testing⁵⁸.

2.1.2.3. Predisposing conditions

Patients with ulcerative colitis and Crohn's disease^{60, 62}, polyps, previous malignancy of the large bowel or radiotherapy to a pelvic malignancy are at increased risk of CRC^{46, 63}.

2.1.2.4. Nutritional and lifestyle factors

High incidence of CRC is possibly modified by western sedentary lifestyle, including obesity, diabetes, and dietary habits⁴⁰. However, the influence of these factors on the risk of RC is somewhat different from that of colon cancer (CC). Body mass index is not associated with an increased risk of RC⁶⁴. Active physical exercise may modestly decrease the risk of CC but not RC⁶⁵. Red and processed meat and alcohol consumption is associated with an increased risk of RC^{66, 67}, whereas fish intake may have a protective effect⁶⁶. No clear association of dietary antioxidant vitamins and carotenoids with CRC risk has been found⁶⁸. Short duration of sleep is shown to increase the risk of colorectal adenoma, and hence, colorectal neoplasia⁶⁹.

Table I. Revised ICG-HNPCC Criteria (Amsterdam Criteria II)*

There should be at least 3 relatives with an HNPCC-associated cancer (CRC, cancer of the endometrium, small bowel, ureter, or renal pelvis)
One should be a first-degree relative of the other 2
At least 2 successive generations should be affected
At least 1 person should be diagnosed before the age of 50
Familial adenomatous polyposis should be excluded in the CRC case(s) if any
Tumours should be verified by pathological examination

**Reprinted from Gastroenterology, Vol 116, Vasen H, Watson P, Mecklin JP, Lynch H and the ICG-HNPCC: New Clinical Criteria for Hereditary Nonpolyposis Colorectal Cancer (HNPCC, Lynch syndrome) Proposed by the International Collaborative Group on HNPCC, pages 1453-1456, 1999, © 2010 with permission from Elsevier.*

2.1.2.5. Other risk factors

Use of acetylsalicylic acid compounds and oral contraceptives reduce the risk of CRC^{70, 71}. Chemically induced RC is shown to be inhibited in male rats by giving exogenous testosterone during carcinogen administration, implicating that hormonal manipulation may have an effect on RC risk in men⁷². Currently there is no clear evidence of an association of CRC with viral infection, but AIDS patients have a 3-4 fold increased risk of CRC, possibly caused by an oncogenic protein related to HIV infection⁷³. Human papilloma virus may be associated with the risk of CRC⁷⁴. Interestingly, observations in animal studies have shown that colonisation of the large intestine with certain E. coli strains induces DNA damage in vivo, implying that these E. coli strains could be involved in sporadic colorectal carcinogenesis⁷⁵.

2.2. RECTAL CANCER

2.2.1. Signs and symptoms of disease

Symptoms of RC include a change in the bowel habits or the consistency of the stools, faecal blood or dark stools, narrow thread-like or mucous faeces, abdominal or pelvic discomfort or bloating, urgency, incompleteness of emptying the bowel, obstructive symptoms or increased flatulence. General

symptoms like tiredness, pain, nausea, vomiting, weight loss or anaemia may be present. However, the disease may give only obscure or slight symptoms.

2.2.2. Diagnostic and screening procedures

The diagnosis of suspected RC is based on digital rectal examination, colonoscopy and histological biopsies. A magnetic resonance imaging (MRI) is performed to define the local extent and resectability of the disease and a computerised tomography (CT) to rule out distant metastasis. CRCs developing from benign adenomas through the adenoma-carcinoma sequence^{46, 47} offer an opportunity for screening. Faecal occult blood test (FOB) is a generally used method^{76, 77}. In the screening programmes, a 16% reduction of mortality from colorectal cancer has been shown⁷⁸. In Finland, a gradually expanding screening programme was started in 2004, including individuals 60-69 years old randomised into screening and control arms. FOB is tested twice a year and in case of a positive faecal occult blood test, a colonoscopy is performed⁷⁹. In this screening programme after the first screening interval, an invasive CRC was reported to be diagnosed in 8.2% of the FOB positive population⁸⁰. However, among the screened FOB negative patients, the proportion of interval cancers was found to be 27.3%⁸⁰.

2.2.3. Pathology and classification

2.2.3.1. Histopathology

Tumours up to 15 cm from the anal verge are considered rectal carcinomas⁵. The majority of them (85-90%) are classified as conventional adenocarcinomas⁸¹. About 8-10% of tumours are mucinous cancers, defined by the composition of $\geq 50\%$ of extracellular mucin⁸². CRC associated with serrated adenoma tends to produce mucin and harbour KRAS and BRAF mutations^{49, 50}. Other types of tumours can be found in the rectum albeit rarely, including angiosarcomas⁸³, melanomas⁸⁴, squamous cell carcinomas⁸⁵, as well as neuroendocrine carcinomas, the incidence of which is rising⁸⁶.

2.2.3.2. Handling and evaluation of the specimen

The specimen is examined macroscopically by the surgeon and the pathologist for the completeness of the dissected mesorectum. The resection margins are inked and the specimen is fixed in formalin and thinly sliced. The pathologist measures the narrowest proximal, distal and circumferential margins. The

mesorectal fat is sliced and carefully examined for lymph nodes, using fat clearance techniques as needed to detect a sufficient amount of lymph nodes⁸⁷.

2.2.3.3. Circumferential margin

The resected tissue is macroscopically examined to assure that the mesorectum is excised completely⁸⁸. Proximal and distal margins are also measured⁸⁸. Moreover, a clear CRM is critical to achieve a curative resection. The radial resection margin encompasses the vessels, lymph nodes and lymphatics of the rectum, surrounded by fascia. CRM less than 1 mm is associated with a higher local relapse rate⁴, and an increased risk is reported to be present with CRM less than 2 mm⁸⁹. Also, tumour location closer than 5 cm from the anal verge increases the probability of positive CRM⁸⁹. Abdominoperineal resection is associated with a higher rate of intraoperative perforation⁹⁰, positive CRM, higher local recurrence rate of 22%, and more unfavourable prognosis than anterior resection⁹¹.

2.2.3.4. Lymph node evaluation

The number of examined lymph nodes strongly affects the patient's prognosis¹. According to the ESMO clinical recommendations, a minimum of twelve lymph nodes is to be examined for adequate staging⁵. However, after long-course preoperative treatment it is not always possible to collect the required number of lymph nodes⁹²⁻⁹⁴. In general, the more lymph nodes are examined, the more adequate is the staging^{1,95}. The impact of the number of examined lymph nodes was studied in T3N0 rectal cancer¹. When 8-12 lymph nodes were examined, a patient's hazard ratio (HR) for death was reduced to 0.81 and if at least 13 lymph nodes are examined, it dropped to 0.68, as compared to patients with 1-7 lymph nodes examined¹.

2.2.3.5. Tumour differentiation grade

Tumour differentiation grade (G) (**Table II**) is assessed from the diagnostic biopsies and the operative samples. However, the size of biopsy may be insufficient to reliably determine the differentiation grade. Also, preoperative RT may affect tumour differentiation grade and after a complete response to preoperative treatment it is not feasible to assess.

2.2.3.6. Perineural and vascular invasion

The interactions of cancer cells and their surrounding microenvironment play an important role in cancer growth and progression as well as in tumour response to treatment⁹⁶. Vascular invasion is shown to predict cancer-specific mortality and local recurrence in RC⁹⁷. Perineural tumour invasion also affects the patient's prognosis, and may help to identify patients with node-negative tumours in need of adjuvant therapy⁹⁸.

Table II. WHO tumour classification

Criterion	Differentiation category	Numerical grade ^a	Descriptive grade
> 95 % with gland formation	Well-differentiated	1	Low
50-95 % with gland formation	Moderately differentiated	2	Low
>0-49 % with gland formation	Poorly differentiated	3	High
High level of microsatellite instability ^b	Variable	Variable	Low
^a The category "undifferentiated carcinoma" (grade 4) is reserved for carcinomas with no gland formation, mucin production or endocrine, squamous or sarcomatoid differentiation; ^b MSI-H			

MSI-H microsatellite instability-high

Reprinted from Hamilton SR, Bosman FT, Boffetta P, Ilyas M, Morreau H, Nakamura SI, Quirke P, Riboli E and Sobin LH: Carcinoma of the colon and rectum in Bosman FT, Carneiro F, Hruban RH and Theise ND (editors). WHO Classification of Tumours of the Digestive System, 4th edition. International Agency for Research on Cancer, Lyon, 2010, page 138, © 2010 with permission from the World Health Organization.

2.2.4. Tumour response to preoperative treatment

Preoperative treatment provokes morphological changes in both cancer tissue and the surrounding stromal cells. These changes can be identified by the appearance of tumour fibrosis and inflammatory cells, increased mucin production and mucin pools, accompanied by eosinophilia, nuclear pyknosis and necrosis as well as cytoplasmic vacuolisation of cancer cells^{99, 100}. Damaged cells do not die immediately after radiation, but divide 1-2 times before their mitotic death, occurring by necrosis or apoptosis¹⁰¹.

2.2.4.1. Methods for predicting response to treatment

Several methods have been used to predict response to radiotherapy, including approaches for detecting DNA variability (e.g. DNA sequencing), RNA expression measurements (e.g. microarray) as well as methods for studying genetic alterations (e.g. gene deletions, amplifications) or protein expression (e.g. IHC, Western blot)¹⁰². In rectal cancer, high TS expression and persistent positive lymph nodes after preoperative chemoradiotherapy have been found to be associated with unfavourable prognosis¹⁰³. Tumour suppressor gene p53 mutations have been detected by complete direct sequencing of exons 2 to 10 and compared with p53 IHC from the preoperative biopsies, showing p53 genotype but not p53 IHC to predict response to preoperative treatment and survival¹⁰⁴. Furthermore, CEA level falling below 5ng/ml after preoperative chemoradiotherapy has been found to be coupled with complete clinical and pathologic response as well as survival¹⁰⁵.

2.2.4.2. Tumour regression grade

Tumour regression grade (TRG) is used to estimate the response to given preoperative treatment. TRG is analysed in HE-stained sections of the tumour after long-course RT. This assessment is generally based on a scale by Mandard and colleagues, further modified by Dworak, Rödel and colleagues^{99, 106, 107}. The response of the tumour is assessed using three to five main categories, depending on the scale used^{99, 106, 107}, with the response varying from complete to no regression. Pathologic complete disappearance of the tumour is achieved in 0-29%^{100, 107-112} of tumours. Good tumour regression is shown to associate with favourable disease outcome^{107, 108, 112}.

2.2.4.3. Imaging in predicting response to treatment

Imaging of hypoxia could give additional prognostic information and help in estimating the response to radiation and the effect of novel therapies¹¹³. The resolution of the current routine imaging methods is not sufficient to measure biological response in tumours¹¹⁴. Vital cancer cells may persist after clinical and pathological complete response¹¹⁴. Thus, additional methods could be helpful in defining the amount of truly vital cancer cells. [18 F]Fluorodeoxyglucose positron emission tomography (FDG-PET) has been studied to evaluate radiation response in solid tumours, including rectal cancer. A favourable disease outcome has been observed with negative FDG-PET after preoperative chemo-RT in locally advanced rectal cancer¹¹⁵. MRI imaging can detect the CRM, reduction in tumour size and nodal down-staging

in rectal cancer. However, radiation-associated fibrosis may be one confounding factor in imaging these post-treatment tumour parameters¹¹⁶. The ability of MRI to detect pCR in rectal cancer after chemoradiation has been evaluated in a prospective study, showing MRI to underestimate tumour response to therapy in about two thirds of patients¹¹⁷.

2.2.5. Staging

RC staging is based on the assessment of the depth of tumour invasion through the basal layer and the bowel wall (T), the regional lymph node involvement (N) and the presence or absence of distant metastases (M)¹¹⁸ as depicted in **Tables III-V**.

2.2.5.1. Preoperative staging

Preoperative staging helps to define the patient's risk of disease recurrence as well as the need and type of preoperative treatment. To assess the local extent of the disease, a digital rectal examination and an endorectal ultrasound (EUS), a magnetic resonance imaging (MRI) (**Figure 1a and b**), or a computerised tomography (CT) scan of the rectum are performed. MRI interpretation tends to over-stage T1 and T2 lesions⁸⁷, while EUS is the most accurate method to define the local invasion of a T1-T2 tumour¹¹⁹. However, in nodal staging, EUS is not markedly superior to MRI and CT⁸⁷. Furthermore, the size and shape of lymph nodes is not a reliable method to rule out metastases in rectal cancer, since even small nodes of 2-3 mm can be metastatic^{87,120}.

To rule out distant metastases, a CT of the chest and abdomen and chest x-ray are performed. A positron-emission tomography (PET) scan may be considered in selected cases. A complete colonoscopy is needed pre- or postoperatively to detect possible synchronous tumours in the colon⁵.

2.2.5.2. Pathology report and postoperative staging

In many centres, a structured pathology report is used to clarify the assessment of the pathological stage. A structured pathology report includes information on the proximal, distal and circumferential margins of the tumour, the depth of invasion, the number of metastatic and examined lymph nodes, tumour differentiation grade and the presence or absence of extramural venous invasion⁵ (**Table VI**). The complete report also comprises information

of perineural and vascular invasion, and the presence or absence of vital cancer cells. A tumour regression grade may also be assessed. Thus, the pathologist estimates the extent of the disease, the accuracy of preoperative imaging, the quality of surgery and the effect of preoperative therapies⁸⁷.

Postoperative staging is based on the pathological findings of the tumour specimens, using the International Union Against Cancer (UICC)¹¹⁸ (**Tables III-V**) or American Joint Committee on Cancer (AJCC)¹²¹ tumour classification tables. In case of complete tumour regression, postoperative tumour grade is not assessable.

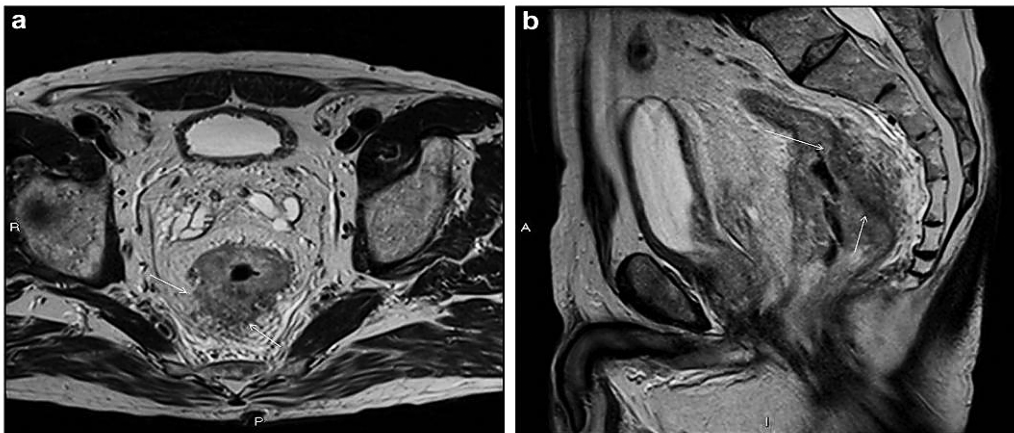


Figure 1a and b. Axial and sagittal MRI scan of advanced rectal cancer (arrows) (with patient's permission).

Table III. TNM Clinical Classification*

T-Primary Tumour	N-Regional Lymph Nodes
Tx Primary tumour cannot be assessed	Nx Regional lymph nodes cannot be assessed
T0 No evidence of primary tumour	N0 No regional lymph node metastasis
Tis ¹ Carcinoma in situ: intraepithelial or invasion of lamina propria	N1 Metastasis in 1-3 regional lymph nodes
T1 Tumour invades submucosa	N1a Metastasis in 1 regional lymph node
T2 Tumour invades muscularis propria	N1b Metastasis in 2-3 regional lymph nodes
T3 Tumour invades subserosa or into the non-peritonealized pericolic or perirectal tissues	N1c Tumour deposit(s), i.e. satellites*, in the subserosa, or in non-peritonealized pericolic or perirectal soft tissue <i>without</i> regional lymph node metastasis
T4 Tumour directly invades other organs or structures and/or perforates visceral peritoneum	N2 Metastasis in 4 or more regional lymph nodes
T4a Tumour perforates visceral peritoneum	N2a Metastasis in 4-6 regional lymph nodes
T4b Tumour directly invades other organs or structures ^{2,3}	N2b Metastasis in 7 or more regional lymph nodes
Notes:	Note:
1. Tis includes cancer cells confined within the glandular basement membrane (intraepithelial) or mucosal lamina propria (intramucosal) with no extension through the muscularis mucosae into the submucosa	*Tumour deposits (satellites), i.e. macroscopic or microscopic nests or nodules, in the pericorectal adipose tissue's lymph drainage area of a primary carcinoma without histological evidence of residual lymph node in the nodule, may represent discontinuous spread, venous invasion with extracellular spread (V1/2) or a totally displaced lymph node (N1/2). If such deposits are observed with lesions that would otherwise be classified as T1 or T2, then the T classification is not changed, but the nodule(s) is recorded as N1c. If a nodule is considered by the pathologist to be totally replaced lymph node (generally having a smooth contour), it should be recorded as a positive lymph node and not as a satellite, and each nodule should be counted separately as a lymph node in the final pN determination.
2. Direct invasion in T4b includes invasion of other organs or segments of the colorectum by way of the serosa, as confirmed on microscopic examination, or for tumours in a retroperitoneal or subperitoneal location, direct invasion of other organs or structures by virtue of extension beyond the muscularis propria.	
3. Tumour that is adherent to other organs or structures, macroscopically, is classified cT4b. However, if no tumour is present in the adhesion, microscopically, the classification should be pT1-3, depending on the anatomical depth of wall invasion.	

* According to *TNM Classification of Malignant Tumours, International Union Against Cancer, 7th Edition, Wiley-Blackwell, Singapore, 2010. Reprinted with permission © 2010 John Wiley and sons Ltd. All rights reserved.*

Table IV. Stage grouping of rectal cancer*

Stage 0	Tis	No	M0
Stage I	T1,T2	N0	M0
Stage II	T3,T4	N0	M0
Stage IIA	T3	N0	M0
Stage IIB	T4a	N0	M0
Stage IIC	T4b	N0	M0
Stage III	Any T	N1,N2	M0
Stage IIIA	T1,T2	N1	M0
Stage IIIB	T1	N2a	M0
	T3,T4a	N1	M0
	T2,T3	N2a	M0
Stage IIIC	T1,T2	N2b	M0
	T4a	N2a	M0
	T3,T4a	N2b	M0
Stage IVA	T4b	N1,N2	M0
	Any T	Any N	M1a
Stage IVB	Any T	Any N	M1b

* M0=no distant metastases, M1 distant metastases present. According to *TNM Classification of Malignant Tumours, International Union Against Cancer, 7th Edition, Wiley-Blackwell, Singapore, 2010*. Reprinted with permission © 2010 John Wiley and sons Ltd. All rights reserved.

Table V. TNM classification of distant metastasis (M)*

M0	No distant metastasis
M1	Distant metastasis
	M1a Metastasis confined to one organ (liver, lung, ovary, non-regional (lymph node (s)))
	M1b Metastasis in more than one organ or the peritoneum

*According to *TNM Classification of Malignant Tumours, International Union Against Cancer, 7th Edition, Wiley-Blackwell, Singapore, 2010*. Reprinted with permission © 2010 John Wiley and sons Ltd. All rights reserved.

Table VI. The structured attachment of pathology report used in Turku University Hospital

Tumour diameter	(mm)
Distance from visceral peritoneum	(mm)
Lateral/circumferential margin	(mm)
Proximal margin	(cm)
Distal margin	(cm)
Histological grade	I/II/III
Depth of penetration	pT1 invasion of submucosa pT2 invasion of muscularis propria pT3 through muscularis propria into subserosa pT4 invasion of visceral peritoneum/lateral margin
Vascular/lymphatic invasion	yes/no
Metastatic/Examined lymph nodes	(number M/E)
Tumour regression grade	1)<25% 2) 2-50% 3)>50% 4) 100%

2.2.6. Surgical treatment of localised rectal cancer

2.2.6.1. Anterior resection and abdominoperineal resection

The goal of RC surgery is to achieve a macroscopically as well as microscopically radical resection (R0) with negative distal and circumferential margins¹²². Two primary modes of operation, anterior resection (AR) and abdominoperineal resection (APR) are used. AR preserves the continuity of the intestine, while APR comprises the pelvic and the perineal incisions and necessitates a colostomy. Low AR is usually feasible for tumours of the upper and mid rectum¹²². It may also be possible when operating distal carcinomas, unless the sphincter mechanism is involved¹²². Usually a protecting temporary stoma, however, is required.

Sufficient proximal, distal and CRM margins are prerequisites for achieving local disease control. For tumours of the upper and mid rectum, a macroscopic distal margin of 4-5 cm is preferred, while 1-2 cm margin may be acceptable for low-situating cancers^{87, 122, 123}.

2.2.6.2. Total mesorectal excision technique

The surgical technique of choice is total mesorectal excision (TME), which involves the removal of the mesorectum intact, including the lymphovascular tissue¹²⁴, in order to obtain a sufficient CRM and improved local control⁴. The local recurrence rate varies from 12-15% to 30-40% after conventional surgery¹²⁴⁻¹²⁶, while it is 5-15 % after TME^{124, 125}.

2.2.6.3. Laparoscopic surgery

The interest in less invasive operative procedures is increasing due to anticipated earlier recovery and better physical functioning^{127, 128}. Early reports from ongoing trials have not shown increased local recurrence rates or worse disease outcome^{129, 130}.

2.2.6.4. Local excision

Local excision of RC can be considered for patients with superficial T1-adenocarcinomas with no adverse histologic or clinical features^{1, 2, 4, 97, 98, 103, 107, 108, 131-142} (**Table VII**). Surgery is generally performed using transanal excision involving the whole rectal wall to the perirectal fat with at least a 1 cm lateral margin¹²². Local excision is also an option for those patients, who have

severe comorbidities or metastatic disease or who refuse APR¹⁴³. However, even selected elderly patients tolerate and benefit from major curative surgery¹⁴⁴.

Table VII. Adverse histopathologic and clinical features in rectal cancer

Advanced postoperative stage (TNM)	Dukes CE, 1958 Kuo LJ et al, 2007 Liersch et al, 2007 Peeters K et al, 2007
Poorly differentiated histology	McDermott et al, 1984
Presence of lymphovascular invasion	Compton et al, 2000 Tilney et al, 2009
Tumour level (low)	Peeters K et al, 2007
Presence of perineural invasion	Liebig et al, 2009
Examination of less than 12 lymph nodes	Tepper JE et al, 2001 Kapiteijn et al, 2001 Swanson et al, 2003 Edler D et al, 2007
Involved circumferential margin	Adam et al, 1994 Hall et al 1998 Nagtegaal 2008
Poor response to preoperative treatment	Ruo et al, 2002 Rödel et al, 2005
Tumour involving the surface of the specimen	Compton et al, 2000
Clinical obstruction or perforation	Griffin et al, 1987
Intraoperative tumour perforation	Zirngibl H et al, 1990
High level of CEA	Compton et al, 2000

2.2.7. Oncological treatments of rectal cancer

2.2.7.1. General aspects of radiotherapy

Local recurrences were previously a major problem after RC, deteriorating the patients' quality and quantity of life. RT is used to improve local control and/or operability and to allow sphincter preservation¹⁴⁵. RT is given before or after surgery, with or without chemotherapy. Several advantages are associated with preoperative radiotherapy, including patient compliance and treatment tolerability, tumour down-staging, operability and possible sphincter preservation. Possible disadvantages include side-effects of treatment and over-treatment due to the limitations of imaging in estimating tumour stage³³. CT-based dose planning is used, encompassing 3-4 radiotherapy fields and 6-24 megavoltage photons with daily fractions of 1.8 Gy to a total dose of 50.4 Gy in 6 weeks (long-course RT) or 5 fractions of 5Gy in a week (short-course RT)¹⁴⁶. The RT portals cover the primary tumour and the adjacent lymph nodes, reaching from the border of the first sacral and the fifth lumbar vertebra down to the obturator foramina, anteriorly from the hip joints to the sacral bone³. Radiotherapy plan of a rectal cancer patient is depicted in **Figures 2a and b**.

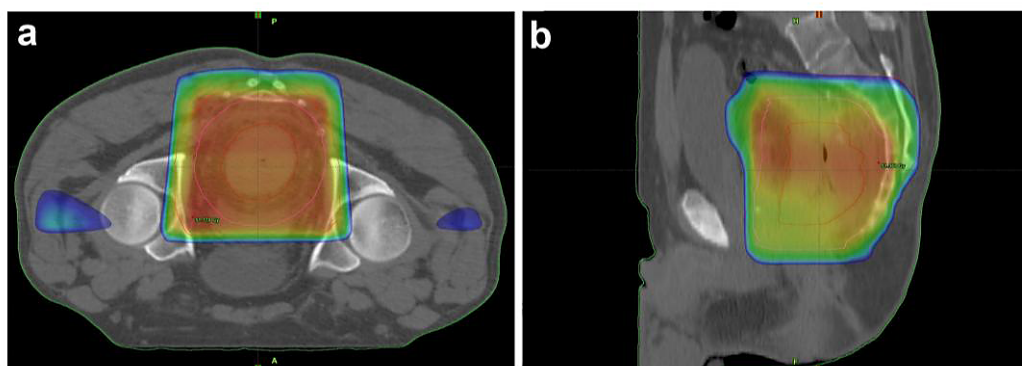


Figure 2 a-b. Radiotherapy plan in the treatment of rectal cancer (patient in a prone position): a, axial b, sagittal scans (with patient's permission).

Patients with T3 tumours and tumours with suspected metastatic lymph nodes are usually treated with short-course preoperative RT, whereas T4, very large or fixed tumours or tumours with predicted CRM involvement are generally treated with long-course RT^{5, 147}. Patients with T1-T2 tumours may be treated without preoperative therapy¹⁴⁷. There is still controversy about the treatment and clinical behaviour of proximal or upper rectal tumours¹⁴⁸. A

British study showed that patients with early disease (T2N0) and tumours in the upper rectum were less likely offered neoadjuvant therapy by multidisciplinary team¹⁴⁹. Subgroups of patients with high risk upper RC may be identified by the depth of tumour infiltration, lymph node status and tumour differentiation grade¹³¹.

Precise RT planning diminishes the risk of possible side effects, which include diarrhoea, frequent defecation, incontinence, urgency and soiling, sexual dysfunction, and bladder irritation³. Interestingly, the risk of prostate cancer, after RT for rectal cancer, is substantially reduced¹⁵⁰.

2.2.7.2. Preoperative short-course radiotherapy

The Swedish Rectal Cancer Group showed that preoperative short-course RT improved local disease control and survival. Local relapse rate decreased from 27% to 11% and five-year overall survival increased from 48% to 58%³. The rate of side effects and surgical complications did not increase significantly after preoperative RT³. The results were reported to persist after median follow-up time of 13 years¹⁵¹. The Dutch Colorectal Cancer Group confirmed that preoperative RT further reduced the risk of local recurrence significantly from 8.2% to 2.4% even after TME- technique was utilised in surgery⁴. After median follow-up of 6 years, the difference persisted between the two groups but no survival advantage was seen¹⁴¹. Patients with positive nodes, tumours of the mid rectum or tumours with negative CRMs benefited most from preoperative RT¹⁴¹. The relative risk reduction of local recurrences in the preoperative radiotherapy group was 71 % at 2 years and 49% at 5 years as compared with the surgery group^{4, 141}.

2.2.7.3. Preoperative chemoradiotherapy

RT is generally combined with fluorouracil-based chemotherapy (fluorouracil or capecitabine), if not contraindicated, based on the results of two EORTC trials^{152, 153}. Fluorouracil (FU) is a fluoropyrimidine analogue, given either as a bolus treatment in 4-5 day-cycles (Mayo regimen)¹⁵⁴ or via continuous intravenous infusion¹¹⁰. Capecitabine is an oral fluoropyrimidine precursor that is activated by a three-step enzymatic cascade, including conversion to active FU by thymidine phosphorylase. Thymidine phosphorylase is more active in the tumour than in normal tissues, resulting in about three-fold concentration of FU in the tumour, as compared to normal tissue¹⁵⁵. Capecitabine is given concomitantly and continuously with RT¹⁵⁶.

Preoperative chemoradiotherapy is utilised in the treatment of locally advanced rectal cancer, showing infiltration to the adjacent structures or threatened CRM. After preoperative treatment, an inoperable carcinoma may turn operable and even pCRs are seen¹⁰⁸. The EORTC-trial reported chemoradiotherapy to improve local disease control, irrespective of timing¹⁵². Sauer and colleagues compared preoperative with postoperative chemoradiotherapy in the treatment of stage II and III RCs in a randomised study³³. Preoperative chemoradiotherapy was shown to improve local control but not overall survival of patients with T3-T4 tumours, as compared to postoperative combined treatment³³. This trial demonstrated a lower local relapse rate and less acute toxicity in the preoperative as compared with the postoperative treatment arm³³. The Polish study randomised patients to receive either short-course radiotherapy or long-course chemoradiotherapy preoperatively. In contrary to the EORTC trial, no increase in survival, local control or late toxicity was found after chemoradiotherapy, as compared with short-course radiotherapy, albeit a down-staging effect was seen¹⁵⁷.

In the study by Guillem and colleagues, excellent response after preoperative treatment was shown to predict improved disease outcome¹⁵⁸, which may reflect tumour biology sensitive to therapeutic interventions¹⁰⁹. Since most studies have not shown survival advantage after preoperative treatment, despite improved local disease outcome, chemotherapy intensification could be feasible. In fact, in locally advanced RC, oxaliplatin in combination with capecitabine in the preoperative setting was shown to be efficient and result in excellent tumour regression in a phase II study¹⁵⁹. The Dutch Colorectal Cancer Group reported a high R0-resection rate of 81% in a phase I-II study of locally advanced rectal cancer, treated by capecitabine and oxaliplatin combined to preoperative radiotherapy¹⁶⁰.

Sphincter-preservation has been one of the focuses in several studies investigating the effects of preoperative chemoradiotherapy. Randomised trials have so far not shown an increased rate of sphincter-saving surgery after CRT¹⁶¹.

2.2.7.4. Time interval between radiotherapy and surgery

Timing of surgery after preoperative treatment is critical, when considering down-staging of the tumour or possible sphincter-saving procedures. The effects of radiotherapy on cancer tissue are not seen immediately¹⁶² but cancer cells may appear morphologically intact shortly after radiation¹⁶³. The

Lyon trial randomised patients in two groups: short (2 weeks) or long (4 weeks) interval between the end of RT and the subsequent operation. This trial showed that a long interval resulted in increased down-staging and complete pathologic remission (pCR) rate, as compared with a short interval¹⁶². The Dutch Colorectal Cancer Group study found no tumour down-staging, if the interval between RT and surgery was less than ten days⁹². A waiting time of at least 8 weeks was found to be the only factor predicting pCR in a non-randomised study by Kalady and colleagues¹⁶⁴. In their study, pCR was also associated with improved local control and survival¹⁶⁴. Even longer intervals of 12 weeks have been reported¹⁶⁵.

Currently, short-course RT is given over five days, with surgery on the following week. The time interval between long-course RT and surgery is about 6-8 weeks. Recently, delayed surgery up to 6-8 weeks after short-course preoperative RT was also shown to result in substantial tumour regression¹⁶⁶. A randomised trial is under way to evaluate this treatment strategy in resectable rectal cancer¹⁶⁶.

2.2.7.5. Postoperative radiotherapy and chemoradiotherapy

Postoperative RT delivered 30-60 days after surgery to a total dose of 40 Gy, was found to reduce and postpone local relapses in RC¹⁶⁷. Postoperative RT in combination with fluorouracil-based chemotherapy, was shown to reduce local recurrences significantly¹⁶⁸, and was recommended by the National Cancer Institute as standard treatment after resection of stage II-III rectal cancer¹⁶⁹. According to the current European guidelines, chemoradiotherapy should preferably be given preoperatively, but postoperative treatment is adequate, when CRM turns out positive or the tumour has been under-staged preoperatively and the patient has thus not received prior chemoradiotherapy⁵.

2.2.7.6. Novel agents in chemoradiation

2.2.7.6.1. Epidermal growth factor receptor inhibitors

Cetuximab is a monoclonal antibody to epidermal growth factor (EGFR). It regulates cellular proliferation, survival, and differentiation¹⁷⁰. In head and neck cancer, cetuximab given concomitantly with RT was shown to improve locoregional disease control and overall survival, as compared to RT alone¹⁷¹. Cetuximab combined with preoperative chemoradiotherapy in locally advanced rectal cancer was demonstrated to result in an overall 9.1% pCR rate

¹⁷⁰, which is not superior to previous reports with chemotherapy and radiation. Cetuximab is known to cause G1 cell cycle arrest ¹⁷², thereby its concurrent use with radiotherapy may revoke the additive effect of FU ¹⁷⁰. Resistance to anti-EGFR-therapy may also be involved ¹⁷³. Currently, cetuximab is not recommended in combination with chemoradiotherapy outside clinical trials in rectal cancer ¹⁷⁰. Panitumumab, a fully human monoclonal antibody to EGFR, is being investigated in the preoperative treatment setting in rectal cancer ¹⁷⁴.

2.2.6.7.2. Bevacizumab

Vascular endothelial growth factor (VEGF) inhibitor bevacizumab was shown to be beneficial in metastatic colorectal cancer ¹⁷⁵. A phase II trial examined bevacizumab in combination with capecitabine and preoperative radiotherapy in locally advanced rectal cancer and showed a promising pathologic complete response rate but also major wound healing complications ¹⁷⁶. Bevacizumab is currently not indicated in the treatment of localised rectal cancer.

2.2.7.7. Adjuvant chemotherapy

In CRC, postoperative adjuvant chemotherapy is usually delivered according to the current standard recommendations, adjusted for a patient's individual recurrence risk. Adjuvant therapy is considered for patients with stage III and high risk stage II cancer ¹⁷⁷ (**Table VII**).

FU combined to levamisole ¹⁷⁸ and later on to leucovorin became the standard adjuvant treatment in stage III CC ¹⁷⁹. A regimen of continuous infusion FU with leucovorin (LVFU2) was then shown to be less toxic than the bolus regimen ¹⁸⁰. Again, capecitabine was shown to be an effective alternative to fluorouracil-leucovorin combination (FULV) in the adjuvant setting in a non-inferiority trial ¹⁸¹. The Mosaic study demonstrated an improved overall survival in stage III CC with the addition of oxaliplatin to FULV (72.9% vs. 68.7%). DFS at five years was 66.4% for the oxaliplatin treatment group, as compared with 58.9% for the FULV group ¹⁸². However, there is somewhat less evidence supporting the use of adjuvant chemotherapy in RC than in CC ^{5, 122, 183}. Nevertheless, adjuvant treatment is generally used in RC along the same principles as in CC. EORTC- 22921 trial showed no significant benefit of adjuvant chemotherapy after preoperative treatment for the whole study population. However, good prognosis patients, who responded to preoperative treatment, had longer DFS after adjuvant chemotherapy than patients who did not receive adjuvant chemotherapy ¹⁸⁴. Interestingly, a meta-analysis of five randomised Japanese trials reported improved overall survival

and DFS for RC patients with uracil-tegafur adjuvant chemotherapy¹⁸⁵. In western countries, this agent is not currently used due to associated severe toxicity. In Japanese population, severe gastrointestinal toxicities appear to be less frequent for an unknown reason¹⁸⁵.

2.2.8. Multidisciplinary approach

The treatment of rectal cancer patients is preferably discussed in a multidisciplinary team, involving representatives from colorectal surgery, pathology, medical oncology, radiotherapy and radiology to ensure adequate treatment planning¹⁸⁶.

2.2.9. Current treatment recommendations

According to the European Society for Clinical Oncology (ESMO) Clinical recommendations⁵, surgery alone is the treatment of choice of T1-T2N0 favourable cancers. Generally, preoperative short-course RT is recommended to most patients with T3-tumours, with surgery by total mesorectal excision technique on the following week. Preoperative chemoradiotherapy is recommended for locally advanced T4-tumours and T3-tumours with threatened CRM margin involvement, followed by radical surgery 6-8 weeks later. Postoperative chemoradiotherapy can be used, if CRM is positive or the tumour has perforated or was under-staged preoperatively, and thus, no treatment has been given preoperatively. Adjuvant chemotherapy can be considered for stage III and high risk stage II RC⁵. After preoperative combined therapy, treatment decisions should rather be based on the pre-treatment than the postoperative pathological stage¹²².

2.2.10. Surveillance

2.2.10.1. Tumour markers

Carcinoembryonic antigen (CEA) is the first and most widely used tumour marker in CRC surveillance¹⁸⁷, known to be up-regulated by hypoxia in cancer cells¹⁸⁸. Normal CEA does not rule out cancer recurrence and values exceeding upper normal limits may not reflect recurrent disease¹⁸⁹. However, the highest CEA values are obtained from patients with metastatic disease¹⁸⁹. Membrane-associated glycoprotein CA19-9 is not recommended for CRC

surveillance¹⁸⁹. Serum lactate dehydrogenase (LDH) level has been used as a prognostic marker in CRC. Removal of the primary tumour is followed by a decline in serum LDH within a week¹⁹⁰. High serum LDH levels are associated with tissue LDH-5 expression by IHC¹⁹⁰. LDH is known to be affected by various conditions, e.g. hypothyroidism, infection and renal insufficiency, and is not routinely used to monitor CRC.

2.2.10.2. ASCO and ESMO recommendations for follow-up

According to the American Society for Clinical Oncology (ASCO) recommendations for the use of tumour markers, the evaluation of CEA is recommended preoperatively, if it would help in staging and treatment planning. Further, the assessment of CEA postoperatively every three months for three years is recommended by ASCO, if the patient would be eligible for surgery or chemotherapy of metastatic disease. No routine use of novel tumour markers is recommended due to insufficient data¹⁹¹. According to the ESMO Clinical Practice Guidelines, a completion colonoscopy should be performed within the first year of diagnosis, as well as medical history and rectosigmoidoscopy, if feasible, every 6 months for 2 years. Thereafter, history and colonoscopy along with the resection of colonic polyps every 5 years are recommended⁵. CEA testing is recommended before treatment, but no directives for post-operative clinical examinations and laboratory tests are given since their value is unknown. Postoperative imaging of the liver and lungs is recommended at 1 and 3 years after curative surgery. A Cochrane intervention review analysis concerning follow-up strategies for patients with non-metastatic colorectal cancer was recently performed. This analysis showed intensive follow-up to be associated with improved all-cause survival, but the optimal follow-up regimen is not known¹⁹².

Lynch syndrome carriers with DNA mismatch repair gene mutation are offered colonoscopies and vaginal ultrasound at 2-3 years intervals¹⁹³, including personal counselling⁵⁸.

2.2.11. Prognosis

2.2.11.1. Adverse prognostic factors

The prognosis of RC depends on the radicality of the operation and the tumour's TNM-stage. Lymph node metastases worsen disease outcome considerably¹. A positive CRM is a strong predictor of both local disease

recurrence and distant metastasis². Histopathologic and clinical features associated with adverse disease outcome^{1, 2, 4, 97, 98, 103, 107, 108, 131-142} are listed in **Table VII**. Precise surgical and pathological procedures ensure adequate staging and postoperative treatment decisions, leading to best possible disease outcome. Optimal, radical surgery is a cornerstone in the treatment of RC and it cannot be compensated by postoperative oncological treatments¹⁹⁴.

2.2.11.2. Survival by stage

The presence of lymph node metastasis adversely affects disease outcome¹. The 5-year relative and observed survival probability of RC patients in Finland¹⁹⁵ is depicted in **Table VIII**.

Table VIII. The 5 year survival data of rectal cancer patients diagnosed in 2000-2007 in Finland.*

Extent of Disease	Relative Survival proportion/ Males	Observed Survival probability/ Males	Relative Survival proportion/ Females	Observed Survival probability/ Females
Localised disease	84,1%	67,6 %	87,2%	73,4%
Regional disease	67,7%	56,0%	64,8%	55,9%
Distant disease	18,9%	17,1 %	18,0%	14,9%

* Finnish Cancer Registry (personal communication, 2010).

2.3. TUMOUR HYPOXIA

2.3.1. General aspects of hypoxia

Hypoxia refers to insufficient levels of oxygen in blood or tissue, which also results in reduced adenosine triphosphate (ATP) production⁶. There are several mechanisms, which can cause tissue hypoxia^{6, 196} (**Table IX**). ATP is a key mediator of energy in cells. Abated ATP production causes changes in Na⁺-K⁺ channels, membrane depolarisation, enhancement of cellular uptake of Cl⁻, swelling of cells, increased Ca²⁺-concentration and decreased pH in the cytoplasm, leading to tumour cell acidosis⁶. Anoxia in turn refers to complete lack of oxygen in blood or tissue. Hypoxia induces programmed cell death, or apoptosis, both in normal and malignant tissue⁶.

Table IX. Causes of hypoxia with examples of associated clinical conditions.*

Cause	Example
Low oxygen partial pressure	Pulmonary diseases High altitude sickness
Reduced ability of blood to carry oxygen	Anemia Methemoglobinemia Carbon monoxide poisoning
Reduced tissue perfusion Impaired oxygen transport	Sepsis, hypotension, dehydration Capillary microthrombosis in sepsis Arterial thrombosis or embolism
Impaired diffusion 1) Decreased oxygen tension gradient 2) Increased diffusion distance 3) Increased vascular permeability	Excessive fluid loading, tissue oedema Capillary microthrombosis in sepsis
Inability to use cellular oxygen	Cyanide poisoning Endotoxins associated with sepsis

* Modified. Based on Höckel M, Vaupel P. *Tumor Hypoxia: Definitions and Current Clinical, Biologic, and Molecular Aspects. J Natl Cancer Inst 93(4): 266-276, 2001 and Leach RM, Treacher DF. ABC of Oxygen. Oxygen Transport-2. Tissue Hypoxia. BMJ 317: 1370-1373, 1998.*

2.3.2. Angiogenesis and hypoxia

A tumour cannot grow beyond a few millimetres without the formation of new blood vessels¹⁹⁷, angiogenesis. Several mechanisms of angiogenesis are involved in neovascularisation. New capillaries can sprout out of the existing vessels, which starts by endothelial cell activation, a process stimulated by angiogenic growth factors, including vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF) and basic fibroblast growth factor (bFGF)^{198, 199}. New vessels can also be formed by splitting of the pre-existing vessels or the tumour can grow along them, enabling avascular growth¹⁹⁹. Even tumour cells per se can dedifferentiate to endothelial phenotype and thus provide a collateral vascular system to the tumour¹⁹⁹.

2.3.3. Tumour vasculature, oxygenation and glycolysis

New tumour vessels are tortuous, disorganised and dilated, branch abnormally and are often more permeable than normal blood vessels, resulting in uneven blood flow²⁰⁰. Inadequate vasculature, insufficient blood flow and uncontrolled rapid growth result in hypoxia in the tumour^{7, 200}. Large tumours

frequently have necrotic areas caused by anoxia. However, tumours have hypoxic areas regardless of their size, stage, differentiation grade and location⁶.

Under hypoxic conditions, cells turn to lactic acid production instead of oxidative phosphorylation from tricarboxylic acid cycle (TCA) to produce energy²⁰¹, a phenomenon called anaerobic glycolysis or Pasteur Effect²⁰². However, even in an environment with normal oxygen tension, cancer cells are inclined to use anaerobic glycolysis as a source of energy (Warburg effect)²⁰³. Since aerobic glycolysis is known to be associated with higher ATP production²⁰³, it is not clear why tumours tend to use anaerobic glycolysis. One reason for this tendency could be that purine and pyrimidine pathway precursors are produced during anaerobic glycolysis, thus providing DNA building material for tumour growth²⁰⁴.

2.3.4. Measuring hypoxia

Tumour oxygenation has been measured by computerised polarographic electrode systems³¹. In general, tumours tend to have lower partial oxygen tension (pO₂) than the tissue they are originated from, and the oxygenation status deteriorates along with disease recurrence²⁰⁴. In contrast to pO₂ of 40-50mmHg in normal tissues, values as low as 5-10 mmHg have been measured in tumours³⁰.

2.3.5. Activation of the hypoxia response element and HIF-1 α

Hypoxia starts a cascade of events, including the activation of hypoxia-response element (HRE). The key protein in tissue response to oxygen deficiency is hypoxia-inducible factor-1 α (HIF-1 α). HIF-1 α is expressed in the nuclei of almost all superficial epithelial cells in the normal colorectal mucosa. The same staining pattern is seen in adenomas. However, in malignant tumours HIF-1 α is mostly present in the nuclei of cancer cells around necrotic areas²⁰⁵.

HIF-1 consists of two protein subunits, HIF-1 α and HIF-1 β ⁷⁻⁹. HIF-1 β is constitutively expressed, whereas HIF-1 α expression is oxygen-dependent⁷. In hypoxia, HIF-1 α stabilises, followed by dimerisation with the β -unit^{8, 206}. HIF-1 α binds to HRE, which is a special DNA sequence^{14, 15, 207}.

In normoxic conditions, HIF-1 α is bound to Von Hippel-Lindau protein (VHL), which is a tumour suppressor gene⁷. Binding results in HIF-1 α degradation within minutes after hypoxia is resolved²⁰⁸, by an enzymatic mechanism involving prolyl hydroxylation^{206, 209}. HIF-1 α up-regulation may also occur independently of the oxygen tension in the tumour due to defects of its degradation pathway, e.g. VHL protein mutations²¹⁰, oncogenic activation leading to increased HIF mRNA transcription, or intracellular hypoxia caused by increased oxygen consumption²¹¹. HIF-1 α stabilisation leads to the activation of several genes involved in angiogenesis, glycolysis, invasion, metastasis, apoptosis, pH regulation and growth factor signalling^{7, 10}. Many of these genes are more abundant in malignant than in normal tissues^{10, 212, 213}.

2.3.6. Association of HIF-1 α with PI3-AKT and mTOR

In breast cancer cell lines, phosphoinositide-3-kinase (PI3K)-inhibitor LY294002 decreased the expression of HIF-1 α , showing that HIF-1 α activation is regulated by the PI3K/AKT pathway²¹⁰. In VHL mutant renal cancer cell line, the PI3K inhibitor was shown to down-regulate the expression of HIF-1 α , in spite of the stabilisation of HIF-1 α . This implies that HIF-1 α is also regulated by VHL-independent pathways²¹⁰.

Mammalian target of rapamycin (mTOR) is a key regulator of HIF-1 α protein synthesis in many cancers¹⁶. Activation of receptor tyrosine kinases including AKT, HER2 and EGFR, leads to increased mTOR activity^{16, 214}, in turn inducing HIF-1 α ^{9, 16, 215}. Rapamycin, a protein kinase that inhibits mTOR activity is shown to act as a regulator of HIF-1 α in hypoxia¹⁷. Ezrin is a protein which acts as a cross-linker between the cell membrane and the cytoskeleton^{18, 19, 21}. Ezrin expression is necessary for metastatic behaviour in cancer cell lines²⁷ and ezrin-related metastatic behaviour in turn is shown to be linked to mTOR pathway²⁸. Blocking the mTOR pathway with rapamycin is found to inhibit experimental lung metastasis in vivo²⁸. Thus, the regulation of mTOR and ezrin as well as mTOR, ezrin and HIF-1 α are interrelated (**Figure 3**).

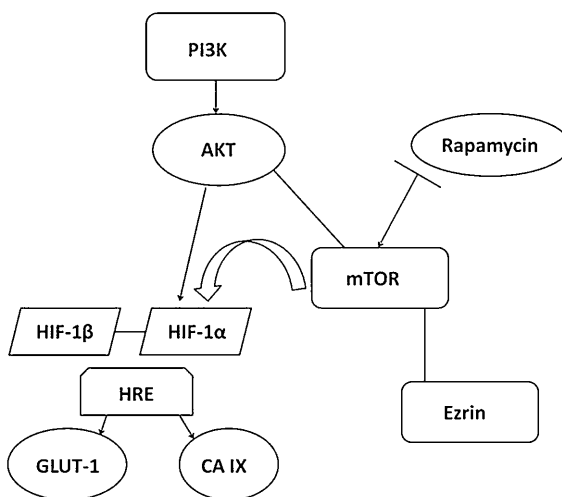


Figure 3. The association of HIF-1 α with PI3K, mTOR and ezrin. Modified, based on Wang G et al, 1993; Semenza G, 2000; Pakkanen R, 1998; Hudson C et al, 2002; Kivelä A et al, 2001; Harris A, 2002, Brambilla D et al, 2009; Niggli V et al, 2008; Fehon R et al, 2010.

2.3.7. Down-stream targets of HIF-1 α

2.3.7.1. Vascular endothelial growth factor (VEGF)

Blood vessels as well as lymphatic vessels have an essential role in tumour metastasis²¹⁶. In normal tissues there is a balance between the factors that induce and inhibit angiogenesis²⁰⁴. Hypoxia induces HIF-1 α and thereby promotes the up-regulation of vascular endothelial growth factor (VEGF), which leads to neoangiogenesis²¹⁷. This concept is called the angiogenic switch⁹, with HIF-1 α as the key player turning the switch on. VEGF up-regulation also results in increasing vascular permeability, which elevates the interstitial pressure in the tumour and deteriorates drug delivery to tissues²⁰⁴.

VEGF-A, often termed as VEGF, belongs to the VEGF-PDGF - family²¹⁸. The expression of VEGF-A and VEGF-C is shown to be increased in CRC²¹⁹ and the physiological effects of VEGF are mediated by its receptors VEGFR-1 and VEGFR-2²¹⁹. VEGFR-3 receptor was reported to be expressed in the endothelial cells of blood and lymphatic vessels²²⁰ and the expression of VEGFR-3 mRNA to be positive in most CRCs²¹⁹.

2.3.7.2. Carbonic anhydrase IX (CA IX)

Carbonic anhydrases are zinc-containing metalloenzymes that are widely present in the gastrointestinal tract. They are important enzymes in the production of saliva, gastric acid and bile and several other physiological functions¹¹. Carbonic anhydrase IX (CA IX) is a transmembrane glycoprotein²²¹, which is expressed on protein level only in some tissues, like the epithelia of bile and pancreatic ducts, gallbladder, gastric mucosa and duodenum²²². The staining weakens in small intestine progressively towards colon²²³. In normal colorectal epithelium, CA IX staining is very weak or absent, confined only to occasional cryptal cells¹¹. Instead in CRC, CA IX is significantly over-expressed, showing membranous staining pattern¹². CA IX expression is thus detectable in CRC, but not in corresponding non-cancerous tissue^{12, 221}.

Cancer cells tend to produce lactic acid²⁰³, which results in acidic environment, in order to maintain optimal acidic pH and malignant biological functions²²⁴. CA IX catalyses the reversible chemical reaction in which carbon dioxide is hydrated to carbonic acid and further to bicarbonate ion and proton^{221, 225-227}. Thus, CA IX is an important enzyme for cancer cells in hypoxic and normoxic conditions^{226, 228} in the regulation of acid-base balance²²⁹. Most hypoxic tumours have acidic pH values of 6²³⁰. Maintaining an acidic environment enhances tumour invasion and relates to poor disease outcome^{204, 225}. Carbonic anhydrases may also have a role in signal transduction²²⁴.

2.3.7.3. GLUT-1

GLUT-1 is one of the down-stream targets of HIF-1 α ¹⁰. GLUT-1 is widely expressed in tissue-blood barriers²³¹ and at variable levels in many tissues^{232, 233}. In the mucosa of normal colon, there is a gradient of GLUT-1 staining from the surface epithelium to the base of the crypts²⁰⁵. The staining is more intense in CRC than in normal mucosa²⁰⁵. In cancer tissue, GLUT-1 staining is primarily membranous^{234, 235}, but a simultaneous cytoplasmic staining is seen along with the dominantly strong membranous staining²³⁶. GLUT-1 is over-expressed in several malignancies, e.g. cancers of the colon²³⁷, and pancreas²³⁸.

Under hypoxic conditions, cells switch to use lactate production instead of oxidative phosphorylation to produce energy²⁰². Hypoxia leads to the stabilisation of HIF-1 α ^{8, 206, 209}, which results in the activation of its down-stream targets including GLUT-1^{10, 227}. Glucose is actively transported into the cell by transporter proteins²³⁹. GLUT-1 is the main carrier of glucose in malignant

tumours²³¹. In addition to hypoxia, GLUT-1 is also regulated by inhibitors of oxidative phosphorylation²⁴⁰.

2.3.7.4. Ezrin

Ezrin belongs to the ezrin-radixin-moesin (ERM) family of proteins^{18, 19}. The ERM proteins function as cross-linkers between the plasma membrane and the cytoskeleton^{21, 22}.

Ezrin, also known as cytovillin, is widely expressed in several normal and malignant tissues of both epithelial and non-epithelial origin^{23, 241}. In normal colorectal mucosa, ezrin is mainly expressed on the apical cell membrane in a polarised fashion, whereas in cancer cells, the staining is predominantly cytoplasmic²⁴². Staining is also less intense in normal colorectal mucosa than in carcinoma²⁴³.

Ezrin has an active C-terminal, which is connected to the actin cytoskeleton and an N-terminal, located to the plasma membrane²¹. It is present in the cytoplasm in its inactive form²⁴⁴. Stimulation leads to ezrin phosphorylation and translocation to the ruffles, which in turn increases motility and proliferation^{18, 19, 241, 245, 246}. In cell cultures and in tumour samples, ezrin is distributed both underneath the plasma membrane and in the cytoplasm, where it links transmembrane proteins, especially adhesion molecules to actin cytoskeleton^{20, 247, 248}. Ezrin is also involved in the control of cell morphology, motility, proliferation and survival, as well as in signal transduction by protein kinase C, Rho-kinase, EGFR, Src, mTOR and PI3K/AKT pathways²⁴⁻²⁶.

2.3.8. Effect of hypoxia on resistance to treatment

The key mechanism for malignant progression as well as resistance to therapy in hypoxia is the selection of neoplastic cells insensitive to apoptosis³¹. Well-oxygenated cells sensitive to radiotherapy are killed, and resistant hypoxic cells continue to proliferate and are thus selected to survive¹⁰¹. Apoptosis is an essential mechanism in the response to radiotherapy and chemotherapy²⁴⁹.

2.3.8.1. Radiotherapy and hypoxia

2.3.8.1.1. Oxygenation and reoxygenation

Radiosensitivity depends on various factors; including cell cycle phase, tumour type and size, and most importantly, oxygen tension¹⁰¹. Radiation causes DNA damages to cancer cells by formation of free radicals. Irreparable lethal double-strand breaks cause cancer cell death by apoptosis. Apoptosis, or programmed cell death is regulated by pro- and antiapoptotic factors, which in turn are regulated by HIF-1 α ²⁵⁰. Cancer cells that have suffered sublethal damages can escape apoptosis, especially in hypoxic conditions²⁵⁰. Oxygen is important for radiosensitivity due to its participation in free radical generation, free radical damage fixation and limiting chemical repair¹⁰¹. Cancer cells can also be injured by the effect of radiation on the microvasculature, supporting stroma or parenchymal cells¹⁰¹. Poorly oxygenated tissues are less sensitive to radiation than well oxygenated tissues, thus impairing the therapeutic effect: a three-fold higher radiation dose is required to achieve the same biological effect as in normoxic conditions²⁵¹. Moreover, oedema, frequently present in malignant tissues during radiotherapy may deteriorate the hypoxic conditions, further aggravating radioresistance²⁵². Thus, oxygen is the most important and potent radiosensitiser^{253, 254}.

Reoxygenation is seen in tumour tissue after exposure to radiation. In mouse mammary carcinoma, the oxygen levels in tumour tissue before radiation have been shown to resemble those of advanced cancer, but after radiation a significant improvement in oxygenation has been found. Reoxygenation is particularly evident after very low pO₂²⁵⁵. Early reoxygenation has been studied in animal tumour models, where increased oxygen pressure has been observed starting 3-4 hours after radiation and being elevated for at least 24 hours after radiation²⁵³. Reoxygenation may occur due to decreased interstitial pressure and better oxygen diffusion after irradiation caused by a loss of tumour cells, resulting in increased radiosensitivity²⁵³.

Proliferating cells are more prone to radiation damages than cells in resting phase¹⁰¹. Fractionated treatment schedule enhances the possibility of targeting proliferating cells^{101, 253}. Repopulation, or accelerated proliferation and repair of sublethal damages also take place in cells after RT, implicating that RT is best given without any additional breaks in the treatment¹⁰¹.

2.3.8.1.2. Nitroimidazoles and anti-inflammatory drugs in radiosensitisation

Nitroimidazoles, including metronidazole and misonidazole, can mimic the effect of oxygen and have therefore been studied for their possible radiosensitising impact. Due to associated toxicities it has not been possible to use them at effective doses. However, a meta-analysis of the trials revealed slightly improved local control with the use of nitroimidazoles²⁵⁶. Cyclooxygenase-2 (COX-2) is an enzyme that catalyses the formation of prostaglandin H₂ from arachidonic acid²⁵⁷. COX-2 over-expression has been shown to associate with poor prognosis in many cancers²⁵⁴ and COX-2 inhibitors to induce apoptosis in colon cancer cell lines²⁵⁷. In tumour model studies, COX-2 inhibitors and other tested non-steroidal anti-inflammatory agents (NSAID) have been found to increase pO₂, leading to enhanced tumour radiosensitivity. These findings implicate that NSAIDs can act as radiosensitisers²⁵⁴. However, COX-2 inhibitors are found to increase the risk of cardiac events and all NSAIDs the risk of upper gastrointestinal tract problems, thus restraining their exploitation as radiosensitisers²⁵⁸.

2.3.8.1.3. Chemotherapeutic agents in radiosensitisation

The addition of fluorouracil to postoperative radiotherapy has been found to improve disease outcome in rectal cancer as compared to radiotherapy alone^{168, 259}. An increase in the rate of pCR and a decrease in local relapses has been demonstrated with the use of preoperative chemoradiotherapy when compared to radiotherapy alone¹⁵³. The benefit of chemotherapy can be explained by its systemic effects, but favourable tumour regression and a decline in local disease recurrences points out the possibility of fluorouracil acting as a radiosensitiser²⁵⁹.

Cisplatin, which forms DNA adducts, leading to increased apoptosis through the activation of several signalling pathways²⁶⁰, has been shown to act as a radiosensitiser²⁶¹. Oxaliplatin is a platinum analogue currently used in the treatment of colorectal cancer, both in the adjuvant and the metastatic setting¹⁸². Oxaliplatin has been reported to increase the efficacy of radiation even when the doses are diminished, as long as a threshold dose is maintained²⁶². In phase II studies, oxaliplatin combined with capecitabine and radiotherapy preoperatively, has been shown to evoke excellent responses in rectal cancer¹⁵⁹.

Some agents are toxic to hypoxic cells only, which are resistant to conventional cytotoxic treatments. These drugs include chemotherapeutic agent mitomycin C and a more selective drug tirapazamine²⁶³, which brings about single and

double strand DNA damages more lethal than those produced by ionising radiation²⁶⁴. In locally advanced cervical cancer, tirapazamine in combination with radiation and weekly cisplatin has, however, been found to associate with more severe toxicity than anticipated²⁶⁵.

2.3.8.1.4. Oxygen manipulation

Hyperbaric oxygen

Since oxygen is critical to the efficacy of treatment, hyperbaric oxygen should in theory potentiate the benefit of radiotherapy. This rationale has been studied in malignant glioma, showing longer median survival with than without hyperbaric oxygen²⁶⁶. This approach, however, is not routinely available and does not take into account other factors related to hypoxia, including impaired blood flow in tumour vessels²⁶⁷.

Treatment of anaemia

Lower than 120 g/L haemoglobin level, has been shown to associate with increased pelvic recurrence in cervical cancer. However, correction of anaemia by blood transfusions and maintaining the haemoglobin level ≥ 120 g/L has been observed to improve survival rates for these patients²⁶⁸. Average weekly haemoglobin levels have also been shown to be coupled with disease outcome in cervical cancer²⁶⁹. Erythropoiesis-stimulating agents (ESAs) have been variably used in the treatment of anaemic cancer patients, but concerns of its safety were brought about by the observations of increased risk of thromboembolism²⁷⁰. A meta-analysis of controlled ESA oncology trials was carried out and no significant influence of ESAs on disease progression or survival was found but an increased risk for thromboembolic events was revealed²⁷¹. Moreover, the activation of hypoxia-induced pathways have been shown to occur independently, irrespective of the patient's haemoglobin levels²⁷², further emphasising the complexity of the effects of tumour hypoxia and its regulation.

2.3.8.3. Chemotherapy and hypoxia

The mechanisms of chemoresistance are not well known²⁷³. One mechanism involved in hypoxia-related chemoresistance could be linked to HIF-1 α , which is shown to regulate VEGF²⁰⁴. HIF-1 α activation leads to the up-regulation of VEGF, which in turn increases vascular permeability. VEGF up-regulation elevates interstitial pressure and causes impaired drug delivery to tissues²⁰⁴. Hypoxia appears to render cells resistant to chemotherapy²⁶³. In malignant

colon, ovarian and liver cell lines, oxygen deprivation before and during etoposide and oxaliplatin treatment have been found to result in down-regulation of proapoptotic proteins and drug resistance to etoposide. These results implicate hypoxia to lead to apoptosis inhibition²⁷³. Loss of tumour-suppressor gene p53 or over-expression of apoptosis inhibitor BCL-2 is known to inhibit apoptosis in hypoxic cervical cancer tissue²⁷⁴. Clinical studies have shown that over-expression of HIF-1 α is associated with unfavourable outcome in several human cancers, linking hypoxia with chemoresistance^{10, 29}.

2.3.8.4. Hypoxia, response to therapy and disease outcome

Hypoxia, oncogene activation or loss of function of suppressor genes result into increased activation or decreased degradation of HIF-1 α ⁹, leading to its over-expression in many cancers, including CRC²⁰⁵. HIF-1 α has a central role in tumour progression, resistance to treatment and disease outcome⁹. A significantly worse DFS and overall survival has been observed with hypoxic cervical cancers, as compared with normoxic tumours³⁰. Also, larger and thus more hypoxic tumours tend to be accompanied by more frequent parametrial and lymphovascular involvement than well oxygenated tumours³⁰. In clinical studies concerning head and neck or cervical cancer, poor tumour oxygenation has been associated with increased tumour invasion, metastasis, unfavourable disease outcome^{29, 275} or resistance to radiotherapy^{6, 9, 276}. HIF-1 α expression has also been shown to be linked to Duke's stage²⁷⁷ and lymph node status in rectal cancer²⁷⁷. Patients, whose tumours express HIF-1 α strongly, have shorter overall^{277, 278} and local relapse-free and disease-free survival^{277, 278}. Previous studies on the prognostic significance of HIF-1 α , CA IX, GLUT-1 and ezrin with regard to outcome of human tumours are depicted in **Table X**. In general, strong expression is related to unfavourable disease outcome^{27, 30, 227, 242, 272, 275, 277-294}. However, in renal cancer, low CA IX expression and absence of VHL mutation have been shown to be related to more advanced tumours and unfavourable outcome²⁸⁵⁻²⁸⁸.

Table X. HIF-1 α , CA IX, ezrin and GLUT-1 as prognostic/predictive markers in human malignancies in previously published studies.

Biomarker	Type of cancer	Prognostic/predictive significance of high expression	Reference
CA IX	Head and neck cancer	Unfavourable	Bache et al, 2006
	Non-small cell lung cancer	Unfavourable	Giatromanolaki et al, 2001 Swinson et al 2003
	Breast cancer	Unfavourable	Brennan et al, 2006 Generali et al, 2006
	Renal cell cancer	Favourable	Bui et al, 2003, 2004 Atkins et al, 2005 Patard et al, 2008
HIF-1α	Cervical cancer	Unfavourable	Höckel M, 1996 Birner et al, 2000
	Head and neck cancer	Unfavourable	Lu X et al, 2006 Koukourakis M et al, 2002
	Rectal cancer	Unfavourable	Theodoropoulos G et al, 2006
Ezrin	Colorectal cancer	Unfavourable	Elzagheid et al, 2008
	Head and neck cancer	Unfavourable	Mhavec-Fauceglia P et al, 2007
	Ovarian cancer	Unfavourable	Köbel M et al, 2006
	Malignant melanoma	Unfavourable	Mäkitie et al, 2001
GLUT-1	Rectal cancer	Unfavourable	Brophy et al, 2009 Cooper et al, 2003
	Colorectal cancer	Unfavourable	Haber et al, 1998
	Gastric cancer	Unfavourable	Kawamura 2001
CA IX & GLUT-1	Bladder cancer	Unfavourable	Hoskin et al, 2003

3. AIMS OF THE STUDY

Hypoxia is shown to have an impact on disease outcome and resistance to treatment^{29,30}. Hypoxia activates HIF-1 α , leading to the activation of its downstream targets CA IX and GLUT-1^{7,8}. The focus of this study is to evaluate hypoxia-associated biological markers HIF-1 α , CA IX and GLUT-1, as well as cell skeleton protein ezrin, also linked to HIF-1 α , in RC treated by preoperative RT or chemoradiotherapy. The aim of the study is on one hand to compare by IHC the expression of these biomarkers in the diagnostic biopsies with the expression in the operative samples, and on the other hand in relation to disease outcome. In addition, the aim is to evaluate histopathologic features in all tumour samples and tumour regression grade in the operative samples after long-course RT and compare them to the results of IHC stainings.

The specific aims of the study are

- 1) To analyse the expression of the key protein in hypoxia, HIF-1 α in the diagnostic biopsies and operative samples after preoperative radiotherapy or chemoradiotherapy, the impact of preoperative treatment on HIF-1 α expression and the association of HIF-1 α expression with disease outcome and tumour regression grade.
- 2) To evaluate the expression of CA IX, regulated by HIF-1 α , in the diagnostic biopsies and operative samples after preoperative radiotherapy or chemoradiotherapy, as well as the influence of preoperative treatment on CA IX expression and the association of CA IX expression with tumour regression grade and survival.
- 3) To study the expression of cell skeleton protein ezrin, also linked to HIF-1 α , in the diagnostic biopsies and operative samples after preoperative radiotherapy or chemoradiotherapy, the impact of preoperative treatment on ezrin expression and ezrin expression in relation to tumour regression grade and survival.
- 4) To investigate the expression of GLUT-1, a downstream target of HIF-1 α , in the diagnostic biopsies and operative samples after preoperative radiotherapy or chemoradiotherapy, the effect of preoperative treatment on GLUT-1 expression and the relation of GLUT-1 expression to disease outcome and tumour regression grade.

4. PATIENTS AND METHODS

4.1. PATIENTS

This retrospective, non-randomised study included 178 consecutive RC patients treated at Turku University Hospital according to the standard treatment protocols in the years 2000-2009. Patients with metastatic disease, those with upper rectal cancer or superficial tumours treated by excision only were excluded and only adenocarcinomas were included, in order to build up a biologically and therapeutically homogenous patient population. Preoperatively, the patients were staged by CT or MRI and digital examination of the rectum, CT of the abdomen and x-ray or CT of the chest. Since 2005, rectal cancer treatments in this hospital have been planned by a multidisciplinary team. The number of patients in each study is depicted in **Table XI** and the patient population described in detail in the original publications I-IV and in **Table XII**.

Table XI. Number of patients and tumour samples in each study

Study/Biomarker		Number of patients/tumour samples				
		Preoperative biopsies	Operative samples			
			Total	Short-course RT	Long-course RT	Control
I	CA IX	80	166	75	37	54
II	HIF-1 α	79	168	75	39	54
III	Ezrin	76	176	76	46	54
IV	GLUT-1	78	175	76	46	53

Table XII. The clinical characteristics of the patients in the study (n=178).

Variable	n (%)
Patient gender	
Male	106 (60)
Female	72 (40)
Type of preoperative treatment	
None	54 (30)
Short-course radiotherapy (RT)	77 (43)
Long-course RT, no chemotherapy	10 (6)
Long-course RT with chemotherapy	37 (21)
Preoperative T	
T1-T2	33 (18)
T3	55 (31)
T4*	47 (26)
TX [†]	43 (24)
Preoperative N	
N+ [‡]	74 (42)
N0	41 (23)
Ns [§]	24 (13)
NX [†]	39 (22)
Preoperative G	
G1	22 (12)
G2	53 (30)
G3	14 (8)
GX [†]	89 (50)
Postoperative stage	
Stage I	44 (25)
Stage II	56 (31)
Stage III	76 (43)
No vital cancer	1 (<1)
Stage not assessable	1 (<1)

* including T3 tumours with threatened CRM involvement, [†] not defined, [‡] lymph nodes visualised in imaging, [§] lymph nodes visualised in imaging, suspected metastatic. RT= radiotherapy

The patients were treated with short-course (5x5 Gy) (n=77) or long-course radiotherapy (50.40 Gy) (n=47) or received no preoperative treatment (n=54), depending on the stage and localisation of the tumour and following the standard treatment protocols ^{5, 147}, based upon the judgment of the surgeon and the multidisciplinary team. Long-course RT was given with (n=37) or without (n=10) concomitant chemotherapy. Postoperative adjuvant chemotherapy was given to patients with lymph node positive or high-risk lymph node negative tumours, according to the standard practice ¹⁸⁵. As a control group (n=54), we studied a third series of patients who had not

received any treatment prior to surgery. Postoperative RT, chemoradiotherapy or adjuvant chemotherapy were given to eligible control group patients when indicated (19/54, 35%). After completion of the treatment protocols, patients were scheduled for further follow-up at the Department of Surgery. The median follow-up time of the patients was 35 months and the mean follow-up time 40 months (range 2-114 months).

The mean age of the patients was 67 years (median 69 years, range 34-92 years). All tumours in this study were adenocarcinomas, of them 4% were classified as mucinous. Of the operations, 96 (54%) were performed by AR. Since the introduction of the multidisciplinary team in 2005, the proportion of AR was higher, 63%. Surgery was macroscopically radical in 175/178 (98%). The CRM was ≤ 1 mm in 23/125 (18%), and more than 1mm but ≤ 2 mm in 7/125 (6%) of the patients. The number of examined lymph nodes was less than 12 in 88 (50%) and at least 12 in the rest of the operative specimens. The number of metastatic lymph nodes was < 4 (N1- tumours) in 47/178 (26%) and ≥ 4 (N2-tumours) in 25/178 (14%) of the tumours. The percentage of patients diagnosed before year 2005 was 39%. During follow-up, 12 patients were diagnosed with locally relapsed and 43 patients with metastatic disease. Of the metastatic disease patients, 4 patients also had verified local relapses concurrently at 3 years follow-up. The survival data (updated until 2010, May) of the patients are depicted in **Table XIII**. The survival curves of the patients with regard to postoperative stage are shown in **Figures 4a and b**. The mean disease-free survival (DFS) was 56 months for stage I, 41 months for stage II and 33 months for stage III patients. The mean disease-specific survival (DSS) was 60 months for stage I, 51 months for stage II and 44 months for stage III patients. The mean DFS was 26 months for patients with CRM < 2 mm and 36 months for those with CRM ≥ 2 mm, and the mean DSS were 34 months and 44 months, respectively. The mean DSS for patients with only local relapse was 41 months and for those with distant relapse 40 months.

The study protocol was approved by the ethics committee of the Hospital District of Southwest Finland and the collection and use of archival tissue material by the National Authority for Medico-Legal affairs. The study was conducted in accordance with the Declaration of Helsinki.

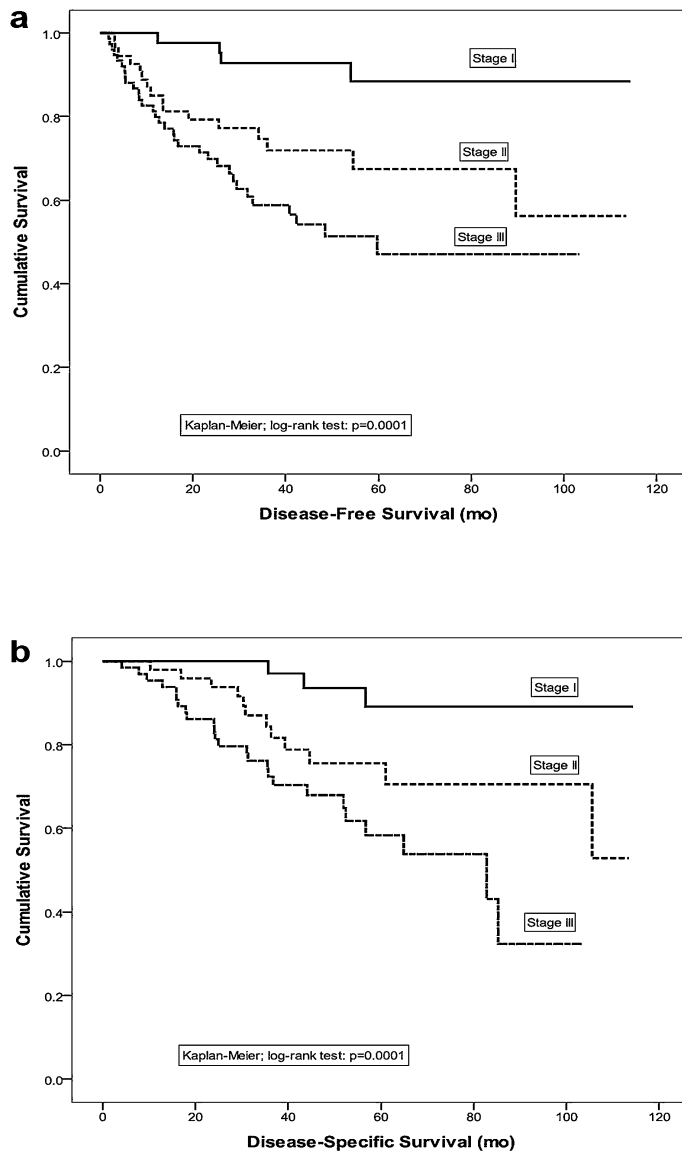


Figure 4. a) Disease-Free Survival and b) Disease-Specific Survival with regard to postoperative stage.

Table XIII. Disease outcome of the patients at the latest follow-up (May 2010)

Disease outcome	n (%)
Alive without cancer	105 (59)
Alive with cancer	12 (7)
Died of cancer	40 (23)
Died of other causes	21 (12)

4. 2. METHODS

4.2.1. Immunohistochemical (IHC) stainings

4.2.1.1. IHC staining procedure

The dilutions of the primary antibodies were optimised by a series of pilot stainings. The pre-treatments were chosen by the recommendations of the manufacturers. In each case, the most representative blocks were selected, cut to 5µm sections. The specimens were pre-treated in microwave oven twice for 7 minutes in 10 mM sodium citrate buffer, pH 6.0, and subjected to IHC staining with the following antibodies: polyclonal rabbit anti-Human CA IX (antibody 15086, Abcam, Cambridge, UK), diluted to 1:8000 concentration (I); monoclonal mouse anti-human HIF-1α (BD Transduction Laboratories San Jose, CA, CAT# 610958), diluted to 1:100 concentration (II); mouse monoclonal IgG antibody to human ezrin (clone 3 C12)²⁹⁵ (III), provided by Antti Vaheri, final dilution 0.3 mg/ml, and polyclonal rabbit anti-human GLUT-1 (Alpha Diagnostic International, rabbit anti-human GLUT-1 IgG, affinity pure, San Antonio, TX, USA), and diluted to 1:1000 (IV). To detect the immunoreactions, a Power Vision poly-HRP IHC Detection system (Immunovision Technologies Co, Burlingame, CA, USA), was used.

4.2.1.2. Evaluation of the IHC staining

The IHC staining of the samples was evaluated by two observers (EK and JS), blinded to all clinical data and radiological findings. Light microscopes with 4x and 10x objectives (EK) and 5x and 10x objectives (JS) were used for evaluation. The analyses of the IHC staining of CA IX, HIF-1α, ezrin and GLUT-1 are described in detail in publications I-IV.

4.2.1.2.1. CA IX (I)

IHC staining in the operative samples was graded using three approaches: 1) general grouping of the cases into positive or negative, 2) proportion of positive staining, and 3) staining intensity. The slides were assessed as negative if the proportion of positive carcinoma cells in the section was less than 10%. For positive cases, the proportion of positively stained carcinoma cells was analysed using three categories: 1) 10-25%, 2) 26-50%, and 3) over 50%. The staining intensity was defined as 0 = negative, 1 = weak, and 2 = moderate and 3 = strong staining intensity. Weak staining intensity was defined as a faint positive staining in the cytoplasm and occasional staining in the cell membranes impossible to detect at a small magnification (objectives

4x-5x). The staining was assessed as moderate or strong, if the positive reaction in the cell membranes could easily be identified at a small magnification (objectives 4x–5x). In preoperative biopsies, only positive/negative staining and staining intensity were assessed.

4.2.1.2.2. HIF-1 α (II)

HIF-1 α staining was graded using three approaches: 1) general grouping of the cases into positive or negative, 2) proportion of positive staining in cancer cells, and 3) staining intensity. For positive cases, the number of visual fields in 10x10 grid containing carcinoma cells was first calculated in the HE-slides with a small magnification (objectives 4-5x). The proportion of fields with stained cancer cell nuclei was further calculated in relation to the fields containing carcinoma cells, giving a percentage of positive fields. Staining intensity was then evaluated using three categories: 0 = negative, 1 = weak, 2 = moderate and 3= strong staining intensity. In weak staining intensity there was a faint positive staining in the nucleus. The staining was assessed moderate or strong, if the positive reaction in the nuclei could be easily identified, corresponding to the staining intensity seen in the nuclei of renal carcinoma cells, which was used as a positive control. From the preoperative biopsies, only positive/negative staining and staining intensity were assessed, because of the small size of the samples.

An immunoreactive staining score (IRS) was calculated in order to take both the percentage and the intensity of staining into account. The proportion of positively stained cells was given scores as follows: 0 = 0-4%, 1= 5-10%, 2=11-29%, 3=30-59% and 4=60% and over. The product of the percentage score and intensity grade (0-3) was calculated, giving a score (IRS) of 0-12.

4.2.1.2.3. Ezrin (III)

Ezrin was evaluated by analysing the staining intensity, using four categories: negative, weak (1+), moderate (2+) and strong (3+), corresponding to the staining intensity in the adjacent lymphocytes. There was a weak membranous staining in normal rectal epithelium, concentrated at the apical cell surface, whereas in RC, the staining pattern was mostly cytoplasmic. However, in some tumours concurrent focal membranous staining was also seen.

4.2.1.2.4. GLUT-1 (IV)

GLUT-1 staining was primarily membranous, but a concurrent focal cytoplasmic staining was seen in some cases. The analyses were based on the membranous staining, graded by two approaches: the percentage of positive

tumour cells and staining intensity. Staining intensity was graded as 0 if negative, 1 if weak, 2 if moderate, and 3 if strongly positive. The proportion of positive cells was given scores in the following way: 0 = 0%, 1 = 0-2%, and 2 = 3-9%, 3 = 10-25%, 4 = 26-50% and 5 >50%. The staining in the biopsies was only assessed as negative or positive, in addition to intensity.

4.2.1.2.5. HE-staining

HE- staining was performed according to the standard laboratory protocols.

4.2.2. Western Blot analysis (WB) (I, III)

4.2.2.1. Confirming CA IX expression in WB (I)

To confirm the IHC results of CA IX staining, a small fresh tissue material from CC patients (n = 4) was further studied for the detection of CA IX protein in WB. CA IX primary antibody (ab15086, Abcam, Cambridge, UK) was diluted 1:1000 for western detection. Horseradish peroxidase conjugated anti-rabbit immunoglobulins (Dako, Glostrup, Denmark) and Pierce ECL Western Blotting Substrate (Thermo Scientific, Rockford, IL, U.S.A.) were used according to the manufacturers' instructions for visualisation of the signal.

4.2.2.2. Ezrin in relation to radiation by WB (III)

Two different cell lines with well characterised ezrin expression pattern^{296, 297} were analysed to test whether ezrin expression is directly regulated in response to radiation. Human osteosarcoma cells (U2OS) express wild-type p53²⁹⁸, whereas p53 is mutated in human glioma cells (U251)²⁹⁹. The cultured cells were incubated, then irradiated and analysed by immunoblotting with ezrin 3C12 mAb²⁹⁵, p53 (DO-1) Ab, anti-phospho-histone H2A.X (Ser 139) Ab (Upstate, Temecula, CA, USA) and α -tubulin mAb (Sigma-Aldrich, St Louis, MO, USA) using enhanced chemiluminescence detection (Amersham Biosciences, Uppsala, Sweden).

4.2.3. Tumour regression grade evaluation (I-IV)

Tumour regression grade (TRG) was analysed from HE-stained sections of the operative samples after long-course RT according to the modified Dworak and Rödel scales, using three categories: poor, moderate and excellent response^{106, 107} (study I). The response was assessed as poor, if only minimal or no tumour regression was seen and there was a dominant tumour mass left (Dworak 0-1). When only a few tumour cells or tumour cell groups were left in the primary

tumour, lymph nodes or perirectal fat, the response was assessed as moderate (Dworak 2). The response was defined as excellent if there were only very few or no tumour cells left (Dworak 3-4). In case of moderate and excellent response, a total of 2-8 separate histological slides were studied to confirm the regression grade. Examples of the different degrees of TRG are presented in **Figure 5**.

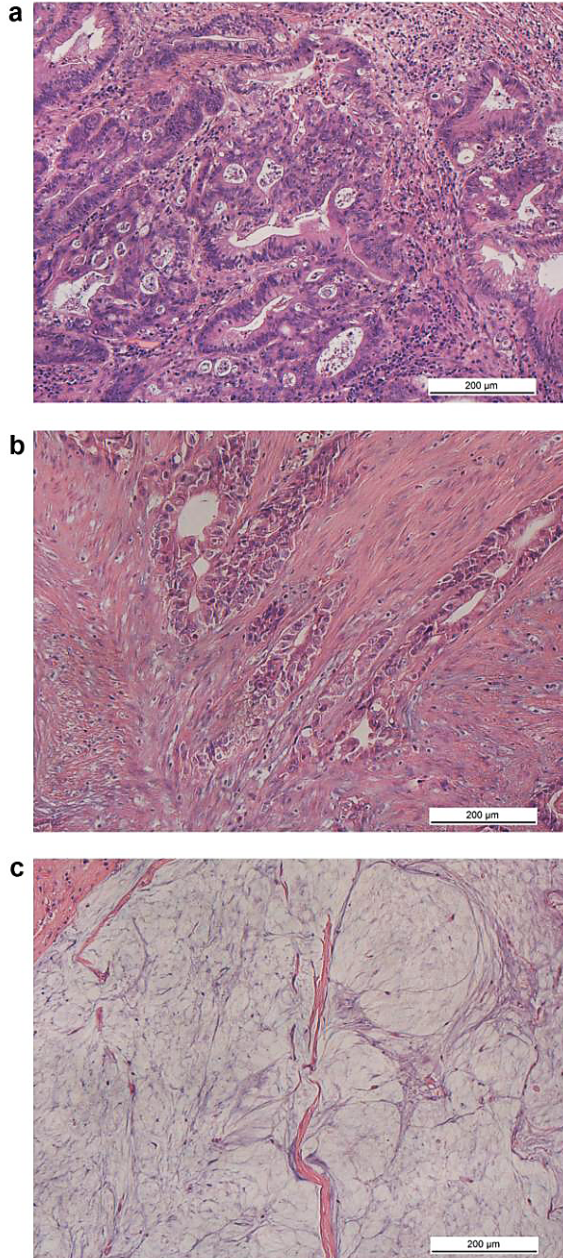


Figure 5. Tumour regression after preoperative long-course radiotherapy: A) poor, B) moderate, and C) excellent tumour regression.

4.2.4. Statistical analysis (I-IV)

All statistical analyses were run using SPSS[®] (SPSS, Inc., Chicago, USA) and STATA[™] (Stata Corp., College Station, TX, USA) software packages (SPSS for Windows, version 17.0.2 and STATA/SE 11.1). Frequency tables were analysed using the χ^2 -test, with the likelihood ratio (LR) or Fisher's exact test for categorical variables. Differences in the means of continuous variables were analysed using Mann-Whitney's test or Kruskal-Wallis test for two and multiple independent samples, respectively. Concordance of IHC staining between preoperative biopsies and operative samples was analysed using non-parametric paired-samples test (Wilcoxon signed ranks test, or McNemar test for dichotomous (+/-) variables. Inter-observer reproducibility of the IHC assessments was tested using regular (Cohen's) kappa and weighted kappa. To calculate the latter, the ICC (intra-class correlation coefficient) test was used, with parallel mode and two-way random model, using consistency assumption and average measures option to interpret the ICC (95%CI). The inter-observer reproducibility of all CA IX assessments was excellent (with κ value 0.988 for CA IX+/-, 0.931 for staining proportion and 0.892 for staining intensity), for HIF-1 α good (with κ value 0.963 for HIF+/-, 0.864 for staining intensity and 0.843 for HIF percentage), for ezrin good (with κ value 0.671) and for GLUT-1 moderate to good (with κ value 0.500 for the percentage of positive cells, 0.500 for staining intensity and 0.587 for GLUT-1 +/-).

To analyse ezrin as a predictor of DSS, performance indicators [sensitivity (SE), specificity (SP), positive (PPV) and negative (NPV) predictive value] were calculated using the contingency tables, and 95% CI calculated with the exact method.

Univariate survival analysis for DFS, and DSS was based on the Kaplan-Meier method, where stratum-specific outcomes were compared using log-rank (Mantel-Cox) statistics. To adjust for covariates, Cox proportional hazards regression model was used, co-variates being entered in a stepwise backward manner. All statistical tests were two-sided and declared significant at p -value <0.05 level.

5. RESULTS

5.1. GENERAL ASPECTS OF IHC STAINING (I-IV)

HIF-1 α staining was primarily confined to the nuclei of cancer cells, while CA IX and GLUT-1 staining patterns were predominantly membranous, and ezrin staining mostly cytoplasmic.

HIF-1 α , CA IX, ezrin or GLUT-1 expression in the operative samples were not related to patient age, gender, preoperative T (the depth of tumour invasion), preoperative N (nodal status), the type or radicality of the operation, postoperative stage, postoperative tumour differentiation (G), the number of metastatic (1-3 vs. ≥ 4) or examined lymph nodes (< 12 or ≥ 12), CRM or vessel invasion. In the multivariate models, the number of metastatic lymph nodes was a significant predictor of DSS in HIF-1 α and ezrin analysis and of DFS in CA IX analysis. In HIF-1 α analysis, preoperative treatment group predicted DFS. Unfavourable disease outcome with regard to IHC analysis and treatment group is shown in **Table XIV**. **Table XV** depicts IHC staining in the preoperative RT group biopsies and control group operative samples (excluding those patients who had received postoperative RT).

5.2. CA IX (I, V)

CAIX expression was positive in 49% of the biopsies and 44% of all the operative samples. There was a significant difference in CA IX staining intensity of the tumours in the different treatment categories ($p=0.006$). Staining intensity was moderate/strong in 10/55 (18%) of the preoperative biopsies in short-course RT group, and 6/25 (24%) in the long-course RT group. Staining intensity in the operative samples was moderate/strong in 18/75 (24%) of the short-course RT group, 7/8 (87%) of the long-course RT only (without chemotherapy), 8/29 (27%) of the chemoradiotherapy group, and 16/54 (30%) of the control group. In all, positive CA IX expression in 18/38 (48 %) biopsies turned into negative and negative expression in 12/40 (30 %) biopsies turned into positive in the operative samples, while the expression in 48/78 (61%) biopsies remained constant. CA IX expression in the preoperative biopsies and the operative samples were not significantly different ($p= 0.268$ for CA IX +/- and $p= 1.000$ for CA IX intensity).

Table. XIV. Unfavourable disease outcome with regard to IHC analysis.

IHC	Expression	Sample	Unfavourable outcome/group	Significance
HIF	positive	operative sample	DSS, long-course RT with or w/o chemotherapy	0.001
CA IX	moderate/strong staining intensity	operative sample	DFS, study population	0.001
			DSS, study population	0.009
			DFS, short-course RT	0.004
			DFS, long-course chemoradiotherapy	0.041
Ezrin	moderate/strong staining intensity	preoperative biopsy	DFS, study population	0.027
			DSS, study population	0.002
		operative sample	DSS, long-course RT with or w/o chemotherapy	0.0001
GLUT-1	moderate/strong staining intensity	operative sample	DFS, long-course RT with or w/o chemotherapy	0.066

RT=radiotherapy, w/o =without

Table XV. Immunohistochemical analyses in the biopsies of the short-course and long-course radiotherapy group patients and the operative samples of the control group patients *

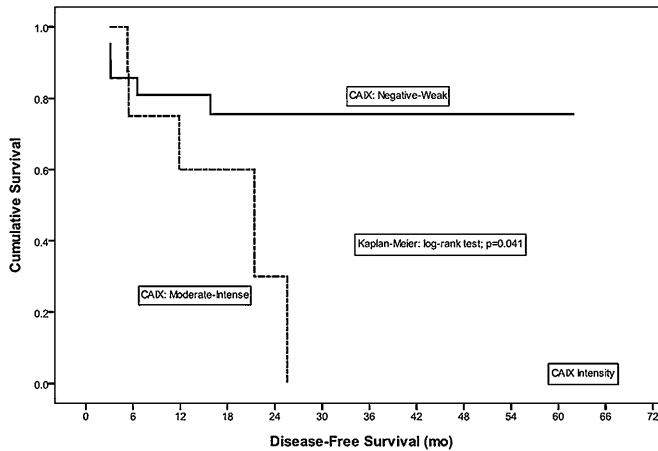
Variable	Sample type	Immunohistochemistry		Significance †
		Negative/Weak intensity n (%)	Moderate/Strong intensity n (%)	
CAIX staining intensity				
Short-course RT	biopsy	45 (82)	10 (18)	p=0.065
Long-course RT	biopsy	19 (76)	6 (24)	
Control	operative	26 (63)	14 (34)	
HIF-1α		Negative n(%)	Positive n (%)	
Short-course RT	biopsy	18 (32)	38 (68)	p=0.084
Long-course RT	biopsy	6 (25)	18 (75)	
Control	operative	20 (49)	20 (49)	
Ezrin staining intensity		Negative/Weak n (%)	Moderate/Strong n (%)	
Short-course RT	biopsy	50 (93)	4 (7)	p=0.0001
Long-course RT	biopsy	12 (52)	11 (48)	
Control	operative	20 (49)	20 (49)	
GLUT-1 staining intensity		Negative/Weak intensity n(%)	Moderate/Strong intensity n (%)	
Short-course RT	biopsy	35 (62)	21 (37)	p=0.014
Long-course RT	biopsy	11 (48)	12 (52)	
Control	operative	13 (32)	27 (66)	

* Control group patients who had received postoperative radiotherapy or chemoradiotherapy (n=13) are excluded from this table. † IHC in the biopsies of the preoperative treatment groups are compared to the operative samples in the control group.

The results of IHC staining were confirmed by WB with CRC tissue samples. Three out of four CRC samples showed strong bands with molecular weights of 50 and 56 kDa, in concordance with CA IX positivity in the IHC slides. However, in the normal colorectal mucosa and in one tumour sample only a faint band of 50 kDa was seen. These samples were CA IX negative in the corresponding IHC slides.

Patients who had tumours with negative/weak CA IX staining intensity in the operative samples had significantly longer DFS ($p=0.001$) than patients who had tumours with moderate/strong staining intensity in Kaplan-Meier (KM) analysis. Likewise, DSS was significantly ($p=0.009$) longer among patients who had tumours with negative/weak CA IX staining intensity in the operative samples. When CA IX was analysed in the preoperative biopsies, there were no significant differences in survival (unpublished data). The control group patients, who had not received any postoperative adjuvant chemotherapy or RT ($n=34$), were further compared to those, who had received postoperative RT, chemoradiotherapy or adjuvant chemotherapy ($n=20$) and no statistically significant differences were seen in DFS or DSS. In the Cox multivariate proportional hazards regression model, only two variables were independent predictors of DFS: 1) number of metastatic lymph nodes; HR 4.44 (95% CI 1.37-14.38) ($p=0.013$) for recurrence if ≥ 4 metastatic lymph nodes, and 2) CA IX intensity in the operative samples; HR 7.54 (95% CI 2.44-23.28) ($p=0.003$) for disease recurrence if moderate/strong CA IX intensity was present. In a similar Cox model, CA IX intensity in the operative samples remained the only significant independent predictor of DSS; HR 9.23 (95% CI 2.26-37.64) for dying of disease if moderate/strong CA IX intensity was present. The cumulative proportion of survivors at the 36-month follow-up time point was 83% and 35% for CA IX negative/weak and moderate/strong groups, respectively, as estimated from the survival curves.

When the treatment groups were analysed separately, most tumours (7/8, 87%) in the long-course RT group that had not been treated with concomitant chemotherapy expressed CA IX in the operative samples with moderate/strong intensity, as compared to tumours that had been treated with chemoradiotherapy (8/29, 27%). Patients who had tumours with negative/weak CA IX staining intensity in the operative samples after short-course RT or long-course chemoradiotherapy had significantly better DFS than patients, whose tumours had moderate/strong CA IX staining intensity ($p=0.004$ and $p=0.041$) (**Figure 6a, b**). No statistical differences in disease outcome with regard to CA IX staining intensity in the operative samples were seen in the control group or the long-course RT without chemotherapy group. (Unpublished data).



b)

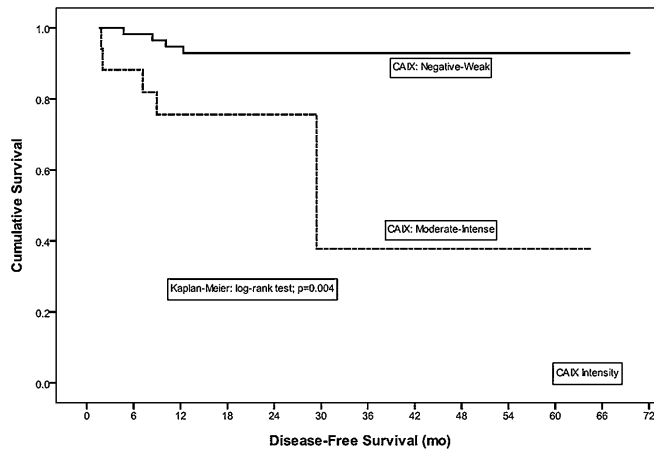


Figure 6. Disease-Free Survival in the a) long-course chemoradiotherapy and b) short-course radiotherapy groups with regard to CA IX intensity.

5.3. HIF-1 α analysis (II)

HIF-1 α was positive in 70% of the diagnostic biopsies and in 38% of the corresponding operative specimens after RT. After short-course RT, positive HIF-1 α expression in 23/37 (62 %) biopsies turned into negative and negative expression in 5/18 (28 %) biopsies turned into positive in the operative samples. After long-course RT, positive expression in 8/18 (44 %) biopsies turned into negative and negative expression in 1/6 (17 %) biopsies turned

into positive. Thus, HIF-1 α in the preoperative biopsies was down-regulated in 31/55 (56 %) and up-regulated in 6/24 (25 %) operative samples. In pair-wise comparison, the staining pattern in the biopsy-operative sample pairs was significantly different ($p=0.0001$ and $p=0.039$). After short-course RT 32% and after long-course RT 38% of the tumours were HIF-1 α positive, whereas in the control group 54% of tumours were positive ($p=0.046$).

In univariate survival analysis, no significant differences in disease outcome were seen, when HIF-1 α in the operative or preoperative samples were analysed in the whole patient population. However, long-course RT group patients whose tumours had HIF-1 α negative expression in the operative sample had significantly longer ($p=0.001$) DSS, as compared to HIF- α positive tumours. There were no significant differences in either DFS or DSS with regard to HIF-1 α in the short-course RT or the control group. In the multivariate regression model (Cox), the preoperative treatment group was the only independent predictor of DFS ($p=0.007$), whereas disease recurrence ($p=0.015$) and the number of metastatic lymph nodes ≥ 4 ($p=0.0001$) were independent predictors of poor DSS.

5.4. EZRIN (III)

Most preoperative biopsies (80%) but only half of the operative specimens (52%) were negative/weak in ezrin IHC. Ezrin expression in the biopsies and the respective operative samples was significantly different ($\kappa=-0.010$ for four-tier and $\kappa= 0.028$ for dichotomised scoring). Negative expression in 18/22 biopsies (82%) turned positive in the operative sample and positive expression in 6/54 (11%) biopsies turned negative in the operative sample.

Osteosarcoma (U2OS) and glioma (U251) cell lines were tested in WB, to study whether RT directly affects ezrin expression in cancer cells. Radiation increased expression of p53 in U2OS but not in U251 cells, whereas the expression of γ H2A.X, another stress marker, was up-regulated in U251 cells. However, there were no changes in ezrin expression in either cell line.

In survival analysis, two thirds (70%) of the patients with negative/weak ezrin expression in the biopsy were alive without disease, as compared to one third (33 %) of the patients with moderate/strong ezrin expression. In Kaplan-Meier survival analysis, DFS and DSS were significantly longer ($p= 0.027$ and $p= 0.002$), if ezrin expression in the preoperative biopsy sample was

negative/weak as compared to moderate/strong expression. When analysed in samples taken after preoperative treatment, ezrin expression did not correlate with disease outcome. In the multivariate model ezrin expression did not reach statistical significance as a predictor of DFS ($p=0.071$). Disease recurrence ($p= 0.0001$) and the number of metastatic lymph nodes ≥ 4 ($p= 0.028$) were the only independent predictors of poor DSS. In a separate analysis of the long-course RT group, survival of patients, whose tumours showed negative/weak ezrin expression in the operative sample, had better DSS as compared to patients, whose tumours had moderate/strong expression ($p=0.0001$). No statistical differences in disease outcome were seen with regard to ezrin expression in the operative samples of the short-course RT group or the control group.

Negative-weak ezrin expression in the preoperative biopsies was a sensitive predictor of DSS (sensitivity 89.1%), with positive predictive value (PPV) of 86% for the patient being alive. High sensitivity and PPV were lost in the operative samples. The expression of ezrin was further compared with the expression of HIF-1 α and CA IX. Ezrin and HIF-1 α were significantly congruent in the operative samples but not in the preoperative biopsies. Low HIF-1 α expression was associated with negative/weak ezrin expression with OR 4.15 (95% CI, 1.44-11.93; $p=0.004$). Likewise, negative/weak CA IX was linked with negative/weak ezrin with OR=6.42 (95% CI, 3.04-13.56; $p=0.0001$). When the three markers were compared in the preoperative biopsies, no such associations were seen.

When the three markers were analysed together (CA IX and HIF-1 α in the operative samples and ezrin in the preoperative biopsies) and the panel was tested for its longitudinal predictive value of DSS, negative/weak expression of all three markers predicted survival with 31.1% sensitivity, 89.2% specificity, 90.2% PPV and 28.7% NPV. The panel lost its predictive value if ezrin expression in the operative samples was used instead of preoperative biopsies.

5.5. GLUT-1 (IV)

GLUT-1 expression was positive in 68% of the preoperative diagnostic biopsies and in 51% of the corresponding operative samples. Positive GLUT-1 expression of 33/53 (62%) preoperative biopsies remained positive, while that of 20/53 (38%) biopsies turned into negative in the operative specimens. Negative GLUT-1 expression in 7/25 (28%) biopsies turned into positive in the

operative sample, while that of 18/25 (72%) biopsies remained negative ($\kappa=0.301$, CI [0.007; 0.204], ICC= 0.319). Moderate staining intensity in 1/26 (4%) and strong intensity in 0/7 (0%) biopsies turned into negative in the operative specimen. In the short-course RT group, staining intensities in the biopsies and the respective operative samples were significantly different ($p=0.005$), but not in the long-course RT group.

In univariate analysis (KM), there were no significant differences in DFS or DSS with regard to GLUT-1 expression in the biopsies or the operative samples. No significant differences in disease outcome with regard to GLUT-1 expression were found in the short-course RT group or the control group. However, patients, who had received long-course preoperative RT and had negative/weak staining intensity in the operative specimen, had a tendency towards better DFS ($p=0.066$) as compared to patients, who had moderate/strong staining intensity. When CA IX and GLUT-1 expression in the operative samples were analysed together ($n=165$), patients who had tumours with negative/weak staining intensity of both markers had longer DFS than patients who had tumours with moderate/high intensity ($p=0.005$) (unpublished data).

5.6. TUMOUR REGRESSION GRADE WITH REGARD TO IHC (I-IV)

TRG was analysed in the operative tumour samples treated by long-course preoperative RT or chemoradiotherapy. In all, TRG was excellent in 9 tumours and poor/moderate in 36 tumours. TRG with regard to each IHC-analysis is shown in **Table XVI**. In the GLUT-1 study (IV), DFS ($p=0.068$) and DSS ($p=0.024$) of the patients who had excellent tumour regression were longer than those of the patients with moderate or poor response. There was one pCR (1/45, 2%) with no viable tumour cells left after preoperative long-course RT in the excellent response group. There have been no local relapses in the excellent response group, whereas in the moderate/poor response group two patients are alive with and five patients have died of locally recurrent disease.

In a further analysis, the expression of all four markers in the preoperative biopsies was compared to TRG in the operative samples (**Table XVII**). There were no statistically significant differences with regard to IHC of any of the four markers in the biopsies in relation to tumour response. However, no excellent responses in the operative samples were seen, if CA IX intensity in the biopsy was strong ($p=0.092$) (unpublished data).

5.8. COMBINED ANALYSIS OF HIF-1 α , CA IX, EZRIN AND GLUT-1

HIF-1 α (positive/negative), CA IX (negative/weak; moderate/strong intensity), ezrin (negative/weak; moderate/strong) and GLUT-1 (negative/weak; moderate/strong staining intensity) in the operative samples were entered in a multivariate (Cox) model as only variables. In this model, moderate/strong CA IX intensity was a significant independent predictor of poor DSS (HR 2.35; 95% CI 1.167-4.74) ($p=0.017$), but none of the markers predicted DFS (data not shown).

Table XVI. Tumour regression grade with regard to the immunohistochemical (IHC) analysis of HIF-1 α , CA IX, ezrin and GLUT-1 in the operative samples. The number of patients in each publication is included in parentheses.

IHC analysis/ Tumour regression	Staining intensity				Significance
	Negative n (%)	Weak n (%)	Moderate n (%)	Strong n (%)	
CAIX (n=36)					
Poor/Moderate	14 (50)	1 (3)	6 (21)	7 (25)	0.057
Excellent	7 (87)	0 (0)	1 (12)	0 (0)	
HIF-1α (n=39)					
Poor/Moderate	16 (52)	7 (22)	7 (22)	1 (3)	0.413
Excellent	7 (87)	0 (0)	1 (12)	0 (0)	
Ezrin (n=38)					
Poor/Moderate	1 (3)	12 (39)	8 (26)	10 (32)	0.012
Excellent	4 (50)	2 (25)	1 (12)	1 (12)	
GLUT-1 (n=45)					
Poor/Moderate	1 (3)	12 (33)	16 (44)	7 (19)	0.013
Excellent	4 (44)	2 (22)	2 (22)	1 (11)	

Table XVII. Tumour regression grade with regard to immunohistochemical analysis in the preoperative biopsies

Immunohistochemistry	Tumour regression grade		Significance
	Poor/Moderate n (%)	Excellent n (%)	
CA IX intensity (n=24)			
Negative/Weak	11 (61)	7 (39)	0.092
Moderate/Strong	6 (100)	0 (0)	
HIF (n=24)			
Positive	13 (72)	5 (28)	0.586
Negative	4 (67)	2 (33)	
Ezrin (n=23)			
Negative/Weak	7 (58)	5 (42)	0.222
Moderate/Strong	9 (82)	2 (18)	
GLUT-1 intensity (n=23)			
Negative/Weak	10 (53)	9 (47)	0.671
Moderate/Strong	2 (50)	2 (50)	

6. DISCUSSION

In spite of improved disease outcome, a significant proportion of RC patients still succumb to their disease. Consequently, more sophisticated prognostic and predictive markers are needed to direct therapeutic interventions. The aim of this study was to evaluate the ability of three investigational hypoxia-associated markers, HIF-1 α , CA IX and GLUT-1, as well as ezrin, an important protein also associated to HIF-1 α , to predict disease outcome in rectal cancer treated by preoperative RT or chemoradiotherapy.

The IHC staining of HIF-1 α , CA IX, ezrin or GLUT-1 in the operative samples did not correlate with established clinicopathologic factors. One explanation to the lack of correlation could be given preoperative treatment, which may influence tumour size, stage and grade as well as nodal status. Tumour differentiation grade assessed from a biopsy specimen may not be representative of the whole tumour. After long-course RT, the evaluation of tumour differentiation grade may be impossible due to the potential disappearance of most or all of vital tumour^{108, 111}. Preoperative staging can also be challenging^{87, 119}. Currently available imaging methods can over- or underestimate the depth of tumour invasion^{87, 119}. Moreover, there are no reliable methods available as yet to sort out metastatic lymph nodes from benign, and thus, exact preoperative staging is currently challenging¹²⁰.

In order to circumvent this problem, the operative samples were on one hand compared to the pre-treatment, diagnostic biopsies and on the other hand to the operative samples of patients, who had not received preoperative therapy. Our hypothesis was that the expression of the biomarkers in the non-irradiated specimens would resemble their expression in the preoperative biopsies. As shown in **Table XV**, the IHC expression in the biopsies of the preoperative RT group patients deviated from the expression in the control group operative samples. This may partly be explained by the size of the biopsies. Generally multiple biopsies are taken to cover different parts of the tumour and to ensure sufficient tissue material for diagnosis. However, biopsies tend to be taken from the tumour surface, which may be problematic considering the potential heterogeneity of tumours. The differences in patient demographics and the tumour features between the treatment groups also need to be taken into account, keeping in mind the non-randomised retrospective approach of the current study. The control group patients had been too frail or had had too severe comorbidities to have received preoperative treatment, or they had presented with less advanced disease

that had not necessitated preoperative intervention. Control group patients, who had been under-staged preoperatively and who had received postoperative RT or chemoradiotherapy were excluded from this analysis.

6.1. CA IX (I)

The current study showed CA IX staining intensity in the operative specimens to predict adverse disease outcome in RC, which has not been reported previously. CA IX staining intensity was significantly different among the tumours in the different treatment categories, with the greatest proportion of tumours having moderate/strong CA IX intensity in the long-course RT only group. CA IX staining intensity in the operative samples was found to be a remarkable predictive factor, significantly related to both DFS and DSS.

Patients, who had tumours with negative or weakly positive CA IX staining intensity in the operative samples had better DFS and DSS than patients who had tumours with moderately or strongly positive staining intensity. The difference in DFS was evident from early postoperative period and it increased over time to about 50% at 3 years. Similar data have not been published before in RC. No such differences in survival were seen when CA IX intensity was analysed in the preoperative biopsies. In multivariate (Cox) regression model, CA IX intensity in the operative samples and the number of metastatic lymph nodes independently predicted DFS. In a similar analysis, CA IX intensity in the operative samples was the only independent predictor of DSS.

The preoperative treatment groups were then analysed separately. When the long-course RT group was divided into two categories (those with and without concomitant chemotherapy), there was a statistically significant difference in staining intensities in the operative samples between the two groups, showing more intense staining in the tumours of those patients who had not received concomitant chemotherapy. Patients, whose tumours had negative/weak CA IX staining intensity in the operative samples after short-course RT or long-course chemoradiotherapy, had significantly better DFS than patients, whose tumours had moderate/strong CA IX staining intensity (data not shown). DSS was also better for patients whose tumours showed negative/weak CA IX staining intensity in the operative samples after long-course preoperative chemoradiotherapy than for patients, whose tumours had moderate/strong CA IX staining intensity. No significant differences in survival with regard to CA IX intensity in the preoperative biopsies in any of the patient groups or the operative samples after long-course RT

without chemotherapy were seen. The findings of this study may imply that chemotherapy can act as a radiosensitiser, improve tumour oxygenation and consequently, also final treatment outcome. CA9 is known to encode carbonic anhydrase isoenzyme CA IX^{12, 226} and to be the most up-regulated gene in CRC³⁰⁰. CA IX in turn is shown to be up-regulated by HIF-1 α ³⁰¹. Thus, CA IX positivity is presumptuously associated with up-regulated HIF-1 α and further, tumour hypoxia³⁰², as well as unfavourable disease outcome⁶.

Hypoxia-associated markers, including CA IX, are known to be linked with resistance to radiotherapy^{30, 34}. Tumours in the short-course RT group were in general less locally advanced and hence, probably better oxygenated than tumours in the long-course RT group. The locally advanced tumours in the long-course RT group likely harboured hypoxic and radiotherapy resistant areas. After combined chemoradiotherapy, improved disease outcome has previously been reported in RC, malignant glioma and other tumour types as well^{168, 169, 303}. Chemotherapy may thus revoke radioresistance, or act as a radiosensitiser. The radiosensitising effect of chemotherapy could explain the differences between the long-course RT only and the chemoradiotherapy groups in this study. However, the results of these analyses may also be affected by the small number of patients in the long-course RT only group.

The control group patients were also analysed separately. There were no differences in survival between the patients who had or who had not received any RT/chemoradiotherapy and on the other hand, between the patients who had or who had not received any RT/ chemoradiotherapy/adjuvant chemotherapy postoperatively. This may implicate that moderate/strong CA IX intensity rather predicts response to therapy than unfavourable disease outcome per se³⁰⁴. However, the number of patients is low, and thus the analysis may not have enough statistical power to show differences. Also, the control group population deviates from the preoperative treatment group patients with regard to disease stage and patient characteristics. Adjuvant postoperative chemotherapy was likewise given after preoperative treatment when implicated. Moreover, since the current study in addition is retrospective and non-randomised, a direct comparison of the groups may be somewhat questionable.

The data of the current study suggest that CA IX intensity in the operative samples is a powerful independent predictive factor, in line with the findings reported in other human malignancies. Studies have shown CA IX to be associated with adverse disease outcome in non-small cell lung cancer^{290, 294}, bladder²³⁵ and breast cancer²²⁷ as well as oligodendroglioma³⁰⁵. Instead, low

CA IX expression and absence of VHL mutation were found to be related to a more advanced tumour and worse outcome in renal cancer²⁸⁸.

These findings could have important therapeutic implications in the future. Also, CA IX expression might prove to be a useful tool in selecting high-risk patients for adjuvant therapies. In fact, in renal cell cancer, a large placebo-controlled trial is ongoing to define the benefit of CA IX-targeted treatment in the adjuvant setting³⁰⁶. Investigational agent cG250 is a CA IX- specific monoclonal antibody. There are phase I and II trials underway with therapeutic high-dose radiolabelled cG250 and CA IX-loaded dendritic cells in metastatic renal cancer, as well as a large randomised study of cG250 imaging for primary renal cancer³⁰⁶.

In conclusion, CA IX staining intensity in the operative sample is a powerful predictive factor in RC. Negative/weak CA IX staining intensity is associated with favourable DFS and DSS. CA IX may prove to be a useful tool in identifying patients with poor prognosis and thus in need of additional postoperative therapies.

6.2. HIF-1 α (II)

To the best of our knowledge, this is the first study to report HIF-1 α expression in the preoperative biopsies and operative tumour samples in RC treated with preoperative RT. The hypothesis of the study was that preoperative radiotherapy may affect tumour oxygenation and thereby, HIF-1 α expression in the tumour. Indeed, in over 50% of cases, HIF-1 α positive expression in the diagnostic sample turned negative in the operative specimens. Hypoxic tumours are more resistant to therapy and result in worse disease outcome than well oxygenated tumours^{30, 35}. After radiation, reoxygenation is shown to be evident especially in the most poorly oxygenated tumour regions²⁵⁵, probably due to the loss of tumour cells, resulting in improved oxygen diffusion and decreased interstitial pressure²⁵⁵. Improved oxygenation should also lead to the inactivation of HIF-1 α and its down-stream targets, which include VEGF²¹⁷. Mathematical models concerning the effects of antiangiogenic treatments have shown that they likely act by decreasing peritumoural oedema and interstitial pressure, thus normalising the tumour vasculature³⁰⁷. Preoperative radiotherapy may also decrease hypoxia in the tumour and its microenvironment, and thus, the expression of HIF-1 α ¹⁵. However, HIF-1 α is also regulated by other mechanisms, which are independent of the tumour oxygenation status. These mechanisms involve oncogenic stimulation^{211, 308} or defects in HIF-1 α degradation, including VHL

protein mutations²¹⁰. Thus, decreased HIF-1 α expression may also result from other causes than improved tumour oxygenation.

There were no differences with regard to IHC in the preoperative biopsies or the operative samples analysed in the whole patient population. Patients in this study, who were treated with long-course preoperative RT and whose tumours were HIF-1 α negative in the operative specimen, had significantly better DSS than patients whose tumours were HIF-1 α positive. HIF-1 α is recognised as a key mediator in tumour hypoxia^{7-9, 16, 206, 209, 211}. It has a central role in tumour progression as well as in resistance to cancer treatment^{29-31, 309}. This implies that tumours, which remain HIF-1 α positive, evaluated in the operative samples after preoperative treatment, may be resistant to RT. In the short-course RT group, there were no significant differences in survival with regard to HIF-1 α expression in the operative samples. This may reflect the tumours being smaller and probably less hypoxic, and thus the patients having an overall better prognosis than in the long-course RT group^{30, 31, 310}.

In multivariate survival analysis, disease recurrence remained the only independent predictor of DFS, while the preoperative treatment group and the number of metastatic lymph nodes were the only predictors of DSS, HIF-1 α losing its predictive value. Potentially, the size of the long-course RT group was too small to have statistical power in multivariate analysis, or the number of metastatic lymph nodes may exceed the power of other predictors of poor outcome^{1, 311}. There are only two previous trials examining the role of HIF-1 α expression in RC, but patients in neither of these trials had received preoperative RT. Nevertheless, the results of the previous studies showed that high HIF-1 α expression correlated with shorter overall survival^{277, 278}. These findings are in line with the current study and trials with other malignancies^{275, 312} as well.

Our study suggests that RC patients with locally advanced HIF-1 α positive tumours may have an aggressive disease with rapid progression. Thereby, they should probably be followed up closely, regardless of the stage of the tumour. HIF-1 α - analysis could be useful in selecting patients for adjuvant postoperative therapies. Also, HIF-1 α might turn out to be an applicable therapeutic target enhancing the impact of adjuvant treatment on disease outcome¹⁶. In cell line studies, the effect of HIF-1 α inhibition has, however, been shown to be dependent on its sequencing with radiotherapy³¹³. HIF-1 α may on one hand increase radiosensitivity by promoting proliferation, apoptosis, ATP metabolism and p53 activation³¹³; on the other hand HIF-1 α blockade may result in the destruction of tumour vasculature, which in turn

can contribute to radioresistance³¹³. Although tumour cells and their signalling *in vivo* and *in vitro* may be different, these results imply that HIF-1 α blockade may not be unequivocal.

In conclusion, HIF-1 α expression was positive in most of the preoperative biopsies but negative in most of the operative samples, implying that preoperative treatment down-regulates HIF-1 α . Importantly, DSS of the patients who had tumours with negative HIF-1 α expression in the operative samples after long-course radiotherapy had favourable DSS.

6.3. EZRIN (III)

The aim of the current study was to evaluate the possible effect of preoperative treatment on ezrin expression and further, the potential influence of ezrin expression on disease outcome. We are not aware of any previous reports studying ezrin expression in the preoperative biopsies and operative samples after preoperative RT of RC.

Most of the preoperative biopsies but only about half of the operative specimens had negative/weak expression. Thus, more intense ezrin staining was seen after preoperative treatment, which could possibly be induced by radiotherapy. Radiotherapy can shrink the tumour and even complete remissions are seen after preoperative treatment varying between 0-29%^{100, 107, 108, 111, 162}. Normalisation of tumour vasculature³⁰⁷ or loss of tumour cells may improve its oxygenation and down-regulate the expression of various hypoxia markers, e.g. HIF-1 α and CA IX (I, II). Therefore, the relation of ezrin, HIF1- α and CAIX expression was further analysed. The expression of HIF-1 α and ezrin, as well as CAIX and ezrin were significantly concordant in the operative samples, but not in the preoperative biopsies. Ezrin is shown to be linked to mTOR signalling²⁸, which in turn controls the translation of cap-mRNA proteins, including HIF-1 α ¹⁶. It can be postulated that in response to radiation, ezrin expression is up-regulated, inducing also HIF-1 α expression. HIF-1 α is associated with resistance to RT³⁰. Hence, increased ezrin expression due to radiation could bring about resistance to RT³¹⁴.

In this study, in WB, the cell lines responded to radiation by up-regulation of a stress-response marker but this did not affect ezrin expression (III). Ezrin expression in cultured cells, however, may be different from its expression in tissue samples^{20, 241, 247, 315}. In tissues, factors that are not linked to tumour

cells per se can have an effect on how the tissues respond to injury³¹⁶. These factors include the extracellular matrix and various soluble factors, e.g. transforming growth factors, which are known to modulate each other and the effects of injury on e.g. vascular cells³¹⁷. Thereby they can modify the phenotype of epithelial cells^{317, 318} and affect cancer growth and invasion³¹⁶. Thus, tumour environment can modify the response of the tumour to injury caused by radiation and explain the differences seen between cell lines and histological tissue samples.

Ezrin expression in the preoperative biopsies as well as in the surgical specimens of the long-course RT group patients predicted disease outcome in this study. However, this association was not seen in the short-course RT group. It is possible that short-course RT causes rapid changes in tumour microenvironment⁹⁶, which may affect the cellular ezrin levels. During long-course RT or the interval between the end of RT and the operation, tumour environment might restore its balance and thus, re-establish the ezrin levels. Instead, surgery is performed on the following week after short-course RT, and the time interval or the treatment per se, may be too short for the tissues to recover.

Disease outcome of the patients with negative/weak ezrin IHC in the biopsy was significantly better than that of patients with moderate/strong expression. A similar trend was seen for ezrin in the operative specimens, but the difference was not statistically significant. In the long-course RT group, negative or weak ezrin expression in the operative samples was associated with excellent tumour regression and superior DSS. These findings imply that moderate/high ezrin expression in itself predicts adverse disease outcome or is associated with the response of the tumour to preoperative treatment.

Ezrin is a key regulator of HIF-1 α protein synthesis in many cancers¹⁶. Ezrin expression is shown to be related to mTOR pathway²⁸, and its inhibition to significantly reduce tumour cell invasion²⁸⁴. Furthermore, mTOR inhibition is shown to inhibit experimental lung metastasis²⁸. Poor survival in CRC seems to be related to intense ezrin expression²⁴² as well as HIF-1 α expression^{277, 278}. Strong ezrin expression has also been related to adverse disease outcome in epithelial ovarian cancer²⁸⁴, head and neck cancer²⁸³ and uveal malignant melanoma²⁹¹. Thereby, therapies targeting ezrin expression could decrease HIF-1 α expression, which in turn may improve the effect of preoperative therapies and disease outcome in RC. These treatment strategies could include an antibody to or inhibitor of ezrin or possibly its downstream molecules. Indeed, there are preclinical and clinical trials ongoing to study the usefulness of rapamycin and its

analogues²⁸. Temsirolimus and everolimus are mTOR inhibitors currently indicated in the treatment of metastatic renal cell carcinoma.

It can be concluded that ezrin expression in the preoperative, pre-treatment biopsies and the subsequent operative specimens was significantly different, which implies that preoperative treatment modulates ezrin expression in RC. Moderate/strong ezrin expression in the preoperative samples as well as in the operative samples of the long-course RT group was linked to unfavourable disease outcome. In the multivariate analysis, however, ezrin did not reach statistical significance as an independent predictor of disease outcome. Ezrin is a promising biomarker that is associated with mTOR pathway signalling. The concept of mTOR inhibition is already exploited in some cancers.

6.4. GLUT-1 (IV)

Our hypothesis was that preoperative treatment shrinks the tumour and alters its microenvironment, which leads to improved tumour oxygenation and a change in GLUT-1 expression. There are no previous reports comparing GLUT-1 expression in the diagnostic biopsies and operative samples after preoperative RT.

In this study, GLUT-1 staining in the preoperative biopsies and the corresponding operative samples was different in that two thirds of the biopsies but only half of the corresponding operative samples were GLUT-1 positive, reflecting a change in GLUT-1 expression in the tumour during the preoperative period. In an earlier study, surgical procedure was associated with significantly increased GLUT-1 levels in RC³¹⁹, when biopsies were taken just before the operation under anaesthesia, which may also have some effect on the results. In contrast, in the current study, GLUT-1 expression levels decreased when pre-RT biopsies were compared to the samples taken at operation. This implies that preoperative treatment interferes with GLUT-1 expression in the tumour. RT destroys tumour cells, leading to down-staging, down-sizing or even complete tumour elimination¹⁰⁸. Importantly, RT can also have an effect on the tumour microenvironment, especially the tumour vasculature^{96, 316, 317}. Thus, it is not unexpected that preoperative treatment seems to have an effect on oxygenation and hence, also tumour metabolism.

In our study, there were no significant differences in DFS or DSS with regard to GLUT-1 expression in the biopsies or the operative samples. However, patients, who had received long-course preoperative RT and had

negative/weak staining intensity in the operative specimens, had a trend towards better DFS as compared to patients, who had moderate/strong staining intensity. This is in parallel to the findings of a previous study, showing an association of increased GLUT-1 expression in the pre-treatment biopsies with poorer response to chemoradiotherapy²⁷⁹. High GLUT-1 expression is shown to be associated with poor prognosis in other malignancies as well^{280, 293}. In a combined analysis with CA IX, negative/weak intensity of both markers in the operative samples was associated with longer DFS, further emphasising the power of CA IX in predicting disease outcome in rectal cancer. The expressions of CA IX and GLUT-1 have earlier been shown to correlate with each other and to be interchangeable prognostic markers in bladder cancer²⁹⁴. However, to the best of our knowledge there are no previous reports of the combined expression of these markers with regard to survival in rectal cancer.

Malignant cells have a tendency to turn to anaerobic glycolysis as a source of energy even in well-oxygenated conditions²⁰³ and an association between GLUT-1 expression and chemoresistance has been shown²³⁶. Thus, the inhibition of glycolysis could be a useful approach for therapeutic purposes in cancer³²⁰. Tyrosine kinase inhibitors with specificity for ATP binding sites are shown to directly inhibit GLUT-1³²¹. GLUT-1 antibodies have been tested in cancer cell lines and they have been shown to inhibit proliferation and to induce apoptosis³²².

Taken together, there was a change in the tumour's GLUT-1 expression during the preoperative period, since GLUT-1 expression was positive in two thirds of the preoperative biopsies but only half of the operative samples. Patients of the long-course RT group with negative/weak GLUT-1 expression in the operative sample had better DFS, as compared to patients with moderate/strong expression, but the difference did not reach statistical significance.

6.5. TUMOUR REGRESSION GRADE (I-IV)

Tumour regression grade was analysed in the long-course RT group operative samples. Good tumour regression was non-significantly linked to negative/weak HIF-1 α and CA IX, and significantly to GLUT-1 and ezrin expression. Excellent TRG was significantly associated with favourable DFS and DSS. In the study population, there was only one pCR with no viable tumour cells left. Thus, the rate of pCR was less than previously reported^{100, 107, 111, 162}.

The rate of pCR depends on the strictness of the criteria and the analysis of sufficient tissue material, including the mural tumour and the lymph nodes. In

the current study, the criteria for complete pathologic regression were set tight and several slides were studied to ensure the complete disappearance of vital cancer in the tumour and lymph nodes as well. Time interval between the end of RT and surgery is shown to be the single most important factor affecting pCR¹⁶⁴. The effects of RT are not visible shortly after its completion. Non-viable cancer cells may appear morphologically intact, before they undergo apoptosis¹⁶³. Extending the interval beyond 12 weeks does not appear to increase pCR since most responses are shown to occur during 8-10 weeks after RT¹⁶⁴. The controversy between the findings of the current study and most of the previously reported trials may partly be explained by the deviating time interval. In this study, waiting time varied between 4-10 weeks, being shorter among some patients who were operated at the start and longer towards the end of patient accrual.

The expression of all four markers in the preoperative biopsies was also compared with TRG in the operative samples after long-course RT, showing no significant differences between response to therapy and IHC. However, no excellent responses were seen if CA IX intensity in the preoperative biopsy was strong. Taken together, in this study, IHC in the biopsies was not a significant predictor of good response to chemoradiotherapy. This may be explained by the small number of patients, for whom biopsies were available, but the relatively small size of the biopsy specimens can also have an influence on the results.

Excellent response, with only a small remaining proportion of possibly vital cancer, was associated with favourable disease outcome. Determining complete pathologic response is challenging and analysing the degree (poor/moderate or excellent) of TRG may be helpful in estimating disease outcome and weighing the possible benefits of additional postoperative therapies to optimise prognosis. Sufficient time interval between the end of RT and surgery is important to achieve pCR.

6.6. COMBINED ANALYSIS OF HIF-1 α , CA IX, EZRIN AND GLUT-1

All four markers, HIF-1 α , CA IX, ezrin and GLUT-1, were tested in multivariate analysis, where CA IX intensity proved to be the only independent predictor of DSS. Albeit preliminary, this multivariate analysis suggests that CA IX indeed is a powerful predictive factor in RC. Further analyses are planned in the future to investigate the potential role of all these markers or their various combinations in predicting disease outcome in RC, using models where classical prognostic factors are included as well.

7. SUMMARY AND CONCLUSIONS

The present study concerning rectal cancer was focused on evaluating the predictive significance of hypoxia-related molecular markers HIF-1 α , CA IX and GLUT-1 as well as ezrin, which is also linked to these markers. The following conclusions can be drawn from the results of this study:

1) CA IX staining intensity is a powerful predictor of disease outcome. In the present study, moderate/strong CA IX staining intensity in the operative samples was related to unfavourable DFS and DSS in rectal cancer. In multivariate analysis, CA IX intensity in the operative samples was an independent predictor of DFS. The risk for dying of rectal cancer was 9.2-fold if moderate/strong CA IX intensity was present.

2) HIF-1 α expression predicts disease outcome in rectal cancer after long-course RT. In the present study, positive HIF-1 α expression in the operative samples after long-course RT was associated with unfavourable DSS in rectal cancer.

3) Ezrin expression in the diagnostic pre-treatment biopsies is associated with disease outcome in rectal cancer. In this study, negative/weak ezrin expression in the biopsies was related to longer DFS and DSS. In the long-course RT group, DSS of the patients with negative/weak ezrin expression in the operative samples was also significantly better than of those patients with moderate/strong ezrin expression.

4) GLUT-1 expression is not a powerful predictive factor in rectal cancer. In the current study, patients with negative/weak GLUT-1 staining intensity in the operative specimen after long-course RT had longer DFS, as compared to patients with moderate/strong staining intensity, but the difference was not statistically significant.

5) Excellent tumour regression after preoperative long-course RT with or without chemotherapy was linked to negative or weak expression of CA IX, ezrin and GLUT-1 in the operative samples. Excellent TRG was associated with favourable DFS and DSS.

8. ACKNOWLEDGEMENTS

The studies for this thesis were carried out at the Departments of Pathology and Oncology in Turku University Hospital during the years 2006-2010. I express my deepest gratitude to Professor Seppo Pyrhönen, my supervisor, and Professor Olli Carpén, my co-writer and monitor, for their scientific visions as well as instructive comments and opinions, and for the opportunity to explore the scientific world. I am extremely grateful to my supervisor Docent Jari Sundström, for his dedication to my research, as well as for valuable guidance, advice and encouragement during these years. Jari introduced me to the world of immunohistochemistry and steered me through the jungles of joy and frustration in research. I am extremely thankful to Docent Panu Jaakkola, MD, PhD, for his ideas, opinions and expertise in the field of hypoxia. I am also grateful to the reviewers for their extensive knowledge and outstanding comments and suggestions to further improve this thesis.

I express my warmest gratitude to Professor Kari Syrjänen, for his skilful help in statistics, to Professor Antti Vaheri for providing ezrin antibody and to Docent Tove Grönroos, for providing GLUT-1 antibody for this research. I am thankful for having had the opportunity of working with my appreciated co-workers and co-writers Professor Heikki Minn, MD, PhD, Kati Talvinen, PhD, Docent Riyad Bendardaf, MD, PhD, and Minja Laulajainen, MSc. I am thankful to Docent Raija Ristamäki, MD, PhD, for her knowledge and skills in the field of gastrointestinal malignancies, as well as for the unforgettable opera experiences we have shared during the past few years. I am grateful to Docent Abdelbaset Buhmeida, MD, PhD, and Dr. Adam Elzagheid, MD, PhD, for collaboration, as well as Dr. Annika Ålgars, MD, and Dr. Tuulia Avoranta, MD, for inspiring conversations in the hospital corridors or over lunch during this work.

The skilful expert help of Mr. Jaakko Liippo and Mrs. Sinikka Collanus as well as researcher Arun Pokhrel, PhD, at Finnish Cancer Registry is greatly appreciated. I am also indebted to Docent Paula Lindholm, MD, PhD, Dr. Kirsti Dean, MD, and physicist Sami Suilamo, MSc as well as colleagues from the Surgical hospital, especially Drs. Erkki Pekkala, MD, PhD, and Heikki Sarparanta, MD, for collaboration.

I am grateful to all my colleagues and co-workers at the Departments of Oncology and Pathology, particularly my past and present room-mates Docent Riikka Huovinen, MD, PhD, Docent Paula Lindholm, MD, PhD and Dr. Outi Hirvonen, MD, PhD, as well as all the colleagues at the departments of surgery and radiology.

I wish to thank my friends and members of the former Banana Therapy Section: Dr. Ritva Valavaara, MD, PhD, Docent Sirkku Jyrkkiö, MD, PhD, Ministerial counsellor Raija Asola, MD, and Dr. Anne Suovuori, MD, for friendship and countless moments of supervision of work and life. I also wish to express my sincere gratitude to my former and current jogging teams: Docents Sirkku Jyrkkiö, MD, PhD, Pia Vihinen, MD, PhD,

Acknowledgements

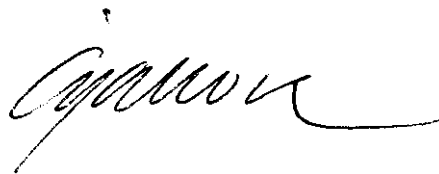
Paula Lindholm, MD, PhD, Drs. Maria Lapela, MD, PhD, Outi Hirvonen, MD, PhD, Marjaana Luukkaa, MD, PhD, Johanna Ruohola, MD, PhD, Kaisa Lehtiö MD, PhD, and Minna Rinne MD. I am thankful for having had a pleasure to drive a snowmobile and ride a reindeer, with my colleague and friend Dr. Anna-Liisa Kautio, MD, and to hike over the Pyrenees with my Santiago de Compostela team: Marita Talvenmaa, RN, Ritva Mylly, RN and Pirjo Nieminen, PT.

I wish to express my gratitude to my deceased parents, to my sister Ulla-Marja and her family, to my parents-in-law Jaakko and Marja-Leena, my siblings-in-law Minna and Jyrki and their families, and my godchildren Nella, Jonas, Simo and Laura. I am thankful to my aunt Seija and her husband Tero for their continued support during the ups and downs of life. My friends Merja, Mervi, Iivi, Sinikka and Kari, Rosana, Kim and Clyde, Agnieszka and Lauri are greatly appreciated for their patience and support and Päivi for inspiring theatre experiences.

I am extremely thankful to my number one team, Team Finland, my family, to whom I dedicate this thesis. I wish to express my love as well as my warmest and deepest gratitude to my very best friend and hiking buddy, my beloved husband Petri and my dear and talented daughters Karoliina, Petra and Tiltu. I am indebted to my loved ones for their unfailing patience, love, support, company and great sense of humour as well as for the tremendous creative atmosphere, which has surrounded me at home. This research work would not have been meaningful without my family. I am thankful for the two awesome young gentlemen, Atte and Ilkka, who have joined our family and bring happiness and joy to our daughters, and the parents alike. I endeavour the memories of Buster, for patience as well as for lessons of love and loyalty taught during the past 13 years.

This research work has been supported by grants from the Special Government Funding (EVO) allocated to Turku University; Finnish Society for Therapeutic Radiology and Oncology; Finnish Cultural Foundation, Varsinais-Suomi Regional Fund; Cancer Society of Finland; Southwest Finland Cancer Society; Turku Graduate School of Clinical Sciences; Lilly Foundation and Maud Kuistila Foundation.

Turku, April 2011,



9. REFERENCES

1. Swanson R, Compton C, Stewart A and Bland K. The prognosis of T3N0 colon cancer is dependent on the number of lymph nodes examined. *Ann Surg Oncol* 2003; 10:65-71.
2. Nagtegaal I and Quirke P. What is the role for the circumferential margin in the modern treatment of rectal cancer? *J Clin Oncol* 2008; 26:303-12.
3. Swedish Rectal Cancer Trial. Improved survival with preoperative radiotherapy in resectable rectal cancer. *N Engl J Med* 1997; 336: 980-7.
4. Kapiteijn E, Marijnen C, Nagtegaal I, Putter H, Steup W, Wiggers T, Rutten H, Pahlman L, Glimelius B, van Krieken J, Leer J and van de Velde C. Preoperative radiotherapy combined with total mesorectal excision for resectable rectal cancer. *N Engl J Med* 2001; 345:638-46.
5. Glimelius B, Pålman L, Cervantes A for ESMO Guidelines Working Group. Rectal cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 2010; 21 Suppl 5:v82-6.
6. Höckel M and Vaupel P. Tumor hypoxia: definitions and current clinical, biologic, and molecular aspects. *J Natl Cancer Inst* 2001; 93: 266-76.
7. Harris A. Hypoxia--a key regulatory factor in tumour growth. *Nat Rev Cancer* 2002; 2:38-47.
8. Wang G and Semenza G. General involvement of hypoxia-inducible factor 1 in transcriptional response to hypoxia. *Proc Natl Acad Sci U S A* 1993; 90:4304-8.
9. Semenza G. Hypoxia, clonal selection, and the role of HIF-1 in tumor progression. *Crit Rev Biochem Mol Biol* 2000; 35:71-103.
10. Semenza G. Targeting HIF-1 for cancer therapy. *Nat Rev Cancer* 2003; 3:721-32.
11. Kivelä A, Saarnio J, Karttunen T, Kivelä J, Parkkila A, Pastorekova S, Pastorek J, Waheed A, Sly W, Parkkila T and Rajaniemi H. Differential expression of cytoplasmic carbonic anhydrases, CA I and II, and membrane-associated isozymes, CA IX and XII, in normal mucosa of large intestine and in colorectal tumors. *Dig Dis Sci* 2001; 46:2179-86.
12. Niemelä A, Hynninen P, Mecklin J, Kuopio T, Kokko A, Aaltonen L, Parkkila A, Pastorekova S, Pastorek J, Waheed A, Sly W, Ørntoft T, Kruhøffer M, Haapasalo H, Parkkila S and Kivelä A. Carbonic anhydrase IX is highly expressed in hereditary nonpolyposis colorectal cancer. *Cancer Epidemiol Biomarkers Prev* 2007; 16:1760-6.
13. Wang G and Semenza G. Molecular basis of hypoxia-induced erythropoietin expression. *Curr Opin Hematol* 1996; 3: 156-62.
14. Wang G and Semenza G. Characterization of hypoxia-inducible factor 1 and regulation of DNA binding activity by hypoxia. *J Biol Chem* 1993; 268: 21513-8.
15. Semenza G, Roth P, Fang H, Wang G. Transcriptional regulation of genes encoding glycolytic enzymes by hypoxia-inducible factor 1. *J Biol Chem* 1994; 269: 23757-63.
16. Semenza G. Evaluation of HIF-1 inhibitors as anticancer agents. *Drug Discov Today* 2007; 12:853-9.
17. Hudson C, Liu M, Chiang G, Otterness D, Loomis D, Kaper F, Giaccia A and Abraham R. Regulation of hypoxia-inducible factor 1 α expression and function by the mammalian target of rapamycin. *Mol Cell Biol* 2002; 22:7004-14.

References

18. Bretscher A, Reczek D and Berryman M. Ezrin: a protein requiring conformational activation to link microfilaments to the plasma membrane in the assembly of cell surface structures. *J Cell Sci* 1997; 110 (Pt 24):3011-8.
19. Pakkanen R. Immunofluorescent and immunochemical evidence for the expression of cytovillin in the microvilli of a wide range of cultured human cells. *J Cell Biochem* 1988; 38: 65-75.
20. Tsukita S, Oishi K, Sato N, Sagara J and Kawai A. ERM family members as molecular linkers between the cell surface glycoprotein CD44 and actin-based cytoskeletons. *J Cell Biol* 1994; 126: 391-401.
21. Algrain M, Turunen O, Vaehri A, Louvard D and Arpin M. Ezrin contains cytoskeleton and membrane binding domains accounting for its proposed role as a membrane-cytoskeletal linker. *J Cell Biol* 1993;120: 129-39.
22. Arpin M, Algrain M and Louvard D. Membrane-actin microfilament connections: an increasing diversity of players related to band 4.1. *Curr Opin Cell Biol* 1994;6: 136-41.
23. Vaehri A, Carpén O, Heiska L, Helander T, Jääskeläinen J, Majander-Nordenswan P, Sainio M, Timonen T and Turunen O. The ezrin protein family: membrane-cytoskeleton interactions and disease associations. *Curr Opin Cell Biol* 1997;9:659-66.
24. Brambilla D and Fais S. The Janus-faced role of ezrin in "linking" cells to either normal or metastatic phenotype. *Int J Cancer* 2009; 125:2239-45.
25. Niggli V and Rossy J. Ezrin/radixin/moesin: versatile controllers of signaling molecules and of the cortical cytoskeleton. *Int J Biochem Cell Biol* 2008; 40:344-9.
26. Fehon R, McClatchey A and Bretscher A. Organizing the cell cortex: the role of ERM proteins. *Nat Rev Mol Cell Biol* 2010; 11:276-87.
27. Khanna C, Wan X, Bose S, Cassaday R, Olomu O, Mendoza A, Yeung C, Gorlick R, Hewitt S and Helman L. The membrane-cytoskeleton linker ezrin is necessary for osteosarcoma metastasis. *Nat Med* 2004; 10: 182-6.
28. Wan X, Mendoza A, Khanna C and Helman L. Rapamycin inhibits ezrin-mediated metastatic behavior in a murine model of osteosarcoma. *Cancer Res* 2005; 65:2406-11.
29. Höckel M, Schlenger K, Aral B, Mitze M, Schäffer U and Vaupel P. Association between tumor hypoxia and malignant progression in advanced cancer of the uterine cervix. *Cancer Res* 1996; 56: 4509-15.
30. Höckel M, Schlenger K, Mitze M, Schäffer U and Vaupel P. Hypoxia and Radiation Response in Human Tumors. *Semin Radiat Oncol* 1996; 6: 3-9.
31. Höckel M, Schlenger K, Höckel S and Vaupel P. Hypoxic cervical cancers with low apoptotic index are highly aggressive. *Cancer Res* 1999; 59: 4525-8.
32. Jokilehto T, Rantanen K, Luukka M, Heikkinen P, Grenman R, Minn H, Kronqvist P and Jaakkola P. Overexpression and nuclear translocation of hypoxia-inducible factor prolyl hydroxylase PHD2 in head and neck squamous cell carcinoma is associated with tumor aggressiveness. *Clin Cancer Res* 2006; 12:1080-7.
33. Sauer R, Becker H, Hohenberger W, Rödel C, Wittekind C, Fietkau R, Martus P, Tschmelitsch J, Hager E, Hess C, Karstens J, Liersch T, Schmidberger H and Raab R. Preoperative versus postoperative chemoradiotherapy for rectal cancer. *N Engl J Med* 2004; 351:1731-40.

References

34. Koukourakis M, Giatromanolaki A, Polychronidis A, Simopoulos C, Gatter K, Harris A and Sivridis E. Endogenous markers of hypoxia/anaerobic metabolism and anemia in primary colorectal cancer. *Cancer Sci* 2006; 97:582-8.
35. Ravizza R, Molteni R, Gariboldi M, Marras E, Perletti G and Monti E. Effect of HIF-1 modulation on the response of two- and three-dimensional cultures of human colon cancer cells to 5-fluorouracil. *Eur J Cancer* 2009; 45:890-898.
36. Parkin D, Bray F, Ferlay J and Pisani P. Global cancer statistics, 2002. *CA Cancer J Clin*; 55:74-108.
37. Jemal A, Siegel R, Ward E, Hao Y, Xu J and Thun M. Cancer statistics, 2009. *CA Cancer J Clin*; 59:225-49.
38. Ferlay J, Autier P, Boniol M, Heanue M, Colombet M and Boyle P. Estimates of the cancer incidence and mortality in Europe in 2006. *Ann Oncol* 2007; 18:581-92.
39. Finnish Cancer Registry. Cancer Statistics, Finland. <http://www.cancerregistry.fi/eng/statistics/AID122.html>, 3.2.2010 (23.11.2010).
40. Ballinger A and Anggiansah C. Colorectal cancer. *BMJ* 2007; 335:715-8.
41. Alhopuro P, Ylisaukko-Oja S, Koskinen W, Bono P, Arola J, Järvinen H, Mecklin J, Atula T, Kontio R, Mäkitie A, Suominen S, Leivo I, Vahteristo P and Aaltonen L. The MDM2 promoter polymorphism SNP309T-->G and the risk of uterine leiomyosarcoma, colorectal cancer, and squamous cell carcinoma of the head and neck. *J Med Genet* 2005; 42:694-8.
42. Hanahan D and Weinberg R. The hallmarks of cancer. *Cell* 2000; 100:57-70.
43. Medema RH and Bos JL. The role of p21ras in receptor tyrosine kinase signaling. *Crit Rev Oncog* 1993; 4: 615-61.
44. Moses HL, Yang EY and Pietenpol JA. TGF-beta stimulation and inhibition of cell proliferation: new mechanistic insights. *Cell* 1990; 63: 245-7.
45. Giancotti FG and Ruoslahti E. Integrin signaling. *Science* 1999; 285: 1028-32.
46. Fearon E and Vogelstein B. A genetic model for colorectal tumorigenesis. *Cell* 1990; 61:759-67.
47. Hill M, Morson B and Bussey H. Aetiology of adenoma--carcinoma sequence in large bowel. *Lancet* 1978; 1:245-7.
48. Goel A, Arnold CN, Niedzwiecki D, Chang DK, Ricciardiello L, Carethers JM, Dowell JM, Wasserman L, Compton C, Mayer RJ, Bertagnolli MM and Boland CR. Characterization of sporadic colon cancer by patterns of genomic instability. *Cancer Res* 2003; 63:1608-14.
49. Laiho P, Kokko A, Vanharanta S, Salovaara R, Sammalkorpi H, Järvinen H, Mecklin J, Karttunen T, Tuppurainen K, Davalos V, Schwartz SJ, Arango D, Mäkinen MJ and Aaltonen LA. Serrated carcinomas form a subclass of colorectal cancer with distinct molecular basis. *Oncogene* 2007; 26:312-20.
50. Mäkinen M. Colorectal serrated adenocarcinoma. *Histopathology* 2007; 50:131-50.
51. Mäkinen M, George S, Jernvall P, Mäkelä J, Vihko P and Karttunen T. Colorectal carcinoma associated with serrated adenoma--prevalence, histological features, and prognosis. *J Pathol* 2001; 193:286-94.

References

52. Bendardaf R, Lamlum H, Ristamäki R, Korkeila E, Syrjänen K and Pyrhönen S. Thymidylate synthase and microsatellite instability in colorectal cancer: implications for disease free survival, treatment response and survival with metastases. *Acta Oncol* 2008; 47:1046-53.
53. Vasen H, Möslein G, Alonso A, Aretz S, Bernstein I, Bertario L, Blanco I, Bulow S, Burn J, Capella G, Colas C, Engel C, Frayling I, Rahner N, Hes FJ, Hodgson S, Mecklin JP, Møller P, Myrthøj T, Nagengast FM, Parc Y, Ponz de Leon M, Renkonen-Sinisalo L, Sampson JR, Stromorken A, Tejpar HJ, Wijnen J, Lubinski J, Järvinen H, Claes E, Heinimann K, Karagiannis JA Lindblom A, Dove-Edwin I and Müller H. Recommendations to improve identification of hereditary and familial colorectal cancer in Europe. *Fam Cancer* 2010; 9:109-15.
54. Lynch P. Standards of care in diagnosis and testing for hereditary colon cancer. *Fam Cancer* 2008; 7:65-72.
55. St John D, McDermott F, Hopper J, Debney E, Johnson W and Hughes E. Cancer risk in relatives of patients with common colorectal cancer. *Ann Intern Med* 1993; 118:785-90.
56. de la Chapelle A. Genetic predisposition to colorectal cancer. *Nat Rev Cancer* 2004; 4:769-80.
57. Mecklin J, Aarnio M, Läärä E, Kairaluoma M, Pylvänäinen K, Peltomäki P, Aaltonen L and Järvinen H. Development of colorectal tumors in colonoscopic surveillance in Lynch syndrome. *Gastroenterology* 2007; 133:1093-8.
58. Mecklin J. The implications of genetics in colorectal cancer. *Ann Oncol* 2008; 19 Suppl 5:v87-90.
59. Balmaña J, Castells A and Cervantes A for ESMO Guidelines Working Group. Familial colorectal cancer risk: ESMO Clinical Practice Guidelines. *Ann Oncol* 2010; 21 Suppl 5:v78-81.
60. Aarnio M, Sankila R, Pukkala E, Salovaara R, Aaltonen L, de la Chapelle A, Peltomäki P, Mecklin J and Järvinen H. Cancer risk in mutation carriers of DNA-mismatch-repair genes. *Int J Cancer* 1999; 81:214-8.
61. Vasen H, Watson P, Mecklin J and Lynch H. New clinical criteria for hereditary nonpolyposis colorectal cancer (HNPCC, Lynch syndrome) proposed by the International Collaborative group on HNPCC. *Gastroenterology* 1999; 116:1453-6.
62. van Staa T, Card T, Logan R and Leufkens H. 5-Aminosalicylate use and colorectal cancer risk in inflammatory bowel disease: a large epidemiological study. *Gut* 2005; 54:1573-8.
63. Baxter N, Tepper J, Durham S, Rothenberger D and Virnig B. Increased risk of rectal cancer after prostate radiation: a population-based study. *Gastroenterology* 2005; 128:819-24.
64. Pischon T, Lahmann P, Boeing H, Friedenreich C, Norat T, Tjønneland A, Halkjaer J, Overvad K, Clavel-Chapelon F, Boutron-Ruault M, Guernec G, Bergmann M, Linseisen J, Becker N, Trichopoulou A, Trichopoulos D, Sieri S, Palli D, Tumino R, Vineis P, Panico S, Peeters PH, Bueno de Mesquita HB, Boschuizen HC, Van Guelpen B, Palmqvist R, Berglund G, Gonzalez CA, Dorronsoro M, Barricarte A, Navarro C, Martinez C, Quiros JR, Roddam A, Allen N, Bingham S, Khaw KT, Ferrari P, Kaaks R, Slimani N and Riboli E. Body size and risk of colon and rectal cancer in the European Prospective Investigation into Cancer and Nutrition (EPIC). *J Natl Cancer Inst* 2006; 98:920-31.
65. Harriss D, Atkinson G, Batterham A, George K, Cable N, Reilly T, Haboubi N and Renehan A. Lifestyle factors and colorectal cancer risk (2): a systematic review and meta-analysis of associations with leisure-time physical activity. *Colorectal Dis* 2009; 11:689-701.
66. Norat T, Bingham S, Ferrari P, Slimani N, Jenab M, Mazuir M, Overvad K, Olsen A, Tjønneland A, Clavel F, Boutron-Ruault M, Kesse E, . Meat, fish, and colorectal cancer risk: the European Prospective Investigation into cancer and nutrition. *J Natl Cancer Inst* 2005; 97:906-16.

References

67. Cho E, Smith-Warner S, Ritz J, van den Brandt P, Colditz G, Folsom A, Freudenheim J, Giovannucci E, Goldbohm R, Graham S, Holmberg L, Kim D, Malila N, Miller AB, Pietinen P, Rohan TE, Sellers TA, Speizer FE, Willet WC, Wolk A and Hunter DJ. Alcohol intake and colorectal cancer: a pooled analysis of 8 cohort studies. *Ann Intern Med* 2004; 140:603-13.
68. Malila N, Virtamo J, Virtanen M, Pietinen P, Albanes D and Teppo L. Dietary and serum alpha-tocopherol, beta-carotene and retinol, and risk for colorectal cancer in male smokers. *Eur J Clin Nutr* 2002; 56:615-21.
69. Thompson C, Larkin E, Patel S, Berger N, Redline S and Li L. Short duration of sleep increases risk of colorectal adenoma. *Cancer* 2011; 117:657-878.
70. Fernandez E, La Vecchia C, Balducci A, Chatenoud L, Franceschi S and Negri E. Oral contraceptives and colorectal cancer risk: a meta-analysis. *Br J Cancer* 2001; 84:722-7.
71. Sandler R, Halabi S, Baron J, Budinger S, Paskett E, Keresztes R, Petrelli N, Pipas J, Karp D, Loprinzi C, Steinbach G and Schilsky R. A randomized trial of aspirin to prevent colorectal adenomas in patients with previous colorectal cancer. *N Engl J Med* 2003; 348:883-90.
72. Pour P and Kazakoff K. Prevention of rectal cancer induced by N-nitrosobis(2-oxopropyl)-amine by exogenous testosterone in MRC rats. *Carcinogenesis* 1989; 10:2015-7.
73. Huynh D, Vincan E, Mantamadiotis T, Purcell D, Chan C and Ramsay R. Oncogenic properties of HIV-Tat in colorectal cancer cells. *Curr HIV Res* 2007; 5:403-9.
74. Damin D, Caetano M, Rosito M, Schwartzmann G, Damin A, Frazzon A, Ruppenthal R and Alexandre C. Evidence for an association of human papillomavirus infection and colorectal cancer. *Eur J Surg Oncol* 2007; 33:569-74.
75. Cuevas-Ramos G, Petit CR, Marcq I, Boury M, Oswald E and Nougayrède JP. *Escherichia coli* induces DNA damage in vivo and triggers genomic instability in mammalian cells. *Proc Natl Acad Sci U S A* 2010; 107:11537-42.
76. Kellokumpu I, Andersson L and Kellokumpu S. Detection of colorectal neoplasia with peanut-agglutinin (PNA)-reactive carbohydrate structures in rectal mucus. *Int J Cancer* 1997; 74:648-53.
77. Vironen J, Kellokumpu S, Andersson L and Kellokumpu I. Comparison of a peanut agglutinin test and an immunochemical faecal occult blood test in detecting colorectal neoplasia in symptomatic patients. *Scand J Clin Lab Invest* 2004; 64:140-5.
78. Towler B, Irwig L, Glasziou P, Kewenter J, Weller D and Silagy C. A systematic review of the effects of screening for colorectal cancer using the faecal occult blood test, hemoccult. *BMJ* 1998; 317:559-65.
79. Finnish Cancer Registry. Screening. <http://www.cancerregistry.fi/eng/screening> 17.12.2008 (23.11.2010)
80. Paimela H, Malila N, Palva T, Hakulinen T, Vertio H and Järvinen H. Early detection of colorectal cancer with faecal occult blood test screening. *Br J Surg* 2010; 97:1567-71.
81. Odze RD GJ. *Surgical Pathology of the GI Tract, Liver, Biliary Tract and Pancreas*, 2nd edition, Saunders Elsevier; Philadelphia, PA, 2009, page 605.
82. Hamilton SR, Bosman FT, Boffetta P, Ilyas M, Morreau H, Nakamura SI, Quirke P, Riboli E and Sobin LH. Carcinoma of the colon and rectum *in* Bosman FT, Carneiro F, Hruban RH and Theise ND (editors). *WHO Classification of Tumours of the Digestive system*. 3rd volume, 4th edition, IARC Press; Lyon, 2010, pages 134-146.

References

83. Ahmad A, Jamieson T, Balsitis M and Diament R. Radiation-induced angiosarcoma of the rectum: a case report and review of literature. *Colorectal Dis* 2008; 10:847-8.
84. Fratesi L, Alhusayen R and Walker J. Case report of primary rectal melanoma and review of the etiology of melanoma. *J Cutan Med Surg*; 12:117-20.
85. Gelas T, Peyrat P, Francois Y, Gerard J, Baulieux J, Gilly F, Vignal J and Glehen O. Primary squamous-cell carcinoma of the rectum: report of six cases and review of the literature. *Dis Colon Rectum* 2002; 45:1535-40.
86. Scherübl H. Rectal carcinoids are on the rise: early detection by screening endoscopy. *Endoscopy* 2009; 41:162-5.
87. Valentini V, Beets-Tan R, Borrás J, Krivokapić Z, Leer J, Pålman L, Rödel C, Schmoll H, Scott N, Velde C and Verfaillie C. Evidence and research in rectal cancer. *Radiother Oncol* 2008; 87:449-74.
88. Wasserberg N and Gutman H. Resection margins in modern rectal cancer surgery. *J Surg Oncol* 2008; 98:611-5.
89. Nagtegaal I, Marijnen C, Kranenbarg E, van de Velde C and van Krieken J. Circumferential margin involvement is still an important predictor of local recurrence in rectal carcinoma: not one millimeter but two millimeters is the limit. *Am J Surg Pathol* 2002; 26:350-7.
90. Nagtegaal I, van de Velde C, Marijnen C, van Krieken J and Quirke P. Low rectal cancer: a call for a change of approach in abdominoperineal resection. *J Clin Oncol* 2005; 23:9257-64.
91. Marr R, Birbeck K, Garvican J, Macklin C, Tiffin N, Parsons W, Dixon M, Mapstone N, Sebag-Montefiore D, Scott N, Johnston D, Sagar P, Finan P and Quirke P. The modern abdominoperineal excision: the next challenge after total mesorectal excision. *Ann Surg* 2005; 242:74-82.
92. Marijnen C, Nagtegaal I, Klein Kranenbarg E, Hermans J, van de Velde C, Leer J and van Krieken J. No downstaging after short-term preoperative radiotherapy in rectal cancer patients. *J Clin Oncol* 2001; 19:1976-84.
93. Baxter N, Morris A, Rothenberger D and Tepper J. Impact of preoperative radiation for rectal cancer on subsequent lymph node evaluation: a population-based analysis. *Int J Radiat Oncol Biol Phys* 2005; 61:426-31.
94. Taflampas P, Christodoulakis M, Gourtsoyianni S, Leventi K, Melissas J and Tsiftsis D. The effect of preoperative chemoradiotherapy on lymph node harvest after total mesorectal excision for rectal cancer. *Dis Colon Rectum* 2009; 52:1470-4.
95. Joseph N, Sigurdson E, Hanlon A, Wang H, Mayer R, MacDonald J, Catalano P and Haller D. Accuracy of determining nodal negativity in colorectal cancer on the basis of the number of nodes retrieved on resection. *Ann Surg Oncol* 2003; 10:213-8.
96. Lorimore S, Coates P, Scobie G, Milne G and Wright E. Inflammatory-type responses after exposure to ionizing radiation in vivo: a mechanism for radiation-induced bystander effects? *Oncogene* 2001; 20:7085-95.
97. Tilney H, Rasheed S, Northover J and Tekkis P. The influence of circumferential resection margins on long-term outcomes following rectal cancer surgery. *Dis Colon Rectum* 2009; 52:1723-9.
98. Liebig C, Ayala G, Wilks J, Verstovsek G, Liu H, Agarwal N, Berger D and Albo D. Perineural invasion is an independent predictor of outcome in colorectal cancer. *J Clin Oncol* 2009; 27:5131-7.

References

99. Mandard A, Dalibard F, Mandard J, Marnay J, Henry-Amar M, Petiot J, Roussel A, Jacob J, Segol P and Samama G. Pathologic assessment of tumor regression after preoperative chemoradiotherapy of esophageal carcinoma. Clinicopathologic correlations. *Cancer* 1994;73:2680-6.
100. Shia J, Guillem J, Moore H, Tickoo S, Qin J, Ruo L, Suriawinata A, Paty P, Minsky B, Weiser M, Temple L, Wong W, Klimstra DS. Patterns of morphologic alteration in residual rectal carcinoma following preoperative chemoradiation and their association with long-term outcome. *Am J Surg Pathol* 2004; 28:215-23.
101. Halperin EC PC, Brady LV (editors). *Principles and Practice of Radiation Oncology*, 5th edition, Lippincott, Williams & Wilking; Philadelphia PA, 2008, pages 114-141.
102. West CM, McKay MJ, Hölscher T, Baumann M, Stratford IJ, Bristow RG, Iwakawa M, Imai T, Zingde SM, Anscher MS, Bourhis J, Begg AC, Haustermans K, Bentzen SM and Hendry JH. Molecular markers predicting radiotherapy response: report and recommendations from an International Atomic Energy Agency technical meeting. *Int J Radiat Oncol Biol Phys* 2005; 62:1264-73.
103. Liersch T, Langer C, Ghadimi B, Kulle B, Aust D, Baretton G, Schwabe W, Häusler P, Becker H and Jakob C. Lymph node status and TS gene expression are prognostic markers in stage II/III rectal cancer after neoadjuvant fluorouracil-based chemoradiotherapy. *J Clin Oncol* 2006; 24:4062-8.
104. Kandioler D, Zwrtek R, Ludwig C, Janschek E, Ploner M, Hofbauer F, Kühner I, Kappel S, Wrba F, Horvath M, Karner J, Renner K, Bergmann M, Karner-Hanusch J, Pötter R, Jakesz R, Teleky B and Herbst F. TP53 genotype but not p53 immunohistochemical result predicts response to preoperative short-term radiotherapy in rectal cancer. *Ann Surg* 2002; 235:493-8.
105. Perez RO, São Julião GP, Habr-Gama A, Kiss D, Proscurshim I, Campos FG, Gama-Rodrigues JJ and Ceconello I. The role of carcinoembryonic antigen in predicting response and survival to neoadjuvant chemoradiotherapy for distal rectal cancer. *Dis Colon Rectum* 2009; 52:1137-43.
106. Dworak O, Keilholz L and Hoffmann A. Pathological features of rectal cancer after preoperative radiochemotherapy. *Int J Colorectal Dis* 1997;12:19-23.
107. Rödel C, Martus P, Papadopoulos T, Füzesi L, Klimpfinger M, Fietkau R, Liersch T, Hohenberger W, Raab R, Sauer R and Wittekind C. Prognostic significance of tumor regression after preoperative chemoradiotherapy for rectal cancer. *J Clin Oncol* 2005; 23:8688-96.
108. Ruo L, Tickoo S, Klimstra D, Minsky B, Saltz L, Mazumdar M, Paty P, Wong W, Larson S, Cohen A and Guillem J. Long-term prognostic significance of extent of rectal cancer response to preoperative radiation and chemotherapy. *Ann Surg* 2002; 236:75-81.
109. Moore H, Gittleman A, Minsky B, Wong D, Paty P, Weiser M, Temple L, Saltz L, Shia J and Guillem J. Rate of pathologic complete response with increased interval between preoperative combined modality therapy and rectal cancer resection. *Dis Colon Rectum* 2004; 47:279-86.
110. Rich T, Skibber J, Ajani J, Buchholz D, Cleary K, Dubrow R, Levin B, Lynch P, Meterissian S and Roubein L. Preoperative infusional chemoradiation therapy for stage T3 rectal cancer. *Int J Radiat Oncol Biol Phys* 1995; 32:1025-9.
111. Vironen J, Juhola M, Kairaluoma M, Jantunen I and Kellokumpu I. Tumour regression grading in the evaluation of tumour response after different preoperative radiotherapy treatments for rectal carcinoma. *Int J Colorectal Dis* 2005; 20:440-5.
112. García VM, Batlle JF, Casado E, Burgos E, de Castro J, Belda C, Barriuso J, Sánchez JJ, García-Cabezas MA, González-Barón M and Cejas P. Immunohistochemical analysis of tumour regression grade for rectal cancer after neoadjuvant chemoradiotherapy. *J Clin Oncol* 27, 2009 (suppl I), abstract 22089.

References

113. Lehtiö K, Eskola O, Viljanen T, Oikonen V, Grönroos T, Sillanmäki L, Grénman R and Minn H. Imaging perfusion and hypoxia with PET to predict radiotherapy response in head-and-neck cancer. *Int J Radiat Oncol Biol Phys* 2004; 59:971-82.
114. Hopkins S, Fakih M and Yang GY. Positron emission tomography as predictor of rectal cancer response during or following neoadjuvant chemoradiation. *World J Gastrointest Oncol* 2010; 2:213-7.
115. Capirci C, Rubello D, Chierichetti F, Crepaldi G, Fanti S, Mandoliti G, Salviato S, Boni G, Rampin L, Polico C and Mariani G. Long-term prognostic value of 18F-FDG PET in patients with locally advanced rectal cancer previously treated with neoadjuvant radiochemotherapy. *AJR Am J Roentgenol* 2006; 187:W202-8.
116. Allen SD, Padhani AR, Dzik-Jurasz AS and Glynne-Jones R. Rectal carcinoma: MRI with histologic correlation before and after chemoradiation therapy. *AJR Am J Roentgenol* 2007; 188:442-51.
117. Guillem JG, Chessin DB, Shia J, Moore HG, Mazumdar M, Bernard B, Paty PB, Saltz L, Minsky BD, Weiser MR, Temple LK, Cohen AM and Wong WD. Clinical examination following preoperative chemoradiation for rectal cancer is not a reliable surrogate end point. *J Clin Oncol* 2005;23:3475-9.
118. UICC. International Union Against Cancer. TNM Classification of Malignant Tumours, 7th edition, Wiley-Blackwell; Singapore, 2010.
119. Bipat S, Glas A, Slors F, Zwinderman A, Bossuyt P and Stoker J. Rectal cancer: local staging and assessment of lymph node involvement with endoluminal US, CT, and MR imaging--a meta-analysis. *Radiology* 2004; 232:773-83.
120. Brown H, Luckasevic T, Medich D, Celebrezze J and Jones S. Efficacy of manual dissection of lymph nodes in colon cancer resections. *Mod Pathol* 2004; 17:402-6.
121. AJCC. Part III Digestive system: Colon and Rectum. *In* Stephen BE, Byrd DR, Compton CC, Fritz AG, Greene FL and Trotti A (editors). *Cancer Staging Handbook*. From the AJCC Cancer Staging Manual, 7th edition, Springer; New York, 2010, pages 173-206.
122. Meredith K, Hoffe S and Shibata D. The multidisciplinary management of rectal cancer. *Surg Clin North Am* 2009; 89:177-215, ix-x.
123. NCCN. Rectal Cancer. Clinical Practice Guidelines in Oncology, Version 2.2011, NCCN org. http://www.nccn.org/professionals/physician_gls/f_guidelines.asp. (12.11.2010).
124. Heald R, Husband E and Ryall R. The mesorectum in rectal cancer surgery--the clue to pelvic recurrence? *Br J Surg* 1982; 69:613-6.
125. Martling A, Holm T, Rutqvist L, Moran B, Heald R and Cedemark B. Effect of a surgical training programme on outcome of rectal cancer in the County of Stockholm. Stockholm Colorectal Cancer Study Group, Basingstoke Bowel Cancer Research Project. *Lancet* 2000; 356:93-6.
126. Wibe A, Møller B, Norstein J, Carlsen E, Wiig J, Heald R, Langmark F, Myrvold H and Søreide O. A national strategic change in treatment policy for rectal cancer--implementation of total mesorectal excision as routine treatment in Norway. A national audit. *Dis Colon Rectum* 2002; 45:857-66.
127. Kellokumpu I, Vironen J and Scheinin T. Laparoscopic repair of rectal prolapse: a prospective study evaluating surgical outcome and changes in symptoms and bowel function. *Surg Endosc* 2000; 14:634-40.
128. Kellokumpu I and Kairaluoma M. Laparoscopic repair of rectal prolapse: surgical technique. *Ann Chir Gynaecol* 2001; 90:66-9.

References

129. Kang S, Park J, Jeong S, Nam BH, Choi HS, Kim DW, Lim SB, Lee TG, Kim DY, Kim J, S Chang HJ, Lee HS, Kim SY, Jung KH, Hong YS, Kim JH, Sohn DK, Kim DH and Oh JH. Open versus laparoscopic surgery for mid or low rectal cancer after neoadjuvant chemoradiotherapy (COREAN trial): short-term outcomes of an open-label randomised controlled trial. *Lancet Oncol* 2010; 11:637-45.
130. Reza M, Blasco J, Andradas E, Cantero R and Mayol J. Systematic review of laparoscopic versus open surgery for colorectal cancer. *Br J Surg* 2006; 93:921-8.
131. Adam I, Mohamdee M, Martin I, Scott N, Finan P, Johnston D, Dixon M and Quirke P. Role of circumferential margin involvement in the local recurrence of rectal cancer. *Lancet* 1994; 344:707-11.
132. Hall N, Finan P, al-Jaberi T, Tsang C, Brown S, Dixon M and Quirke P. Circumferential margin involvement after mesorectal excision of rectal cancer with curative intent. Predictor of survival but not local recurrence? *Dis Colon Rectum* 1998; 41:979-83.
133. Compton C, Fenoglio-Preiser C, Pettigrew N and Fielding L. American Joint Committee on Cancer Prognostic Factors Consensus Conference: Colorectal Working Group. *Cancer* 2000; 88:1739-57.
134. Dukes C and Bussey H. The spread of rectal cancer and its effect on prognosis. *Br J Cancer* 1958; 12:309-20.
135. Kuo L, Liu M, Jian J, Horng C, Cheng T, Chen C, Fang W and Chung Y. Is final TNM staging a predictor for survival in locally advanced rectal cancer after preoperative chemoradiation therapy? *Ann Surg Oncol* 2007; 14:2766-72.
136. McDermott F, Hughes E, Pihl E, Milne B and Price A. Influence of tumour differentiation on survival after resection for rectal cancer in a series of 1296 patients. *Aust N Z J Surg* 1984; 54:53-8.
137. Kim C, Anne, PR, Mitchell EP, Palazzo J, Pequinot E, fishel R, Goldstein SD, Isenberg GA and Curran W. Tumor size as a predictor of pathologic complete response (pCR) and survival in a prospective trial of chemoradiation for locally advanced rectal cancer. *J Clin Oncol* 23 (16S): 3672, 2005.
138. Tepper J. Adjuvant radiation therapy of rectal cancer. *J Clin Oncol* 2001; 19:3709-11.
139. Griffin M, Bergstralh E, Coffey R, Beart RJ and Melton Lr. Predictors of survival after curative resection of carcinoma of the colon and rectum. *Cancer* 1987; 60:2318-24.
140. Edler D, Ohrling K, Hallström M, Karlberg M and Ragnhammar P. The number of analyzed lymph nodes - a prognostic factor in colorectal cancer. *Acta Oncol* 2007; 46:975-81.
141. Peeters K, Marijnen C, Nagtegaal I, Kranenbarg E, Putter H, Wiggers T, Rutten H, Pahlman L, Glimelius B, Leer J and van de Velde C. The TME trial after a median follow-up of 6 years: increased local control but no survival benefit in irradiated patients with resectable rectal carcinoma. *Ann Surg* 2007; 246:693-701.
142. Zirngibl H, Husemann B and Hermanek P. Intraoperative spillage of tumor cells in surgery for rectal cancer. *Dis Colon Rectum* 1990; 33:610-4.
143. Kosciński T, Malinger S and Drews M. Local excision of rectal carcinoma not-exceeding the muscularis layer. *Colorectal Dis* 2003; 5:159-63.
144. Vironen J, Halme L, Sainio P, Kyllönen L, Scheinin T, Husa A and Kellokumpu I. New approaches in the management of rectal carcinoma result in reduced local recurrence rate and improved survival. *Eur J Surg* 2002; 168:158-64.
145. Kachnic L and Willett C. Radiation therapy in the management of rectal cancer. *Curr Opin Oncol* 2001; 13:300-6.

References

146. Díaz-González J, Calvo F, Cortés J, García-Sabrido J, Gómez-Espí M, Del Valle E, Muñoz-Jiménez F and Alvarez E. Prognostic factors for disease-free survival in patients with T3-4 or N+ rectal cancer treated with preoperative chemoradiation therapy, surgery, and intraoperative irradiation. *Int J Radiat Oncol Biol Phys* 2006; 64:1122-8.
147. Lepistö A, Österlund P, Kouri M and Järvinen H. [Rectal cancer]. *Duodecim* 2009; 125:857-65.
148. Rosenberg R, Maak M, Schuster T, Becker K, Friess H and Gertler R. Does a rectal cancer of the upper third behave more like a colon or a rectal cancer? *Dis Colon Rectum* 2010; 53:761-70.
149. Scott N, Susnerwala S, Gollins S, Myint A and Levine E. Preoperative neo-adjuvant therapy for curable rectal cancer--reaching a consensus 2008. *Colorectal Dis* 2009; 11:245-8.
150. Hoffman K, Hong T, Zietman A and Russell A. External beam radiation treatment for rectal cancer is associated with a decrease in subsequent prostate cancer diagnosis. *Cancer* 2008; 112:943-9.
151. Folkesson J, Birgisson H, Pahlman L, Cedermark B, Glimelius B and Gunnarsson U. Swedish Rectal Cancer Trial: long lasting benefits from radiotherapy on survival and local recurrence rate. *J Clin Oncol* 2005; 23:5644-50.
152. Bosset J, Collette L, Calais G, Mineur L, Maingon P, Radosevic-Jelic L, Daban A, Bardet E, Beny A, and Ollier J. EORTC Radiotherapy Group Trial 22921. Chemotherapy with preoperative radiotherapy in rectal cancer. *N Engl J Med* 2006; 355:1114-23.
153. Gérard J, Conroy T, Bonnetain F, Bouché O, Chapet O, Closon-Dejardin M, Untereiner M, Leduc B, Francois E, Maurel J, Seitz J, Buecher B, Mackiewicz R, Ducreux M and Bedenne L. Preoperative radiotherapy with or without concurrent fluorouracil and leucovorin in T3-4 rectal cancers: results of FFCD 9203. *J Clin Oncol* 2006; 24:4620-5.
154. Poon M, O'Connell M, Moertel C, Wieand H, Cullinan S, Everson L, Krook J, Mailliard J, Laurie J and Tschetter L. Biochemical modulation of fluorouracil: evidence of significant improvement of survival and quality of life in patients with advanced colorectal carcinoma. *J Clin Oncol* 1989; 7:1407-18.
155. Reigner B, Blesch K and Weidekamm E. Clinical pharmacokinetics of capecitabine. *Clin Pharmacokinet* 2001; 40:85-104.
156. De Paoli A, Chiara S, Luppi G, Friso M, Beretta G, Del Prete S, Pasetto L, Santantonio M, Sarti E, Mantello G, Innocente R, Frustaci S, Corvò R and Rosso R. Capecitabine in combination with preoperative radiation therapy in locally advanced, resectable, rectal cancer: a multicentric phase II study. *Ann Oncol* 2006; 17:246-51.
157. Bujko K, Nowacki MP, Nasierowska-Guttmejer A, Michalski W, Bebenek M and Kryj M. Long-term results of a randomized trial comparing preoperative short-course radiotherapy with preoperative conventionally fractionated chemoradiation for rectal cancer. *Br J Surg* 2006; 93:1215-23.
158. Guillem J, Chessin D, Cohen A, Shia J, Mazumdar M, Enker W, Paty P, Weiser M, Klimstra D, Saltz L, Minsky B and Wong W. Long-term oncologic outcome following preoperative combined modality therapy and total mesorectal excision of locally advanced rectal cancer. *Ann Surg* 2005; 241:829-38.
159. Carlomagno C, Farella A, Bucci L, D'Armiento F, Pesce G, Pepe S, Cannella L, Pacelli R, De Stefano A, Solla R, D'Armiento M and De Placido S. Neo-adjuvant treatment of rectal cancer with capecitabine and oxaliplatin in combination with radiotherapy: a phase II study. *Ann Oncol* 2009; 20:906-12.
160. Hospers GA, Punt CJ, Tesselaar ME, Cats A, Havenga K, Leer JW, Marijnen CA, Jansen EP, Van Krieken HH, Wiggers T, Van de Velde CJ and Mulder NH. Preoperative chemoradiotherapy with capecitabine and oxaliplatin in locally advanced rectal cancer. A phase I-II multicenter study of the Dutch Colorectal Cancer Group. *Ann Surg Oncol* 2007; 14:2773-9.

References

161. Valentini V and Cellini F. Surgery: neoadjuvant chemoradiation and sphincter preservation. *Nat Rev Gastroenterol Hepatol* 2009; 6:327-9.
162. Francois Y, Nemoz C, Baulieux J, Vignal J, Grandjean J, Partensky C, Souquet J, Adeleine P and Gerard J. Influence of the interval between preoperative radiation therapy and surgery on downstaging and on the rate of sphincter-sparing surgery for rectal cancer: the Lyon R90-01 randomized trial. *J Clin Oncol* 1999; 17:2396.
163. Suit HD and Gallager HS. Intact tumor cells in irradiated tissue. *Arch Pathol* 1964; 78:648-51.
164. Kalady MF, de Campos-Lobato LF, Stocchi L, Geisler DP, Dietz D, Lavery IC and Fazio VW. Predictive factors of pathologic complete response after neoadjuvant chemoradiation for rectal cancer. *Ann Surg* 2009; 250:582-589.
165. Smith FM, Waldron D and Winter DC. Rectum-conserving surgery in the era of chemoradiotherapy. *Br J Surg* 2010; 97:1752-64.
166. Radu C, Berglund A, Pählman L and Glimelius B. Short-course preoperative radiotherapy with delayed surgery in rectal cancer - a retrospective study. *Radiother Oncol* 2008; 87:343-9.
167. Medical Research Council Rectal Cancer Working Party. Randomised trial of surgery alone versus radiotherapy followed by surgery for potentially operable locally advanced rectal cancer. *Lancet* 1996; 348:1605-10.
168. Gastrointestinal Tumor Study Group. Prolongation of the Disease-Free Interval in Surgically Treated Rectal Carcinoma *N Engl J Med* 1985; 1985:1465-72.
169. Krook J, Moertel C, Gunderson L, Wieand H, Collins R, Beart R, Kubista T, Poon M, Meyers W and Mailliard J. Effective surgical adjuvant therapy for high-risk rectal carcinoma. *N Engl J Med* 1991; 324:709-15.
170. Glynne-Jones R, Mawdsley S and Harrison M. Cetuximab and chemoradiation for rectal cancer--is the water getting muddy? *Acta Oncol* 2010; 49:278-86.
171. Bonner J, Harari P, Giralt J, Cohen R, Jones C, Sur R, Raben D, Baselga J, Spencer S, Zhu J, Yousoufian H, Rowinsky E and Ang K. Radiotherapy plus cetuximab for locoregionally advanced head and neck cancer: 5-year survival data from a phase 3 randomised trial, and relation between cetuximab-induced rash and survival. *Lancet Oncol* 2010; 11:21-8.
172. Harari P and Huang S. Combining EGFR inhibitors with radiation or chemotherapy: will preclinical studies predict clinical results? *Int J Radiat Oncol Biol Phys* 2004; 58:976-83.
173. Nevo J, Mattila E, Pellinen T, Yamamoto D, Sara H, Iljin K, Kallioniemi O, Bono P, Heikkilä P, Joensuu H, Wärri A and Ivaska J. Mammary-derived growth inhibitor alters traffic of EGFR and induces a novel form of cetuximab resistance. *Clin Cancer Res* 2009; 15:6570-81.
174. Pinto C, Di Fabio F, Maiello E, Di Tullio P, Pini S, Aschele C, Garufi A, Bochicchio G, Pinotti G, Latiano T and Martoni A: Phase II study of preoperative panitumumab, 5-fluorouracil, and oxaliplatin with concurrent radiotherapy in locally advanced rectal cancer: Preliminary safety results. *J Clin Oncol, ASCO Annual Meeting Proceedings* 2009; 27(15S): 4110.
175. Bono P and Joensuu H. [Anti-angiogenic treatment is effective for treating metastatic colorectal cancers]. *Duodecim* 2004; 120:261-2.
176. Crane C, Eng C, Feig B, Das P, Skibber J, Chang G, Wolff R, Krishnan S, Hamilton S, Janjan N, Maru D, Ellis L and Rodriguez-Bigas M. Phase II trial of neoadjuvant bevacizumab, capecitabine, and radiotherapy for locally advanced rectal cancer. *Int J Radiat Oncol Biol Phys* 2010; 76:824-30.

References

177. Labianca R, Nordlinger B, Beretta G, Brouquet A and Cervantes A for ESMO Guidelines Working Group. Primary colon cancer: ESMO Clinical Practice Guidelines for diagnosis, adjuvant treatment and follow-up. *Ann Oncol* 2010; 21 Suppl 5:v70-7.
178. Moertel C, Fleming T, Macdonald J, Haller D, Laurie J, Goodman P, Ungerleider J, Emerson W, Tormey D and Glick J. Levamisole and fluorouracil for adjuvant therapy of resected colon carcinoma. *N Engl J Med* 1990; 322:352-8.
179. O'Connell M, Laurie J, Kahn M, Fitzgibbons RJ, Erlichman C, Shepherd L, Moertel C, Kocha W, Pazdur R, Wieand H, Rubin J, Vukov A, Donohue J, Krook J and Figueredo A. Prospectively randomized trial of postoperative adjuvant chemotherapy in patients with high-risk colon cancer. *J Clin Oncol* 1998; 16:295-300.
180. André T, Colin P, Louvet C, Gamelin E, Bouche O, Achille E, Colbert N, Boaziz C, Piedbois P, Tubiana-Mathieu N, Boutan-Laroze A, Flesch M, Buyse M and Gramont A. Semimonthly versus monthly regimen of fluorouracil and leucovorin administered for 24 or 36 weeks as adjuvant therapy in stage II and III colon cancer: results of a randomized trial. *J Clin Oncol* 2003; 21:2896-903.
181. Twelves C, Wong A, Nowacki M, Abt M, Burris Hr, Carrato A, Cassidy J, Cervantes A, Fagerberg J, Georgoulas V, Hussein F, Jodrell D, Koralewski P, Kröning H, Maroun J, Marschner N, McKendrick J, Pawliski M, Rosso R, Schüller J, Seitz JF, Stabuc B, Tujakowski J, Van Hazel G, Zaluski J and Scheithauer W. Capecitabine as adjuvant treatment for stage III colon cancer. *N Engl J Med* 2005; 352:2696-704.
182. André T, Boni C, Navarro M, Tabernero J, Hickish T, Topham C, Bonetti A, Clingan P, Bridgewater J, Rivera F and de Gramont A. Improved overall survival with oxaliplatin, fluorouracil, and leucovorin as adjuvant treatment in stage II or III colon cancer in the MOSAIC trial. *J Clin Oncol* 2009; 27:3109-16.
183. Bujko K, Glynne-Jones R and Bujko M. Does adjuvant fluoropyrimidine-based chemotherapy provide benefit for patients with resected rectal cancer who have already received neoadjuvant radiochemotherapy? A systematic review of randomised trials. *Ann Oncol* 2010; 21: 1743-1750.
184. Collette L, Bosset JF, den Dulk M, Nguyen F, Mineur L, Maingon P, Radosevic-Jelic L, Piérart M and Calais G. Patients with curative resection of cT3-4 rectal cancer after preoperative radiotherapy or radiochemotherapy: does anybody benefit from adjuvant fluorouracil-based chemotherapy? A trial of the European Organisation for Research and Treatment of Cancer Radiation Oncology Group. *J Clin Oncol* 2007; 25:4379-86.
185. Sakamoto J, Hamada C, Yoshida S, Kodaira S, Yasutomi M, Kato T, Oba K, Nakazato H, Saji S and Ohashi Y. An individual patient data meta-analysis of adjuvant therapy with uracil-tegafur (UFT) in patients with curatively resected rectal cancer. *Br J Cancer* 2007; 96:1170-7.
186. Van Cutsem E, Dico M, Haustermans K, Arber N, Bosset J, Cunningham D, De Gramont A, Diaz-Rubio E, Ducreux M, Goldberg R, Glynne-Jones R, Haller D, Kang YK, Kerr D, Labianca R, Minsky RD, Moore M, Nordlinger B, Rougier P, Scheithauer W, Schmoll HJ, Sobrero A, Tabernero M, Van de Velde C and Zalberg J. The diagnosis and management of rectal cancer: expert discussion and recommendations derived from the 9th World Congress on Gastrointestinal Cancer, Barcelona, 2007. *Ann Oncol* 2008; 19 Suppl 6:vi1-8.
187. Eskelinen M, Pasanen P, Kulju A, Janatuinen E, Miettinen P, Poikolainen E, Tarvainen R, Nuutinen P, Pääkkönen M and Alhava E. Clinical evaluation of serum tumour markers CEA, CA 50 and CA 242 in colorectal cancer. *Anticancer Res*; 14:1427-32.

References

188. Kokkonen N, Ulibarri I, Kauppila A, Luosujärvi H, Rivinoja A, Pospiech H, Kellokumpu I and Kellokumpu S. Hypoxia upregulates carcinoembryonic antigen expression in cancer cells. *Int J Cancer* 2007; 121:2443-50.
189. Plebani M, De Paoli M, Basso D, Roveroni G, Giacomini A, Galeotti F and Corsini A. Serum tumor markers in colorectal cancer staging, grading, and follow-up. *J Surg Oncol* 1996; 62:239-44.
190. Koukourakis M, Giatromanolaki A, Sivridis E, Gatter K and Harris A. Lactate dehydrogenase 5 expression in operable colorectal cancer: strong association with survival and activated vascular endothelial growth factor pathway--a report of the Tumour Angiogenesis Research Group. *J Clin Oncol* 2006; 24:4301-8.
191. Locker G, Hamilton S, Harris J, Jessup J, Kemeny N, Macdonald J, Somerfield M, Hayes D and Bast RJ for ASCO. ASCO 2006 update of recommendations for the use of tumor markers in gastrointestinal cancer. *J Clin Oncol* 2006; 24:5313-27.
192. Jeffery M, Hickey BE and Hider PN. Follow-up strategies for patients treated for non-metastatic colorectal cancer. *Cochrane Database Syst Rev* 2007; 1:CD002200.
193. Järvinen H, Renkonen-Sinisalo L, Aktán-Collán K, Peltomäki P, Aaltonen L, Mecklin J. Ten years after mutation testing for Lynch syndrome: cancer incidence and outcome in mutation-positive and mutation-negative family members. *J Clin Oncol* 2009; 27:4793-7.
194. Marijnen C, Nagtegaal I, Kapiteijn E, Kranenbarg E, Noordijk E, van Krieken J, van de Velde C and Leer J. Radiotherapy does not compensate for positive resection margins in rectal cancer patients: report of a multicenter randomized trial. *Int J Radiat Oncol Biol Phys* 2003; 55:1311-20.
195. Finnish Cancer Registry. Rectal cancer survival data. (personal communication) (27.1.2010).
196. Leach R and Treacher D. Oxygen transport-2. Tissue hypoxia. *BMJ* 1998; 317:1370-3.
197. Folkman J. Tumor angiogenesis: therapeutic implications. *N Engl J Med* 1971; 285:1182-6.
198. Rofstad E and Halsør E. Vascular endothelial growth factor, interleukin 8, platelet-derived endothelial cell growth factor, and basic fibroblast growth factor promote angiogenesis and metastasis in human melanoma xenografts. *Cancer Res* 2000; 60:4932-8.
199. Hillen F and Griffioen A. Tumour vascularization: sprouting angiogenesis and beyond. *Cancer Metastasis Rev* 2007; 26:489-502.
200. Miller J, Pien H, Sahani D, Sorensen A and Thrall J. Imaging angiogenesis: applications and potential for drug development. *J Natl Cancer Inst* 2005; 97:172-87.
201. Krebs H and Johnson W. Metabolism of ketonic acids in animal tissues. *Biochem J* 1937; 31:645-60.
202. Stickland L. The Pasteur effect in normal yeast and its inhibition by various agents. *Biochem J* 1956; 64:503-15.
203. Warburg O. On the origin of cancer cells. *Science* 1956; 123:309-14.
204. Höpfl G, Ogunshola O and Gassmann M. HIFs and tumors--causes and consequences. *Am J Physiol Regul Integr Comp Physiol* 2004; 286:R608-23.
205. Greijer A, Delis-van Diemen P, Fijneman R, Giles R, Voest E, van Hinsbergh V and Meijer G. Presence of HIF-1 and related genes in normal mucosa, adenomas and carcinomas of the colorectum. *Virchows Arch* 2008;452:535-44.

References

206. Jokilehto T and Jaakkola P. The role of HIF prolyl hydroxylases in tumour growth. *J Cell Mol Med* 2010; 14:758-70.
207. Semenza G. HIF-1: using two hands to flip the angiogenic switch. *Cancer Metastasis Rev* 2000; 19:59-65.
208. Wang G and Semenza G. Purification and characterization of hypoxia-inducible factor 1. *J Biol Chem* 1995; 270:1230-7.
209. Jaakkola P, Mole D, Tian Y, Wilson M, Gielbert J, Gaskell S, Kriegsheim Av, Hebestreit H, Mukherji M, Schofield C, Maxwell P, Pugh C and Ratcliffe P. Targeting of HIF- α to the von Hippel-Lindau ubiquitylation complex by O₂-regulated prolyl hydroxylation. *Science* 2001; 292:468-72.
210. Blancher C, Moore J, Robertson N and Harris A. Effects of ras and von Hippel-Lindau (VHL) gene mutations on hypoxia-inducible factor (HIF)-1 α , HIF-2 α , and vascular endothelial growth factor expression and their regulation by the phosphatidylinositol 3'-kinase/Akt signaling pathway. *Cancer Res* 2001; 61:7349-55.
211. Koukourakis M, Pitiakoudis M, Giatromanolaki A, Tsarouha A, Polychronidis A, Sivridis E and Simopoulos C. Oxygen and glucose consumption in gastrointestinal adenocarcinomas: correlation with markers of hypoxia, acidity and anaerobic glycolysis. *Cancer Sci* 2006; 97:1056-60.
212. Maxwell P, Pugh C and Ratcliffe P. Activation of the HIF pathway in cancer. *Curr Opin Genet Dev* 2001; 11:293-9.
213. Weidemann A and Johnson R. Biology of HIF-1 α . *Cell Death Differ* 2008; 15:621-7.
214. Sekulić A, Hudson C, Homme J, Yin P, Otterness D, Karnitz L and Abraham R. A direct linkage between the phosphoinositide 3-kinase-AKT signaling pathway and the mammalian target of rapamycin in mitogen-stimulated and transformed cells. *Cancer Res* 2000; 60:3504-13.
215. Laughner E, Taghavi P, Chiles K, Mahon P and Semenza G. HER2 (neu) signaling increases the rate of hypoxia-inducible factor 1 α (HIF-1 α) synthesis: novel mechanism for HIF-1-mediated vascular endothelial growth factor expression. *Mol Cell Biol* 2001; 21:3995-4004.
216. Alitalo K, Tammela T and Petrova T. Lymphangiogenesis in development and human disease. *Nature* 2005; 438:946-53.
217. Vaupel P and Harrison L. Tumor hypoxia: causative factors, compensatory mechanisms, and cellular response. *Oncologist* 2004; 9 Suppl 5:4-9.
218. Carmeliet P. VEGF as a key mediator of angiogenesis in cancer. *Oncology* 2005;69 Suppl 3:4-10.
219. George M, Tutton M, Janssen F, Arnaout A, Abulafi A, Eccles S and Swift R. VEGF-A, VEGF-C, and VEGF-D in colorectal cancer progression. *Neoplasia* 2001; 3:420-7.
220. Petrova T, Bono P, Holnthoner W, Chesnes J, Pytowski B, Sihto H, Laakkonen P, Heikkilä P, Joensuu H and Alitalo K. VEGFR-3 expression is restricted to blood and lymphatic vessels in solid tumors. *Cancer Cell* 2008; 13:554-6.
221. Pastorek J, Pastoreková S, Callebaut I, Mornon J, Zelník V, Opavský R, Zat'ovicová M, Liao S, Portetelle D and Stanbridge E. Cloning and characterization of MN, a human tumor-associated protein with a domain homologous to carbonic anhydrase and a putative helix-loop-helix DNA binding segment. *Oncogene* 1994; 9:2877-88.
222. Ivanov S, Liao S, Ivanova A, Danilkovitch-Miagkova A, Tarasova N, Weirich G, Merrill M, Proescholdt M, Oldfield E, Lee J, Zavada J, Waheed A, Sly W, Lerman M and Stanbridge E. Expression of hypoxia-

References

- inducible cell-surface transmembrane carbonic anhydrases in human cancer. *Am J Pathol* 2001; 158:905-19.
223. Saarnio J, Parkkila S, Parkkila A, Haukipuro K, Pastoreková S, Pastorek J, Kairaluoma M and Karttunen T. Immunohistochemical study of colorectal tumors for expression of a novel transmembrane carbonic anhydrase, MN/CA IX, with potential value as a marker of cell proliferation. *Am J Pathol* 1998; 153:279-85.
224. Chiche J, Ilc K, Laferrrière J, Trottier E, Dayan F, Mazure N, Brahim-Horn M and Pouyssegur J. Hypoxia-inducible carbonic anhydrase IX and XII promote tumor cell growth by counteracting acidosis through the regulation of the intracellular pH. *Cancer Res* 2009; 69:358-68.
225. Pastorekova S, Kopacek J and Pastorek J. Carbonic anhydrase inhibitors and the management of cancer. *Curr Top Med Chem* 2007; 7:865-78.
226. Wykoff C, Pugh C, Maxwell P, Harris A and Ratcliffe P. Identification of novel hypoxia dependent and independent target genes of the von Hippel-Lindau (VHL) tumour suppressor by mRNA differential expression profiling. *Oncogene* 2000; 19:6297-305.
227. Brennan D, Jirstrom K, Kronblad A, Millikan R, Landberg G, Duffy M, Rydén L, Gallagher W and O'Brien S. CA IX is an independent prognostic marker in premenopausal breast cancer patients with one to three positive lymph nodes and a putative marker of radiation resistance. *Clin Cancer Res* 2006; 12:6421-31.
228. Robertson N, Potter C and Harris A. Role of carbonic anhydrase IX in human tumor cell growth, survival, and invasion. *Cancer Res* 2004; 64:6160-5.
229. Hilvo M, Supuran C and Parkkila S. Characterization and inhibition of the recently discovered carbonic anhydrase isoforms CA XIII, XIV and XV. *Curr Top Med Chem* 2007; 7:893-9.
230. Supuran C. Carbonic anhydrases: novel therapeutic applications for inhibitors and activators. *Nat Rev Drug Discov* 2008; 7:168-81.
231. Pessin J and Bell G. Mammalian facilitative glucose transporter family: structure and molecular regulation. *Annu Rev Physiol* 1992; 54:911-30.
232. Birnbaum M, Haspel H and Rosen O. Cloning and characterization of a cDNA encoding the rat brain glucose-transporter protein. *Proc Natl Acad Sci U S A* 1986; 83:5784-8.
233. Macheda M, Rogers S and Best J. Molecular and cellular regulation of glucose transporter (GLUT) proteins in cancer. *J Cell Physiol* 2005; 202:654-62.
234. Mellanen P, Minn H, Grénman R and Härkönen P. Expression of glucose transporters in head-and-neck tumors. *Int J Cancer* 1994; 56:622-9.
235. Airley R, Loncaster J, Raleigh J, Harris A, Davidson S, Hunter R, West C and Stratford I. GLUT-1 and CAIX as intrinsic markers of hypoxia in carcinoma of the cervix: relationship to pimonidazole binding. *Int J Cancer* 2003; 104:85-91.
236. Evans A, Bates V, Troy H, Hewitt S, Holbeck S, Chung Y, Phillips R, Stubbs M, Griffiths J and Airley R. Glut-1 as a therapeutic target: increased chemoresistance and HIF-1-independent link with cell turnover is revealed through COMPARE analysis and metabolomic studies. *Cancer Chemother Pharmacol* 2008; 61:377-93.
237. Cleven A, van Engeland M, Wouters B and de Bruïne A. Stromal expression of hypoxia regulated proteins is an adverse prognostic factor in colorectal carcinomas. *Cell Oncol* 2007; 29:229-40.

References

238. Ito H, Duxbury M, Zinner M, Ashley S and Whang E. Glucose transporter-1 gene expression is associated with pancreatic cancer invasiveness and MMP-2 activity. *Surgery* 2004; 136:548-56.
239. Gould G, and Bell G. Facilitative glucose transporters: an expanding family. *Trends Biochem Sci* 1990; 15:18-23.
240. Behrooz A and Ismail-Beigi F. Dual control of glut1 glucose transporter gene expression by hypoxia and by inhibition of oxidative phosphorylation. *J Biol Chem* 1997; 272:5555-62.
241. Bretscher A. Rapid phosphorylation and reorganization of ezrin and spectrin accompany morphological changes induced in A-431 cells by epidermal growth factor. *J Cell Biol* 1989; 108:921-30.
242. Elzagheid A, Korkeila E, Bendardaf R, Buhmeida A, Heikkilä S, Vaheeri A, Syrjänen K, Pyrhönen S and Carpén O. Intense cytoplasmic ezrin immunoreactivity predicts poor survival in colorectal cancer. *Hum Pathol* 2008; 39:1737-43.
243. Wang H, Zhu J, Zhang Q, Sun Q and Guo H. High level of ezrin expression in colorectal cancer tissues is closely related to tumor malignancy. *World J Gastroenterol* 2009; 15:2016-9.
244. Bretscher A, Edwards K and Fehon R. ERM proteins and merlin: integrators at the cell cortex. *Nat Rev Mol Cell Biol* 2002; 3:586-99.
245. Di Cristofano C, Leopizzi M, Miraglia A, Sardella B, Moretti V, Ferrara A, Petrozza V and Rocca C. Phosphorylated ezrin is located in the nucleus of the osteosarcoma cell. *Mod Pathol* 2010, 23: 1012-20.
246. Gould K, Bretscher A, Esch F and Hunter T. cDNA cloning and sequencing of the protein-tyrosine kinase substrate, ezrin, reveals homology to band 4.1. *EMBO J* 1989; 8:4133-42.
247. Heiska L, Kantor C, Parr T, Critchley D, Vilja P, Gahmberg C and Carpén O. Binding of the cytoplasmic domain of intercellular adhesion molecule-2 (ICAM-2) to alpha-actinin. *J Biol Chem* 1996; 271:26214-9.
248. Bono P, Cordero E, Johnson K, Borowsky M, Ramesh V, Jacks T and Hynes R. Layilin, a cell surface hyaluronan receptor, interacts with merlin and radixin. *Exp Cell Res* 2005; 308:177-87.
249. D'Amico A and McKenna W. Apoptosis and a re-investigation of the biologic basis for cancer therapy. *Radiother Oncol* 1994; 33:3-10.
250. Greijer A and van der Wall E. The role of hypoxia inducible factor 1 (HIF-1) in hypoxia induced apoptosis. *J Clin Pathol* 2004; 57:1009-14.
251. Milas L CJ. Principles of Combining Radiation Therapy and Chemotherapy, Chapter IV in Cox JD and Ang KK (editors): *Radiation Oncology*, 8th edition, Mosby; St Louis Kentucky, 2003, pages 108-124.
252. Gray L, Conger A, Ebert M, Hornsey S and Scott O. The concentration of oxygen dissolved in tissues at the time of irradiation as a factor in radiotherapy. *Br J Radiol* 1953; 26:638-48.
253. Crockart N, Jordan BF, Baudelet C, Ansiaux R, Sonveaux P, Grégoire V, Beghein N, DeWever J, Bouzin C, Feron O and Gallez B. Early reoxygenation in tumors after irradiation: determining factors and consequences for radiotherapy regimens using daily multiple fractions. *Int J Radiat Oncol Biol Phys* 2005;63:901-10.
254. Crockart N, Radermacher K, Jordan BF, Baudelet C, Cron GO, Grégoire V, Beghein N, Bouzin C, Feron O and Gallez B. Tumor radiosensitization by antiinflammatory drugs: evidence for a new mechanism involving the oxygen effect. *Cancer Res* 2005; 65:7911-6.

References

255. Vaupel P, Frinak S and O'Hara M. Direct measurement of reoxygenation in malignant mammary tumors after a single large dose of irradiation. *Adv Exp Med Biol* 1984; 180:773-82.
256. Overgaard J. Clinical evaluation of nitroimidazoles as modifiers of hypoxia in solid tumors. *Oncol Res* 1994; 6:509-18.
257. Elder DJ, Halton DE, Hague A and Paraskeva C. Induction of apoptotic cell death in human colorectal carcinoma cell lines by a cyclooxygenase-2 (COX-2)-selective nonsteroidal anti-inflammatory drug: independence from COX-2 protein expression. *Clin Cancer Res* 1997; 3:1679-83.
258. Sooriakumaran P. COX-2 inhibitors and the heart: are all coxibs the same? *Postgrad Med J* 2006; 82:242-5.
259. Tveit KM, Guldvog I, Hagen S, Trondsen E, Harbitz T, Nygaard K, Nilsen JB, Wist E and Hannisdal E. Randomized controlled trial of postoperative radiotherapy and short-term time-scheduled 5-fluorouracil against surgery alone in the treatment of Dukes B and C rectal cancer. Norwegian Adjuvant Rectal Cancer Project Group. *Br J Surg* 1997; 84:1130-5.
260. Siddik ZH. Cisplatin: mode of cytotoxic action and molecular basis of resistance. *Oncogene* 2003; 22:7265-79.
261. Yapp DT, Lloyd DK, Zhu J and Lehnert SM. The potentiation of the effect of radiation treatment by intratumoral delivery of cisplatin. *Int J Radiat Oncol Biol Phys* 1998; 42:413-20.
262. Cividalli A, Ceciarelli F, Livdi E, Altavista P, Cruciani G, Marchetti P and Danesi DT. Radiosensitization by oxaliplatin in a mouse adenocarcinoma: influence of treatment schedule. *Int J Radiat Oncol Biol Phys* 2002; 52:1092-8.
263. Brown JM. Exploiting the hypoxic cancer cell: mechanisms and therapeutic strategies. *Mol Med Today* 2000; 6:157-62.
264. Wang J, Biedermann KA and Brown JM. Repair of DNA and chromosome breaks in cells exposed to SR 4233 under hypoxia or to ionizing radiation. *Cancer Res* 1992; 52:4473-7.
265. Rischin D, Narayan K, Oza AM, Mileskin L, Bernshaw D, Choi J, Hicks R, McClure B and Fyles A. Phase 1 study of tirapazamine in combination with radiation and weekly cisplatin in patients with locally advanced cervical cancer. *Int J Gynecol Cancer* 2010; 20:827-33.
266. Kohshi K, Kinoshita Y, Imada H, Kunugita N, Abe H, Terashima H, Tokui N and Uemura S. Effects of radiotherapy after hyperbaric oxygenation on malignant gliomas. *Br J Cancer* 1999;80:236-41.
267. Brown JM and Wilson WR. Exploiting tumour hypoxia in cancer treatment. *Nat Rev Cancer* 2004;4:437-47.
268. Bush RS, Jenkin RD, Allt WE, Beale FA, Bean H, Dembo AJ and Pringle JF. Definitive evidence for hypoxic cells influencing cure in cancer therapy. *Br J Cancer Suppl* 1978;3:302-6.
269. Grogan M, Thomas GM, Melamed I, Wong FL, Pearcey RG, Joseph PK, Portelance L, Crook J and Jones KD. The importance of hemoglobin levels during radiotherapy for carcinoma of the cervix. *Cancer* 1999; 86:1528-36.
270. Hershman DL, Buono DL, Malin J, McBride R, Tsai WY and Neugut AI. Patterns of use and risks associated with erythropoiesis-stimulating agents among Medicare patients with cancer. *J Natl Cancer Inst* 2009; 101:1633-41.
271. Glaspy J, Crawford J, Vansteenkiste J, Henry D, Rao S, Bowers P, Berlin JA, Tomita D, Bridges K and Ludwig H. Erythropoiesis-stimulating agents in oncology: a study-level meta-analysis of survival and other safety outcomes. *Br J Cancer* 2010; 102:301-15.

References

272. Koukourakis M, Giatromanolaki A, Sivridis E, Pastorek J, Karapantzos I, Gatter K and Harris A. Hypoxia-activated tumor pathways of angiogenesis and pH regulation independent of anemia in head-and-neck cancer. *Int J Radiat Oncol Biol Phys* 2004; 59:67-71.
273. Erler J, Cawthorne C, Williams K, Koritzinsky M, Wouters B, Wilson C, Miller C, Demonacos C, Stratford I, Dive C. Hypoxia-mediated down-regulation of Bid and Bax in tumors occurs via hypoxia-inducible factor 1-dependent and -independent mechanisms and contributes to drug resistance. *Mol Cell Biol* 2004; 24:2875-89.
274. Graeber T, Osmanian C, Jacks T, Housman D, Koch C, Lowe S and Giaccia A. Hypoxia-mediated selection of cells with diminished apoptotic potential in solid tumours. *Nature* 1996; 379:88-91.
275. Birner P, Schindl M, Obermair A, Plank C, Breitenecker G and Oberhuber G. Overexpression of hypoxia-inducible factor 1alpha is a marker for an unfavorable prognosis in early-stage invasive cervical cancer. *Cancer Res* 2000;60:4693-6.
276. Vaupel P, Mayer A and Höckel M. Oxygenation status of primary and recurrent squamous cell carcinomas of the vulva. *Eur J Gynaecol Oncol* 2006;27:142-6.
277. Lu X, Xing C, Feng Y, Chen J and Deng C. Clinical significance of immunohistochemical expression of hypoxia-inducible factor-1alpha as a prognostic marker in rectal adenocarcinoma. *Clin Colorectal Cancer* 2006;5:350-3.
278. Theodoropoulos G, Lazaris A, Theodoropoulos V, Papatheodosiou K, Gazouli M, Bramis J, Patsouris E and Panoussopoulos D. Hypoxia, angiogenesis and apoptosis markers in locally advanced rectal cancer. *Int J Colorectal Dis* 2006; 21:248-57.
279. Brophy S, Sheehan K, McNamara D, Deasy J, Bouchier-Hayes D and Kay E. GLUT-1 expression and response to chemoradiotherapy in rectal cancer. *Int J Cancer* 2009; 125:2778-82.
280. Haber R, Rathan A, Weiser K, Pritsker A, Itzkowitz S, Bodian C, Slater G, Weiss A and Burstein D. GLUT1 glucose transporter expression in colorectal carcinoma: a marker for poor prognosis. *Cancer* 1998; 83:34-40.
281. Koukourakis M, Giatromanolaki A, Sivridis E, Simopoulos C, Turley H, Talks K, Gatter K and Harris A. Hypoxia-inducible factor (HIF1A and HIF2A), angiogenesis, and chemoradiotherapy outcome of squamous cell head-and-neck cancer. *Int J Radiat Oncol Biol Phys* 2002; 53:1192-202.
282. Bache M, Reddemann R, Said H, Holzhausen H, Taubert H, Becker A, Kuhnt T, Hänsgen G, Dunst J and Vordermark D. Immunohistochemical detection of osteopontin in advanced head-and-neck cancer: prognostic role and correlation with oxygen electrode measurements, hypoxia-inducible-factor-1alpha-related markers, and hemoglobin levels. *Int J Radiat Oncol Biol Phys* 2006; 66:1481-7.
283. Mhavec-Fauceglia, P. Value of ezrin, moesin and radixin nm23-H1 protein expressions in predicting outcome of patients with head and neck squamous cell carcinoma treated with radical radiotherapy. *J Clin Pathol* 2007; 60:185-89.
284. Köbel M, Gradhand E, Zeng K, Schmitt W, Kriese K, Lantzsch T, Wolters M, Dittmer J, Strauss H, Thomssen C and Hauptmann S. Ezrin promotes ovarian carcinoma cell invasion and its retained expression predicts poor prognosis in ovarian carcinoma. *Int J Gynecol Pathol* 2006; 25:121-30.
285. Bui M, Seligson D, Han K, Pantuck A, Dorey F, Huang Y, Horvath S, Leibovich B, Chopra S, Liao S, Stanbridge E, Lerman M, Palotie A, Figlin RA and Belldegrun AS. Carbonic anhydrase IX is an independent predictor of survival in advanced renal clear cell carcinoma: implications for prognosis and therapy. *Clin Cancer Res* 2003; 9:802-11.

References

286. Bui M, Visapää H, Seligson D, Kim H, Han K, Huang Y, Horvath S, Stanbridge E, Palotie A, Figlin R and Belldegrün A. Prognostic value of carbonic anhydrase IX and KI67 as predictors of survival for renal clear cell carcinoma. *J Urol* 2004; 171:2461-6.
287. Atkins M, Regan M, McDermott D, Mier J, Stanbridge E, Youmans A, Febbo P, Upton M, Lechpammer M and Signoretti S. Carbonic anhydrase IX expression predicts outcome of interleukin 2 therapy for renal cancer. *Clin Cancer Res* 2005; 11:3714-21.
288. Patard J, Fergelot P, Karakiewicz P, Klatter T, Trinh Q, Rioux-Leclercq N, Said J, Belldegrün A and Pantuck A. Low CAIX expression and absence of VHL gene mutation are associated with tumor aggressiveness and poor survival of clear cell renal cell carcinoma. *Int J Cancer* 2008; 123:395-400.
289. Giatromanolaki A, Koukourakis M, Sivridis E, Pastorek J, Wykoff C, Gatter K and Harris A. Expression of hypoxia-inducible carbonic anhydrase-9 relates to angiogenic pathways and independently to poor outcome in non-small cell lung cancer. *Cancer Res* 2001; 61:7992-8.
290. Swinson D, Jones J, Richardson D, Wykoff C, Turley H, Pastorek J, Taub N, Harris A and O'Byrne K. Carbonic anhydrase IX expression, a novel surrogate marker of tumor hypoxia, is associated with a poor prognosis in non-small-cell lung cancer. *J Clin Oncol* 2003; 21:473-82.
291. Mäkitie T, Carpén O, Vaheri A and Kivelä T. Ezrin as a prognostic indicator and its relationship to tumor characteristics in uveal malignant melanoma. *Invest Ophthalmol Vis Sci* 2001; 42:2442-9.
292. Generali D, Berruti A, Brizzi M, Campo L, Bonardi S, Wigfield S, Bersiga A, Allevi G, Milani M, Aguggini S, Gandolfi V, Dogliotti L, Bottini A, Harris AL and Fox SB. Hypoxia-inducible factor-1alpha expression predicts a poor response to primary chemoendocrine therapy and disease-free survival in primary human breast cancer. *Clin Cancer Res* 2006; 12:4562-8.
293. Kawamura T, Kusakabe T, Sugino T, Watanabe K, Fukuda T, Nashimoto A, Honma K and Suzuki T. Expression of glucose transporter-1 in human gastric carcinoma: association with tumor aggressiveness, metastasis, and patient survival. *Cancer* 2001; 92:634-41.
294. Hoskin P, Sibtain A, Daley F and Wilson G. GLUT1 and CAIX as intrinsic markers of hypoxia in bladder cancer: relationship with vascularity and proliferation as predictors of outcome of ARCON. *Br J Cancer* 2003; 89:1290-7.
295. Böhling T, Turunen O, Jääskeläinen J, Carpen O, Sainio M, Wahlström T, Vaheri A and Haltia M. Ezrin expression in stromal cells of capillary hemangioblastoma. An immunohistochemical survey of brain tumors. *Am J Pathol* 1996; 148:367-73.
296. Westermark B, Pontén J and Hugosson R. Determinants for the establishment of permanent tissue culture lines from human gliomas. *Acta Pathol Microbiol Scand A* 1973; 81:791-805.
297. Sainio M, Zhao F, Heiska L, Turunen O, den Bakker M, Zwarthoff E, Lutchman M, Rouleau G, Jääskeläinen J, Vaheri A and Carpén O. Neurofibromatosis 2 tumor suppressor protein colocalizes with ezrin and CD44 and associates with actin-containing cytoskeleton. *J Cell Sci* 1997; 110 (Pt 18):2249-60.
298. Flørenes V, Maelandsmo G, Forus A, Andreassen A, Myklebost O and Fodstad O. MDM2 gene amplification and transcript levels in human sarcomas: relationship to TP53 gene status. *J Natl Cancer Inst* 1994; 86:1297-302.
299. Gomez-Manzano C, Fueyo J, Kyritsis A, Steck P, Roth J, McDonnell T, Steck K, Levin V and Yung W. Adenovirus-mediated transfer of the p53 gene produces rapid and generalized death of human glioma cells via apoptosis. *Cancer Res* 1996; 56:694-9.

References

300. Talvinen K, Tuikkala J, Grönroos J, Huhtinen H, Kronqvist P, Aittokallio T, Nevalainen O, Hiekkänen H, Nevalainen T and Sundström J. Biochemical and clinical approaches in evaluating the prognosis of colon cancer. *Anticancer Res* 2006; 26:4745-51.
301. Opavský R, Pastoreková S, Zelník V, Gibadulinová A, Stanbridge E, Závada J, Kettmann R and Pastorek J. Human MN/CA9 gene, a novel member of the carbonic anhydrase family: structure and exon to protein domain relationships. *Genomics* 1996; 33:480-7.
302. Wykoff C, Beasley N, Watson P, Turner K, Pastorek J, Sibtain A, Wilson G, Turley H, Talks K, Maxwell P, Pugh C, Ratcliffe P and Harris A. Hypoxia-inducible expression of tumor-associated carbonic anhydrases. *Cancer Res* 2000; 60:7075-83.
303. Abrey L. Concomitant chemoradiotherapy followed by adjuvant temozolomide improves survival in glioblastoma multiforme. *Curr Neurol Neurosci Rep* 2005; 5:167-8.
304. Span PN, Nagtegaal ID and Bussink J. Expression of carbonic anhydrase IX suggests poor response to therapy in rectal cancer. *Br J Cancer* 2009;101:372; author reply 3.
305. Järvelä S, Parkkila S, Bragge H, Kähkönen M, Parkkila A, Soini Y, Pastorekova S, Pastorek J and Haapasalo H. Carbonic anhydrase IX in oligodendroglial brain tumors. *BMC Cancer* 2008; 8:1.
306. Stillebroer A, Mulders P, Boerman O, Oyen W and Oosterwijk E. Carbonic anhydrase IX in renal cell carcinoma: implications for prognosis, diagnosis, and therapy. *Eur Urol* 2010; 58:75-83.
307. Jain RK, Tong RT and Munn LL. Effect of vascular normalization by antiangiogenic therapy on interstitial hypertension, peritumor edema, and lymphatic metastasis: insights from a mathematical model. *Cancer Res* 2007; 67:2729-35.
308. Niu G, Briggs J, Deng J, Ma Y, Lee H, Kortylewski M, Kujawski M, Kay H, Cress W, Jove R and Yu H. Signal transducer and activator of transcription 3 is required for hypoxia-inducible factor-1alpha RNA expression in both tumor cells and tumor-associated myeloid cells. *Mol Cancer Res* 2008; 6:1099-105.
309. Semenza G. HIF-1 and tumor progression: pathophysiology and therapeutics. *Trends Mol Med* 2002; 8:S62-7.
310. Vaupel P, Schlenger K, Knoop C and Höckel M. Oxygenation of human tumors: evaluation of tissue oxygen distribution in breast cancers by computerized O₂ tension measurements. *Cancer Res* 1991; 51:3316-22.
311. Joseph M and Benson Ar. Current issues in rectal cancer chemotherapy. *Cancer J*;13:198-203.
312. Brizel D, Scully S, Harrelson J, Layfield L, Bean J, Prosnitz L and Dewhirst M. Tumor oxygenation predicts for the likelihood of distant metastases in human soft tissue sarcoma. *Cancer Res* 1996; 56:941-3.
313. Moeller BJ, Dreher MR, Rabbani ZN, Schroeder T, Cao Y, Li CY and Dewhirst MW. Pleiotropic effects of HIF-1 blockade on tumor radiosensitivity. *Cancer Cell* 2005; 8:99-110.
314. Rademakers S, Span P, Kaanders J, Sweep F, van der Kogel A and Bussink J. Molecular aspects of tumour hypoxia. *Mol Oncol* 2008; 2:41-53.
315. Tsukita S and Yonemura S. ERM (ezrin/radixin/moesin) family: from cytoskeleton to signal transduction. *Curr Opin Cell Biol* 1997; 9:70-5.
316. Micke P and Ostman A. Tumour-stroma interaction: cancer-associated fibroblasts as novel targets in anti-cancer therapy? *Lung Cancer* 2004;45 Suppl 2:S163-75.

References

317. Madri J, Bell L, Marx M, Merwin J, Basson C and Prinz C. Effects of soluble factors and extracellular matrix components on vascular cell behavior in vitro and in vivo: models of de-endothelialization and repair. *J Cell Biochem* 1991; 45:123-30.
318. DeClerck Y, Mercurio A, Stack M, Chapman H, Zutter M, Muschel R, Raz A, Matrisian L, Sloane B, Noel A, Hendrix M, Coussens L and Padarathasingh M. Proteases, extracellular matrix, and cancer: a workshop of the path B study section. *Am J Pathol* 2004; 164:1131-9.
319. Atkin G, Daley F, Bourne S, Glynne-Jones R, Northover J and Wilson G. The impact of surgically induced ischaemia on protein levels in patients undergoing rectal cancer surgery. *Br J Cancer* 2006; 95:928-33.
320. Pelicano H, Martin D, Xu R and Huang P. Glycolysis inhibition for anticancer treatment. *Oncogene* 2006; 25:4633-46.
321. Vera JC, Reyes AM, Velásquez FV, Rivas CI, Zhang RH, Strobel P, Slebe JC, Núñez-Alarcón J and Golde DW. Direct inhibition of the hexose transporter GLUT1 by tyrosine kinase inhibitors. *Biochemistry* 2001; 40:777-90.
322. Rastogi S, Banerjee S, Chellappan S and Simon G. Glut-1 antibodies induce growth arrest and apoptosis in human cancer cell lines. *Cancer Lett* 2007; 257:244-51.