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Factors influencing the microbial composition of human milk

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ABSTRACT

Aside from nutritional components, human milk is rich in microorganisms. Through breastfeeding these microorganisms are introduced to the infant gut where they may transiently or persistently colonize it. Therefore, the human milk microbiota may be an important factor which shapes the infant gut microbiota further influencing infant health and disease. In the current review we aim to give a brief updated insight into the putative origin of the human milk microbiota, its constituents and the possible factors that shape it. Understanding the factors that determine the human milk microbiota composition and function will aid developing optimal postnatal feeding and intervention strategies to reduce the risk of communicable and noncommunicable diseases.

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Introduction

Early life nutrition is associated with the risk of developing non-communicable diseases (NCDs) including obesity, cardiovascular and chronic respiratory diseases, cancers, and diabetes. While NCDs are now considered the leading causes of death worldwide evidence has shown that breastfeeding can exert beneficial effects for both mother and infant. For instance, breastfeeding may protect mothers from ovarian cancer, breast cancer and diabetes and offer long-term maternal cardiovascular health benefits.^{1,2} On the other hand, prolonged breastfeeding for at least eight months has been associated with low average blood glucose levels in infants born to mothers with gestational diabetes mellitus.³ In addition, multiple studies have highlighted that the use of human milk (HM) has a positive effect on various infant diseases and conditions including

respiratory infections and diarrhea as well as necrotizing enterocolitis in preterm neonates.

This protective role of breastfeeding might be mediated through effects on the infant gut microbiome. Increasing evidence links changes in the composition of the intestinal microbiota with adverse health outcomes. Therefore, the development of the gut microbiome during infancy is essential for the maturation and function of the infant's immune system. However, this development is largely modulated by the mode of delivery, perinatal use of antibiotics and infant diet among other factors. Indeed HM is not only viewed as the ideal source of nutrients for infants but also contains a variety of compounds that might affect infant immunity (i.e., human milk oligosaccharides, antibodies, cytokines, human cells and extracellular vesicles).^{4,5} Besides essential nutrients and bioactive molecules, HM although at first thought sterile, is found to contain commensal microbes, which have the ability to modulate the colonization of the infant gut. Several

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studies have discussed the overlap between human milk and infant gut microbiota. Of special interest is the genus *Bifidobacterium*, which has been identified in both niches and is considered a dominant constituent of the gut microbiota of breastfed infants.⁶ Generally, lower abundance of this genus in the infant gut could trigger the overgrowth of *Bacteroides*, which has been associated with the risk of developing of allergies and obesity later in life.⁷ The composition of the human milk microbiota (HMM) may play an essential role in shaping the infant gut microbiome and therefore it is important to identify the factors that influence the composition of the human milk microbiome (Figure 1).

Origin of human milk microbiota

Human milk has for long been considered sterile, however over the past decade various reports describe the presence of viable bacteria in milk produced by healthy women free from infection. Currently, human milk is regarded as a continuous source of microbes with more than several hundred different bacterial species identified.⁸ Initially, the presence of microbes in milk was thought to be a product of contamination, due to vaginal exposure of the neonate or due to mother's skin and infant's mouth contact.⁹

Three different hypotheses have been proposed to explain how these microbes found themselves in milk: the entero-mammary pathway, the retrograde inoculation pathway and the notion of resident mammary microbiota. The entero-mammary pathway is an endogenous route that is described by translocation of maternal gut bacteria from the gut to the mammary glands. Through this pathway, dendritic cells (and possibly macrophages) can introduce openings in the tight junctions between the epithelial cells of the gut and trap bacteria through dendrites. It is possible that hormonal changes during late pregnancy create a favoring environment for immune cells to open the tight junctions in the intestinal epithelium. Dendritic cells then travel through the lymphatic and blood circulation and finally reach the mammary ducts where they release the bacteria in the milk.⁹

Alternatively, microorganisms present in the milk of healthy women might not only originate from the maternal gut but also from the maternal skin, infant oral cavity or even the environment. For instance, some bacterial genera of the skin, such as *Staphylococcus*, *Corynebacterium* and *Propionibacterium*, were also frequently found in breast milk.¹⁰ Retrograde backflow during breastfeeding may also play a role in the establishment of oral bacteria in human milk. Indeed, human milk and the infant oral cavity were found to share several bacterial genera such as *Streptococcus*, *Veillonella* and *Variovorax*.^{11,12} Nonetheless, typical oral bacterial colonizers were

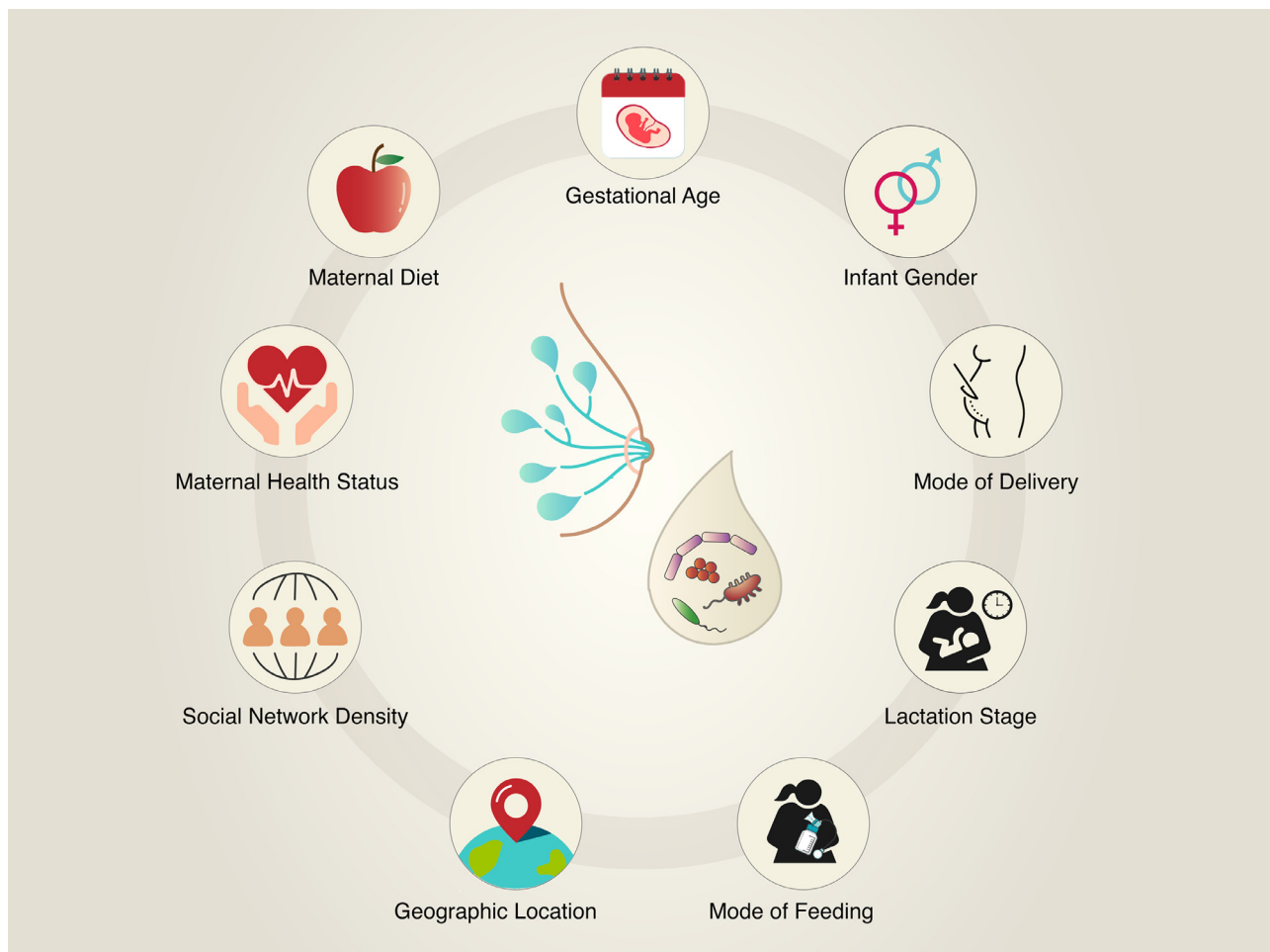


Fig. 1 – Factors influencing the microbial composition of human milk.

found in precolostrum before delivery when the breast had not yet been exposed to the infant's mouth.¹³ In addition, the retrograde inoculation pathway can not fully explain the fact that some bacteria in human milk are strictly anaerobic (eg. *Bifidobacterium* and *Faecalibacterium* strains). These observations suggest the possibility that both pathways may be potential sources of the human milk microbiota although the mechanisms are not yet clear.¹⁴ Finally, it has long been known that mammary tissue of non-lactating women has its own microbiota although previously described as similar to the one on the skin. Specifically, in their recent systematic review, Togo and colleagues identified seven species including *C. acnes*, *S. epidermidis* and *S. agalactiae* that are also present in human milk supporting the thought of a commensal mammary microbiota that could affect the HMM composition.¹¹

Human milk microbiota

The presence of bacteria in human milk has been explored in a number of studies using culture-dependent and culture-independent techniques. While culture-dependent methods enable the identification of viable and culturable bacteria in human milk, they also lead to an underestimation of the total bacterial count since dormant or unculturable microbes remain undetected. To overcome this limitation culture-independent techniques, such as quantitative polymerase chain reaction (qPCR) and next generation sequencing (NGS) have been employed. These molecular approaches have revealed that the diversity and the inter-individual variability of HMM is higher than previously thought. Nevertheless, a recent systematic review highlighted the importance of using a combination of the afore mentioned methodologies to accurately assess the HMM composition.¹¹ In short, molecular methods allowed the detection of twice the number of bacterial species compared to culture methods with only 26% species detected by both approaches. In addition, 62% species were found only by molecular methods, while 12% were found only by traditional culturing.

Despite these variations, both methods have shown that human milk is characterized by a core microbiota comprised of 7-9 taxa depending on the selected methodology and study population.^{15,16} The majority of performed studies have identified *Staphylococcus* and *Streptococcus* as the most consistently and frequently observed genera in human milk. Interestingly, breast-fed infants also have higher levels of *Staphylococcus* in their feces compared to formula-fed ones.¹⁷ While growing in the gut, *Staphylococcus* and *Streptococcus* utilize oxygen creating a favorable environment for the proliferation of beneficial anaerobic bacteria. A few anaerobes which are also detected in human milk include *Bifidobacterium*, *Faecalibacterium* and *Akkermansia* and may reduce the risk of dysbiosis-related diseases through the production of short chain fatty acids. For instance, *Bifidobacterium* and *Akkermansia muciniphila* produce acetate, which may help prevent pathogenic infection while *Faecalibacterium prausnitzii* produces butyrate, which can modulate inflammation.^{18,19} Other identified genera include lactic acid bacteria (i.e., *Lactobacillus*), skin (i.e., *Propionibacterium*, *Cutibacterium* and *Corynebacterium*) and gut commensals

(i.e. *Acinetobacter*, *Bacteroides*, *Blautia*, *Clostridium*, *Dorea*, *Enterococcus*, *Escherichia*, *Mucispirillum*, *Pseudomonas*, *Rothia*, *Salmonella*, *Serratia*, *Shigella*, and *Veillonella*).^{7,11} Further characterization of the HMM has revealed the presence of fungi (*Malassezia*, *Candida*, *Saccharomyces*, *Rhodotorula*, *Davidiella*, *Sistotrema*, and *Penicillium*), archaea (*Haloarcula*, *Halorhabdus*, and *Halomicrobium*), protozoa (*Giardia* and *Toxoplasma*), and viruses in the milk of healthy mothers.^{15,20,21} Specifically regarding viruses, HMM of healthy mothers consists mainly of lytic bacteriophages (*Myoviridae*, *Podoviridae* and *Siphoviridae*) and a minority of eukaryotic viruses (*Phycodnaviridae*, *Iridoviridae* and *Imiviridae*).²² Lytic bifidophages in particular have been shown to suppress the growth of dominant *Bifidobacterium* in the gut.²³ Thus, the presence of bacteriophages could not only affect the bacterial diversity and relative abundance in human milk but also in the gut.

Factors shaping the human milk microbiota

Maternal health status

While obesity has mainly been considered a problem of developed countries, its prevalence has now reached global epidemic proportions. Besides being one of the major risk factors for premature death, maternal obesity is a major risk factor for childhood obesity. Although the mechanisms behind this are poorly understood, differences between HMM diversity and composition between obese and normal weight mothers may play a role. Indeed, overweight and obese mothers reportedly produce milk with a lower microbial diversity and lower counts of *Bifidobacterium* and a higher level of *Staphylococcus*, *Akkermansia* and *Granulicatella* than normal-weight women.^{8,24,25} Obese mothers have also been observed to display a higher incidence of *Staphylococcus*, *Corynebacterium* and *Brevundimonas* in their milk when compared to overweight or normal weight mothers²⁶ Consequently, these changes may affect the human milk metabolome and further influence the infant microbiome development. For instance, a recent study showed that human milk metabolite levels differ significantly between overweight or obese and normal weight mothers.²⁷ Specifically, milk hormones such as insulin and leptin are found in elevated levels in milk of obese mothers and may impact the development of the infant intestinal microbiome.²⁸ Gestational prehypertensive status is another factor that has been reported to contribute to lower bacterial diversity and lower relative abundance of *Lactobacillus* in human milk.²⁹ If this translates to colonization of the infant gut with lower numbers of *Lactobacillus*, it could limit the protective functions associated with this genus.

HMM composition is influenced by chronic illnesses such as celiac disease and allergic disorders. Recently, Spanish researchers found that the milk of healthy mothers was richer in *Bifidobacterium* and *Bacteroides fragilis* and had higher concentration of the tolerogenic cytokine TGF- β 1 than mothers with celiac disease.³⁰ Higher TGF- β 1 levels in milk during the first month of lactation, have been associated with reduced risk of eczema in infants.³¹ Milk of healthy mothers was also found to be richer in *Bifidobacterium* compared to the milk of allergic mothers.³² It has been suggested that the immune responses associated with these chronic diseases could be responsible for the lower levels

of *Bifidobacterium* in the milk of mothers with celiac disease or allergic disorders.^{30,33}

A primary concern for women in the postpartum period is maternal postnatal distress, which is characterized by symptoms of depression or anxiety. Mild postpartum depressive symptoms have been associated with increased cortisol levels, which could alter the fecal microbiota diversity through the recently proposed gut-brain axis.^{34,35} In accordance with the enteromammary pathway hypothesis, this could also influence the HMM composition. Indeed, in a previous study, Browne and coworkers found that maternal postnatal distress can also influence the HMM composition.³⁶ Mothers with high psychosocial distress had a less diverse HMM at 3 months postpartum compared to those with low psychosocial distress. Moreover, both groups saw a significant decrease in the relative abundance of *Staphylococcus*, while milk from mothers in the low group had a significant increase in the relative abundance of *Lactobacillus*, *Acinetobacter*, and *Flavobacterium*. Finally, no significant changes in the relative abundance of Actinobacteria (eg. *Bifidobacterium*) were noted. Mothers who adhered to postpartum confinement practices such as “doing-the-month” programs in Taiwan had lower postpartum physical and depressive symptoms and a higher prevalence of *Lactobacillus*, *Bifidobacterium* and Archaea in their milk compared to mothers who did not.^{37,38} However, these are complex 20–30 day programs, which include a unique combination of dietary and herbal therapies and it is still not yet clear which factor contributes to this modulation of the HMM.

Other important factors that seem to shape the HMM are bacterial and viral infections. For example, lactational mastitis and breast abscess, which may develop as a complication of mastitis, most often caused by *Staphylococcus aureus*, have been associated with dominance of *S. aureus*, complete absence of *Bifidobacterium breve* and overall lower bacterial diversity in the milk of infected mothers.^{11,15} In addition, *Salmonella enterica* and *Burkholderia ambifaria* are only detected in human milk of mothers with breast abscess, while several *Bifidobacteria* and *Lactobacilli* are associated with absence of both mastitis and breast abscess. Lower diversity, increased bacterial loads, and altered microbiota composition were also confirmed in human milk of mothers with sub-acute mastitis by Amorós and colleagues.³⁹

Changes in HMM composition triggered by viral infections have also been reported. A study on neonatal rotavirus (RV) infection in India, showed that the milk of mothers of symptomatic infants had a higher relative abundance of *Enterobacter/Klebsiella*. In contrast, the milk of mothers of asymptomatic and RV negative infants was dominated by *Staphylococcus* and *Streptococcus*.⁴⁰ Increased abundance of *Staphylococcus* was also noted in the milk of healthy African mothers when compared to the milk of HIV infected mothers that was more diverse and had higher counts of *Lactobacillus*.⁴¹ In conclusion, accumulating evidence suggests that eukaryotic viruses often directly and indirectly interact with bacteria and thus, their presence in human milk may not only modulate its virome but also work synergistically with the milk bacteriome.^{42,43}

Antibiotic therapy is common during pregnancy and particularly during delivery either as prophylaxis to protect the

mother and her infant or treatment of infection. Numerous reports indicate that the use of antibiotics has significant effects on several HMM parameters. Recently, Hermansson and coworkers described an increase in the levels of different bacterial species and their diversity in the milk of mothers who received antibiotics during delivery.⁴⁴ In addition, previous studies have reported lower relative abundance of *Lactobacillus* and *Bifidobacterium* in the milk of mothers who received antibiotics during pregnancy or lactation.^{45,46} Besides antibiotherapy, chemotherapy for the treatment of malignancies can also modulate HMM composition. Before and after treatment analysis of HMM of a woman with Hodgkin's lymphoma revealed reduced levels of certain genera, especially *Bifidobacterium*, *Eubacterium*, *Staphylococcus* and *Cloacibacterium*. In contrast, when compared to the milk of healthy mothers, the milk collected during chemotherapy was characterized by a significant increase of *Acinetobacter* and *Xanthomonadaceae*.⁴⁷

Maternal diet

It is well known that maternal diet can influence the bacterial taxonomic composition of human milk since many of its nutrients may be utilized by bacteria. For example, a study on healthy lactating mothers from Brazil showed that the presence of *Staphylococcus* in their milk was positively correlated with Vitamin C intake during pregnancy. During the lactation period, increased sugar intake is associated with a decrease of *Pseudomonas* while an increased Vitamin B9 intake is associated with an increase its numbers in milk. Another positive correlation was found between *Bifidobacterium* and the intake PUFAs and linoleic acid.⁴⁸ Conversely, Kumar and coworkers found a negative correlation between *Bifidobacterium* and *Lactobacillus* and MUFA and n-3 PUFA in milk phospholipids. In addition, in the same study, saturated fatty acids were negatively associated with *Corynebacterium* and *Streptococcus*.^{25,49} Moreover, greater protein consumption was related to an increased abundance of *Gemella*, *Bacillus*, *Peptoniphilus*, and *Anaerococcus*.^{25,50} It is also possible that other milk components are able to modulate its microbial composition. For example, concentrations of the polyamine putrescine are correlated with the levels of Gammaproteobacteria and a strongly negative correlation with other Proteobacteria, Clostridia and Actinobacteria.⁵¹ A recent *in vitro* study indicated that *Bifidobacteria* which utilize specific human milk oligosaccharides (HMOs) can enhance the growth of non-HMO degrading *Bifidobacteria*.⁵² This could potentially lead to their dominance in human milk and thus affect its microbial composition. In line with this notion, correlations between human milk HMO composition and bacteria including *Bifidobacteria* have been reported.⁵³ It is also important to bear in mind that maternal secretor status profoundly affects the specific HMO content of milk. The milk of secretor mothers was found to contain a specific HMO composition distinct from secretor mothers. This observation could also partially explain why infants of secretor mothers have higher bifidobacterial abundance in their gut microbiota.⁵⁴

Finally, a number of studies indicates that although pre and postnatal maternal administration of specific probiotics can result in their presence in human milk, the intervention does

not affect HMM diversity and composition.^{44,55(p12)} In contrast, other probiotic strains, were shown to be effective in reducing the numbers of *S. aureus* in the milk of mothers with mastitis or increasing the relative abundance of lactobacilli and bifidobacteria.^{56,57} This difference could be due to the ability of human milk isolated probiotics to directly metabolize HMOs or other factors such as mode of delivery.

Gestational age, infant gender and mode of delivery

Preterm infants have immune systems characterized by developmental immaturities. This makes them susceptible to detrimental infections and necrotizing enterocolitis (NEC). Interestingly, preterm infants fed with human milk instead of infant formula are less likely to develop NEC. A potential mechanism behind this finding is that some beneficial human milk bacteria may colonize the infant gut, assisting in the development of the immune system and thus provide protection against infections.⁵⁸ However, significant differences in the HMM according to the gestational age have been reported, with higher levels of *Enterococcus* and lower levels of *Bifidobacterium* observed in the milk of preterm-delivering mothers compared to those who gave birth on the expected birth date.⁵⁹ Nonetheless, despite differences in the relative abundances, *Bifidobacterium*, *Lactobacillus*, *Staphylococcus*, *Streptococcus* and *Enterococcus*, were all present in milk from mothers who had delivered preterm or at full term. When examining the different degrees of prematurity, the same group detected an overall lower bacterial load in the milk of mothers of extremely preterm as compared with those of late preterm infants. On the other hand, Urbaniak and colleagues did not detect any changes in the HMM composition correlating with gestational age.⁶⁰ Similar contrasting results have been reported for infant gender, which in some studies seemed to affect HMM composition, but in others no correlation was observed. For instance, a higher relative abundance of *Rothia* was detected in the milk from mothers of female infants compared with mothers of male infants.^{55(p12)} In contrast, Urbaniak and coworkers, found no differences in microbial profiles based on gender of the infant.⁶⁰ Although the exact cause of these inconsistencies is unknown, different collection practices, sample sizes and analytical methods could be considered possible factors.

Mode of delivery is reportedly associated with profound compositional changes of the HMM. Although human milk from C-section mothers is correlated with higher total bacterial counts (especially *Streptococcus*, *Proteobacteria* and *Carnobacteriaceae*), it is less diverse with lower levels of *Bifidobacterium*, *Lactobacillus* and *Leuconostocaceae* compared to the milk of mothers who underwent a vaginal delivery.^{8,44} Conversely, another report showed no association between the composition of the HMM and delivery mode.⁶⁰

Lactation stage and mode of feeding

It has long been observed that the composition of human milk adapts to the immediate needs of the infant and is divided in three distinct stages: colostrum, transitional milk, and mature milk. The microbiota during the first stage, colostrum, is characterized by increased bacterial diversity with

predominant bacteria belonging to the *Weisella*, *Leuconostoc*, *Staphylococcus*, *Streptococcus*, and *Lactococcus* genus.⁸ As lactation progresses, total bacterial concentration increases while bacterial diversity decreases. In transitional and mature milk, *Bifidobacterium*, *Enterococcus*, *Veillonella*, *Leptotrichia*, *Prevotella*, *Lactobacillus* and *Staphylococcus* exhibit an upward trend in their relative abundances when compared to colostrum.⁵⁹ This variation across the lactation period may be partly explained by the retrograde inoculation pathway since increased abundance of typical oral bacteria has been reported in milk from the later stages. Indeed, levels of oral bacteria in milk (mainly *Streptococcus* and *Rothia*), are on the rise after each breastfeeding session accompanied by an increased bacterial diversity.⁶¹

Differences in microbial composition have been observed between the two breasts of the same mother.⁶² Regarding the milk collection protocol, mothers who expressed milk through a pump had lower abundance of cultivable staphylococci and higher content of bacterial DNA than mothers who expressed their milk manually.⁶³ Moreover, mothers who fed their infants pumped milk had lower abundance of *Bifidobacterium*, and higher abundance of *Enterobacteriaceae* and *Pseudomonadaceae* in their milk.⁶⁴ Conversely, mothers who fed their infants through direct breastfeeding had higher numbers of *Gemellaceae*, *Vogesella*, and *Nocardioides*.

Geographic location and social network density

Various studies have explored the impact of geographic location on the HMM composition. In a recent pilot study, the HMM of the Old Order Mennonites (OOMs) population was found to be more diverse compared to the one of mothers from urban areas. This could be due to the different dietary and environmental exposure patterns of this community, mainly characterized by cultivation of own food, consumption of raw milk and preference for home births.⁶⁵ Similarly, population differences were also found in Iran, with total counts of *Lactobacillus* being higher in the milk of mothers residing in rural areas compared to those in urban areas.⁶⁶ Meehan and colleagues examined the HMM of hunter-gatherer and horticulturalist women in the Central African Republic, reporting lower levels of *Lactobacillus* in the milk of hunter-gatherers than horticulturalists. The most abundant genera were similar to the ones found in the milk of Western women.⁶⁷ However, despite the overlap of several genera, lower relative abundances of *Bifidobacterium*, *Propionibacterium*, *Veillonella* and *Serratia* were reported in the milk of Central African Republic mothers compared to US or Swiss mothers.^{16,68} In addition, in the same study, the social and caregiving environment showed a significant association with HMM diversity with high HMM diversity exhibiting a positive correlation with a simultaneous increase in allomaternal and decrease in maternal care. Another study compared the HMM of ethnically distinct mothers.⁴⁹ When the HMM composition was studied in mothers who had delivered vaginally, Spanish mothers had the highest relative abundance of Bacteroidetes in milk when compared to mothers from Finland, China or South Africa. In the same study, Chinese women who delivered by C-section harbored the highest levels of Actinobacteria in their milk compared to their

European or African counterparts. Finally, and in accordance with the previous reports, Lackey and colleagues showed that the composition of HMM varies significantly within and across cohorts from international sites (Ethiopia, Gambia, US, Kenya, Peru, Spain, and Sweden).⁶⁹

Conclusion

Several factors including maternal health status and diet, gestational age, infant gender, mode of delivery, lactation stage, mode of feeding, geographic location and social network density influence the composition of the human milk microbiota and therefore which microorganisms are transferred to the infant through breastfeeding. However, many questions still remain unanswered not only related to the HMM composition but also its function. Lastly, breastfeeding is not always possible and, in many cases, banked milk is the next optimal option. Therefore, further research is needed to determine how factors associated with human milk banking such as storage and pasteurization affect the composition of these microbial communities.

Disclosures

The authors report no proprietary or commercial interest in any product mentioned or concept discussed in this article.

REFERENCES

- Binh Nguyen, Joanne Gale, Natasha Nassar, Adrian Bauman, Grace Joshy, Ding Ding. Breastfeeding and Cardiovascular Disease Hospitalization and Mortality in Parous Women: Evidence From a Large Australian Cohort Study. *J Am Heart Assoc.* 2019;8(6):e011056.
- Victoria CG, Bahl R, Barros AJD, et al. Breastfeeding in the 21st century: epidemiology, mechanisms, and lifelong effect. *The Lancet.* 2016;387(10017):475–490.
- Dugas C, Kearney M, Mercier R, et al. Early life nutrition, glycemic and anthropometric profiles of children exposed to gestational diabetes mellitus in utero. *Early Hum Dev.* 2018;118:37–41.
- Kim SY, Yi DY. Components of human breast milk: from macronutrient to microbiome and microRNA. *J Korean Pediatr Soc.* March 23, 2020 Published online.
- Kim SY, Yi DY. Analysis of the Human Breast Milk Microbiome and Bacterial Extracellular Vesicles in Healthy Mothers. Published online April 23, 2020.
- Rautava S. Early microbial contact, the breast milk microbiome and child health. *J Dev Orig Health Dis.* 2016;7(1):5–14.
- Zimmermann P, Curtis N. Breast milk microbiota: A review of the factors that influence composition. *J Infect.* February 6, 2020 Published online.
- Cabrera-Rubio R, Collado MC, Laitinen K, Salminen S, Isolauri E, Mira A. The human milk microbiome changes over lactation and is shaped by maternal weight and mode of delivery. *Am J Clin Nutr.* 2012;96(3):544–551.
- Rodriguez JM. The Origin of Human Milk Bacteria: Is There a Bacterial Entero-Mammary Pathway during Late Pregnancy and Lactation? *Adv Nutr Int Rev J.* 2014;5(6):779–784.
- Lee J-E, Kim G-B. Human Milk Microbiota: A Review. *J Milk Sci Biotechnol.* 2019;37(1):15–26.
- Togo A, Dufour J-C, Lagier J-C, Dubourg G, Raoult D, Million M. Repertoire of human breast and milk microbiota: a systematic review. *Future Microbiol.* 2019;14(7):623–641.
- Murphy K, Curley D, O’Callaghan TF, et al. The Composition of Human Milk and Infant Faecal Microbiota Over the First Three Months of Life: A Pilot Study. *Sci Rep.* 2017;7.
- Ruiz L, Bacigalupe R, García-Carral C, et al. Microbiota of human precolostrum and its potential role as a source of bacteria to the infant mouth. *Sci Rep.* 2019;9(1):8435.
- Gueimonde M, Laitinen K, Salminen S, Isolauri E. Breast milk: a source of bifidobacteria for infant gut development and maturation? *Neonatology.* 2007;92(1):64–66.
- Jiménez E, de Andrés J, Manrique M, et al. Metagenomic Analysis of Milk of Healthy and Mastitis-Suffering Women. *J Hum Lact.* 2015;31(3):406–415.
- Hunt KM, Foster JA, Forney LJ, et al. Characterization of the Diversity and Temporal Stability of Bacterial Communities in Human Milk. *PLOS ONE.* 2011;6(6):e21313.
- Balmer SE, Wharton BA. Diet and faecal flora in the newborn: breast milk and infant formula. *Arch Dis Child.* 1989;64(12):1672–1677.
- Belzer C, de Vos WM. Microbes inside—from diversity to function: the case of Akkermansia. *ISME J.* 2012;6(8):1449–1458.
- Martín R, Bermúdez-Humarán LG, Langella P. Searching for the Bacterial Effector: The Example of the Multi-Skilled Commensal Bacterium *Faecalibacterium prausnitzii*. *Front Microbiol.* 2018;9.
- Boix-Amorós A, Martínez-Costa C, Querol A, Collado MC, Mira A. Multiple Approaches Detect the Presence of Fungi in Human Breastmilk Samples from Healthy Mothers. *Sci Rep.* 2017;7(1):13016.
- Boix-Amorós A, Puente-Sánchez F, du Toit E, et al. Mycobiome Profiles in Breast Milk from Healthy Women Depend on Mode of Delivery, Geographic Location, and Interaction with Bacteria. *Appl Environ Microbiol.* 2019;85(9).
- Pannaraj PS, Ly M, Cerini C, et al. Shared and Distinct Features of Human Milk and Infant Stool Viromes. *Front Microbiol.* 2018;9.
- Lugli GA, Milani C, Turroni F, et al. Prophages of the genus Bifidobacterium as modulating agents of the infant gut microbiota. *Environ Microbiol.* 2016;18(7):2196–2213.
- Collado MC, Laitinen K, Salminen S, Isolauri E. Maternal weight and excessive weight gain during pregnancy modify the immunomodulatory potential of breast milk. *Pediatr Res.* 2012;72(1):77–85.
- Williams JE, Carrothers JM, Lackey KA, et al. Human Milk Microbial Community Structure Is Relatively Stable and Related to Variations in Macronutrient and Micronutrient Intakes in Healthy Lactating Women. *J Nutr.* 2017;147(9):1739–1748.
- LeMay-Nedjelski L, Butcher J, Ley SH, et al. Examining the relationship between maternal body size, gestational glucose tolerance status, mode of delivery and ethnicity on human milk microbiota at three months post-partum. *BMC Microbiol.* 2020:20.
- Isganaitis E, Venditti S, Matthews TJ, Lerin C, Demerath EW, Fields DA. Maternal obesity and the human milk metabolome: associations with infant body composition and postnatal weight gain. *Am J Clin Nutr.* 2019;110(1):111–120.
- Lemas DJ, Young BE, Baker PR, et al. Alterations in human milk leptin and insulin are associated with early changes in the infant intestinal microbiome. *Am J Clin Nutr.* 2016;103(5):1291–1300.
- Wan Y, Jiang J, Lu M, et al. Human milk microbiota development during lactation and its relation to maternal geographic location and gestational hypertensive status. *Gut Microbes.* 2020;11(5):1438–1449.
- Olivares M, Albrecht S, De Palma G, et al. Human milk composition differs in healthy mothers and mothers with celiac disease. *Eur J Nutr.* 2015;54(1):119–128.

31. Morita Y, Campos-Alberto E, Yamaide F, et al. TGF- β Concentration in Breast Milk is Associated With the Development of Eczema in Infants. *Front Pediatr*. 2018;6.
32. Grönlund M-M, Gueimonde M, Laitinen K, et al. Maternal breast-milk and intestinal bifidobacteria guide the compositional development of the Bifidobacterium microbiota in infants at risk of allergic disease. *Clin Exp Allergy J Br Soc Allergy Clin Immunol*. 2007;37(12):1764–1772.
33. Dixon D-L, Forsyth KD. Leukocytes in expressed breast milk of asthmatic mothers. *Allergol Immunopathol (Madr)*. 2017;45(4):325–332.
34. Garcia-Leal C, Rezende MGD, Corsi-Zuelli FM das G, Castro MD, Del-Ben CM. The functioning of the hypothalamic-pituitary-adrenal (HPA) axis in postpartum depressive states: a systematic review. *Expert Rev Endocrinol Metab*. 2017;12(5):341–353.
35. Jiang H, Ling Z, Zhang Y, et al. Altered fecal microbiota composition in patients with major depressive disorder. *Brain Behav Immun*. 2015;48:186–194.
36. Browne PD, Aparicio M, Alba C, et al. Human Milk Microbiome and Maternal Postnatal Psychosocial Distress. *Front Microbiol*. 2019;10.
37. Chen P-W, Kuo Y-H, Lin Y-L. The Impact of the Postpartum “Doing-the-Month” Practice on Human Milk Microbiota: A Pilot Study in Taiwan. *Microorganisms*. 2020;8(9).
38. Chien L-Y, Tai C-J, Ko Y-L, Huang C-H, Sheu S-J. Adherence to “Doing-the-month” practices is associated with fewer physical and depressive symptoms among postpartum women in Taiwan. *Res Nurs Health*. 2006;29(5):374–383.
39. Boix-Amorós A, Hernández-Aguilar MT, Artacho A, Collado MC, Mira A. Human milk microbiota in sub-acute lactational mastitis induces inflammation and undergoes changes in composition, diversity and load. *Sci Rep*. 2020;10.
40. Ramani S, Stewart CJ, Laucirica DR, et al. Human milk oligosaccharides, milk microbiome and infant gut microbiome modulate neonatal rotavirus infection. *Nat Commun*. 2018;9.
41. González R, Mandomando I, Fumadó V, et al. Breast Milk and Gut Microbiota in African Mothers and Infants from an Area of High HIV Prevalence. *PLoS ONE*. 2013;8(11).
42. Almand EA, Moore MD, Jaykus L-A. Virus-Bacteria Interactions: An Emerging Topic in Human Infection. *Viruses*. 2017;9(3).
43. Domínguez-Díaz C, García-Orozco A, Riera-Leal A, Padilla-Arellano JR, Fafutis-Morris M. Microbiota and Its Role on Viral Evasion: Is It With Us or Against Us? *Front Cell Infect Microbiol*. 2019;9.
44. Hermansson H, Kumar H, Collado MC, Salminen S, Isolauri E, Rautava S. Breast Milk Microbiota Is Shaped by Mode of Delivery and Intrapartum Antibiotic Exposure. *Front Nutr*. 2019;6.
45. Soto A, Martín V, Jiménez E, Mader I, Rodríguez JM, Fernández L. Lactobacilli and Bifidobacteria in Human Breast Milk: Influence of Antibiotherapy and Other Host and Clinical Factors. *J Pediatr Gastroenterol Nutr*. 2014;59(1):78–88.
46. Padilha M, Iaucci JM, Cabral VP, Diniz EMA, Taddei CR, Saad SMI. Maternal antibiotic prophylaxis affects Bifidobacterium spp. counts in the human milk, during the first week after delivery. *Benef Microbes*. 2019;10(2):155–163.
47. Urbaniak C, McMillan A, Angelini M, et al. Effect of chemotherapy on the microbiota and metabolome of human milk, a case report. *Microbiome*. 2014;2(1):24.
48. Padilha M, Danneskiold-Samsøe NB, Brejnrod A, et al. The Human Milk Microbiota is Modulated by Maternal Diet. *Microorganisms*. 2019;7(11).
49. Kumar H, du Toit E, Kulkarni A, et al. Distinct Patterns in Human Milk Microbiota and Fatty Acid Profiles Across Specific Geographic Locations. *Front Microbiol*. 2016;7.
50. Boix-Amorós A, Collado MC, Mira A. Relationship between Milk Microbiota, Bacterial Load, Macronutrients, and Human Cells during Lactation. *Front Microbiol*. 2016;7.
51. Gómez-Gallego C, Kumar H, García-Mantrana I, et al. Breast Milk Polyamines and Microbiota Interactions: Impact of Mode of Delivery and Geographical Location. *Ann Nutr Metab*. 2017;70(3):184–190.
52. Lawson MAE, O’Neill IJ, Kujawska M, et al. Breast milk-derived human milk oligosaccharides promote Bifidobacterium interactions within a single ecosystem. *ISME J*. 2020;14(2):635–648.
53. Aakko J, Kumar H, Rautava S, et al. Human milk oligosaccharide categories define the microbiota composition in human colostrum. *Benef Microbes*. July 20, 2017:1–6 Published online.
54. Lewis ZT, Totten SM, Smilowitz JT, et al. Maternal fucosyltransferase 2 status affects the gut bifidobacterial communities of breastfed infants. *Microbiome*. 2015;3.
55. Simpson MR, Avershina E, Storrø O, Johnsen R, Rudi K, Øien T. Breastfeeding-associated microbiota in human milk following supplementation with *Lactobacillus rhamnosus* GG, *Lactobacillus acidophilus* La-5, and *Bifidobacterium animalis* ssp. *lactis* Bb-12. *J Dairy Sci*. 2018;101(2):889–899.
56. Jiménez E, Fernández L, Maldonado A, et al. Oral Administration of *Lactobacillus* Strains Isolated from Breast Milk as an Alternative for the Treatment of Infectious Mastitis during Lactation. *Appl Environ Microbiol*. 2008;74(15):4650–4655.
57. Mastromarino P, Capobianco D, Miccheli A, et al. Administration of a multistrain probiotic product (VSL#3) to women in the perinatal period differentially affects breast milk beneficial microbiota in relation to mode of delivery. *Pharmacol Res*. 2015;95-96:63–70.
58. Griz EC, Bhandari V. The Human Neonatal Gut Microbiome: A Brief Review. *Front Pediatr*. 2015;3.
59. Khodayar-Pardo P, Mira-Pascual L, Collado MC, Martínez-Costa C. Impact of lactation stage, gestational age and mode of delivery on breast milk microbiota. *J Perinatol*. 2014;34(8):599–605.
60. Urbaniak C, Angelini M, Gloor GB, Reid G. Human milk microbiota profiles in relation to birthing method, gestation and infant gender. *Microbiome*. 2016;4:1.
61. Biagi E, Aceti A, Quercia S, et al. Microbial Community Dynamics in Mother’s Milk and Infant’s Mouth and Gut in Moderately Preterm Infants. *Front Microbiol*. 2018;9.
62. Tušar T, Žerdoner K, Bogovic Matijašič B, et al. Cultivable Bacteria from Milk from Slovenian Breastfeeding Mothers. *Food Technol Biotechnol*. 2014;52(2):242–247.
63. Treven P, Mahnič A, Rupnik M, et al. Evaluation of Human Milk Microbiota by 16S rRNA Gene Next-Generation Sequencing (NGS) and Cultivation/MALDI-TOF Mass Spectrometry Identification. *Front Microbiol*. 2019;10.
64. Moossavi S, Sepehri S, Robertson B, et al. Composition and Variation of the Human Milk Microbiota Are Influenced by Maternal and Early-Life Factors. *Cell Host Microbe*. 2019;25(2):324–335: e4.
65. Järvinen KM. Variations in Human Milk Composition: Impact on Immune Development and Allergic Disease Susceptibility. *Breastfeed Med*. 2018;13(S1):S–11.
66. Taghizadeh M, Mirlohi M, Poursina F, et al. The influence of impact delivery mode, lactation time, infant gender, maternal age and rural or urban life on total number of *Lactobacillus* in breast milk Isfahan - Iran. *Adv Biomed Res*. 2015;4:141.
67. Meehan CL, Lackey KA, Hagen EH, et al. Social networks, cooperative breeding, and the human milk microbiome. *Am J Hum Biol*. 2018;30(4):e23131.
68. Jost T, Lacroix C, Braegger C, Chassard C. Assessment of bacterial diversity in breast milk using culture-dependent and culture-independent approaches. *Br J Nutr*. 2013;110(7):1253–1262.
69. Lackey KA, Williams JE, Meehan CL, et al. What’s Normal? Microbiomes in Human Milk and Infant Feces Are Related to Each Other but Vary Geographically: The INSPIRE Study. *Front Nutr*. 2019;6.