5',5'-Phosphodiesters and esterase labile triesters of 2'-C-methylribonucleosides

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Dedicated to Prof Ferenc Fülöp on the occasion of his 60th birthday

Abstract

Bis(2'-C-methyladenosin-5'-yl) (11),bis(2´-C-methylguanosin-5´-yl) (13),bis(2'-*C*methyluridin-5'-yl) (15) and 2'-C-methylguanosin-5'-yl 2'-C-methyluridin-5'-yl (16) phosphodiesters have been prepared as pro-drug candidates for the respective 2'-Cmethylribonucleoside 5'-monophosphates, expectedly exhibiting antiviral activity against Hepatitis C virus. Additionally, the bis(2'-C-methyladenosine) diester has been converted to 3acetyloxymethoxy-2,2-bis(ethoxycarbonyl)propyl (19) or pivaloyloxymethyl (20) triester. The underlying idea is that the 5′,5′-phosphodiester is first released by intracellular carboxyesterases and subsequently cleaved to nucleoside and nucleoside 5'-monophosphate phosphodiesterases.

Keywords: Pro-drug, antiviral, nucleotide, phosphoester, synthesis

Introduction

Hepatitis C virus (HCV) infections constitute a worldwide health threat with an estimated 170 million infected individuals. ^{1,2} Besides interferon based therapies, ³ inhibitors of the viral NS3-4A serine protease and the NS5B RNA polymerase have emerged as potential drug candidates. ^{4,5} Among the RNA polymerase inhibitors, 2′-C-methylribonucleosides have received particular attention. 2′-C-methyl-3′-O-(L-valinyl)cytidine (Valopicitabine)^{6,7} and 4′-azido-3′,5′-di-O-(2-methylpropanoyl)cytidine showed antiviral activity in phase II and phase I clinal trials, respectively, and 2′-C-methyladenosine and 2′-C-methylguanosine inhibited HCV RNA replication *in vitro*. ⁹ To terminate the synthesis of viral RNA, antiviral nucleoside analogues need to be converted to their 5′-triphosphates. Since conversion to the monophosphate often is

the rate-limiting step, low antiviral activity may result from lack of this transformation by nucleoside kinases, and biodegradably protected nucleotides might, hence, be more potent drugs. For example, 2´-C-methylguanosine has been reported to suffer from inefficient uptake and phosphorylation. We now report on synthesis of novel type of *pro-2´-C*-methylribonucleotides. Two 2´-C-methylribonucleosides are linked to each other via a 5´,5´-phosphodiester bond that additionally bears an esterase labile protecting group, either a pivaloyloxymethyl or 3-acetyloxymethoxy-2,2-bis(ethoxycarbonyl)propyl group. The underlying idea is that intracellular carboxyesterases first releases the 5´,5´-phosphodiester, which is then cleaved by phosphodiesterases to nucleoside and nucleoside 5´-monophosphate.

Results and Discussion

Conversion of 2'-C-methyladenosine, 2'-C-methylguanosine and 2'-C-methyluridine to a protected form allowing their 5'-phosphitylation and, hence, the assembly of their 5',5'-phosphodiesters is outlined in Schemes 1-3, respectively. Accordingly, the 5'-OH of 2'-O-methyladenosine was first protected with a *tert*-butyldimethylsilyl group to give 1 and the 6-amino function then with a 4-methoxytrityl group (2; Scheme 1). Finally, the 2'- and 3'-OH groups were esterified with levulinic anhydride, prepared from levulinic acid *in situ* (3), and the 5'-silyl protection was removed (4). These two reactions deserve special attention.

Scheme 1. Reagents and conditions: (i) TBDMSCl, Py; (ii) MMTrCl, Py, Δ ; (iii) 1. Lev₂O, Py, 1,4-dioxane, 1 day 2. Lev₂O, DCM, 4 days; (iv) TBAF, THF, AcOH.

The levulinoylation of the tertiary 2´-OH turned out to be exceptionally difficult. Excess of levulinic anhydride (4 equiv.) and a 4 days reaction time had to be used. Fluoride ion promoted desilylation had, in turn, to be carried out in the presence of acetic acid to prevent removal of the levulinoyl groups. A closely related strategy was used for the protection of 2´-C-methylguanosine: the 5´-OH was converted to a *tert*-butyldimethylsilyl ether and the 2-amino function was alkylated with 4-methoxytrityl chloride (5; Scheme 2). The 3´-OH was protected either as a levulinic (6a) or benzoic acid (6b) ester, while the 2´-OH was left unprotected. 2´-C-Methyluridine, in turn, was protected by 4-methoxytritylation of the 5´-OH (7), benzolylation of the 2´- and 3´-OH (8) and removal of the trityl protection (9).

Scheme 2. Reagents and conditions: (i) 1. TBDMSCl, Py, 2. MMTrCl, Py; (ii) Lev₂O, DCM to obtain **6a**, BzCl, Py to obtain **6b**; (iii) TBAF, THF, AcOH.

Scheme 3. Reagents and conditions: (i) MMTrCl, Py; (ii) BzCl, Py; (iii) 80% AcOH.

Conversion of appropriately protected 2'-C-methylnucleosides (**4**, **6a**, **6b**, **9**) to dinucleoside-5',5'-monophosphates is depicted in Schemes 4-7. Phosphitylation of 2',3'-di-O-levulinoyl- N^6 -(4-methoxytrityl)-2'-C-methyladenosine (**4**) with half an equiv. of 1,1-dichloro-N,N-diisopropylphosphinamine, followed by tetrazole promoted displacement of the diisopropylamino ligand with a water molecule and oxidation with iodine under aqueous conditions gave the protected dimer (**10**; Scheme 4). The desired fully deprotected dimer (**11**) was obtained by removal of the levulinoyl groups with hydrazinium acetate in MeOH and the 4-methoxytrityl group with aqueous acetic acid. 3'-O-Levulinoyl- N^2 -(4-methoxytrityl)-2'-C-methylguanosine (**6a**) was, in turn, phosphitylated with 1 equiv. of 1-chloro-1-(2-cyanoethoxy)-N,N-diisopropylphosphinamine. Another equivalent of **6a** was coupled by tetrazole activation

and the phosphite triester was oxidized to phosphate ester (12; Scheme 5). The levulinoyl and 2-cyanoethyl groups were removed with methanolic ammonia and the 4-methoxytrityl with aqueous acetic acid (13). Finally, the ammonium counter ion was exchanged to sodium ion with the aid of a strong cation exchange resin.

Scheme 4. Reagents and conditions: (i) iPr₂NPCl₂, Et₃N, DCM; (ii) H₂O, TetH, MeCN; (iii) I₂, 2,6-lutidine, H₂O, THF; (iv) H₂NNH₃OAc, MeOH; (v) 80% AcOH

Scheme 5. Reagents and conditions: (i) 1-chloro-1-(2-cyanoethoxy)-*N*,*N*-diisopropylphosphinamine, Et₃N, DCM; (ii) **6a**, TetH, MeCN; (iii) I₂, 2,6-lutidine, H₂O, THF; (iv) NH₃, MeOH; (v) 80% AcOH; (vi) Dowex 50WX8 Na⁺ form, H₂O.

The same method was applied to preparation of bis(2′-*C*-methyluridin-5′-yl) phosphate (**15**; Scheme 6) from 2′,3′-di-*O*-benzoyl-2′-*C*-methyluridine (**9**) and preparation of 2′-*C*-methyluridin-5′-yl phosphate (**16**; Scheme 7) from **9** and 3′-*O*-

benzoyl- N^2 -(4-methoxytrityl)-2´-C-methylguanosine (**6b**). The low yields of compounds **11**, **12** and **13** resulted from difficulties in chromatographic purification. The migration of the derivatives of 2´-C-methylguanosine, in particular, was accompanied by marked broadening of the bands

Scheme 6. Reagents and conditions: (i) 1-chloro-1-(2-cyanoethoxy)-*N*,*N*-diisopropylphosphinamine, Et₃N, DCM; (ii) **9**, TetH, MeCN; (iii) I₂, 2,6-lutidine, H₂O, THF; (iv) NH₃, MeOH; (v) Dowex 50WX8 Na⁺ form, H₂O

Scheme 7. Reagents and **c**onditions: (i) 1-chloro-1-(2-cyanoethoxy)-*N*,*N*-diisopropylphosphinamine, Et₃N, DCM; (ii) **9**, TetH, MeCN; (iii) I₂, 2,6-lutidine, H₂O, THF; (iv) NH₃, MeOH; (v) 80% AcOH

Two different esterase labile groups were used to protect the internucleosidic 5',5'-phosphodiester linkage. 3-Acetyloxymethoxy-2,2-bis(ethoxycarbonyl)propyl group was introduced by phosphitylating the corresponding alcohol (17)¹² with bis[2',3'-di-O-levulinoyl- N^6 -(4-methoxytrityl)-2'-C-methyladenosin-5'-yl-N,N-diisopropylphosphoramidite and oxidizing the phosphite triester obtained to a phosphate triester (18; Scheme 8). The levulinoyl protections were removed by hydrazinium acetate in methanol. The rather labile acetyloxymethoxy function

of the phosphate protection group withstood this treatment. The deprotected triester (19) was obtained by removal of the 4-methoxytrityl group with 80% aq acetic acid.

Scheme 8. Reagents and conditions: (i) iPr₂NPCl₂, Et₃N, DCM; (ii) **17**, TetH, MeCN; (iii) I₂, 2,6-lutidine, H₂O, THF; (iv) H₂NNH₃OAc, MeOH; (v) 80% AcOH.

Introduction of the commonly used pivaloyloxymethyl group turned out to be difficult. Reaction of $bis[2',3'-di-O-levulinoyl-N^6-(4-methoxytrityl)-2'-C-methyladenosin-5'-yl]$ phosphate with chloromethyl pivalate was attempted in many solvents. The best result was obtained in N-methylpyrrolidone. The overall yield after removal of the levulinoyl and 4-methoxytrityl protections was, however, only 15% (20; Scheme 9).

Scheme 9. Reagents and conditions: (i) PivOCH₂Cl, NMP, Et₃N, 60 °C, 4 days ; (ii) H₂NNH₃OAc, DCM, MeOH; (iii) 80% AcOH.

The antiviral activity of the compounds will be tested by AliosBiopharma.

Experimental Section

General. Chemicals were purchased from Sigma-Aldrich, Fluka, Merck and Ramidus. MeCN, DCM, THF and pyridine were dried over 4Å molecular sieves and 1,4-dioxane over 3Å molecular sieves. Et₃N was dried by refluxing over CaH₂, distilled before use and stored over CaH₂ lumps. Reactions were monitored with TLC (*Merck* silica gel 60 F₂₅₄ aluminium sheets) or with ³¹P-NMR. Column chromatography was performed on silica gel (Fluka silica gel 60 230-400 mesh). ¹H-, ¹³C-, ³¹P- and 2D-NMR spectra were recorded on Bruker Avance 400 or 500 spectrometer at 25°C. The chemical shifts are given in ppm with reference to internal TMS, the coupling constants *J* are given in Hz. High resolution mass spectra were recorded on Bruker Daltonics microTOF-Q using electrospray ionization.

5′-O-(tert-Butyldimethylsilyl)-2′-C-methyladenosine (1). 2′-C-Methyladenosine 1 (5.26 mmol; 1.48 g) was coevaporated twice from dry pyridine and dissolved in the same solvent (10 mL). *tert-*Butyldimethylsilyl chloride (5.21 mmol; 0.81 g) was added and the reaction mixture was stirred for 18 h at r.t. Volatiles were removed and the residue was dissolved in DCM, washed with saturated aqueous NaHCO₃ and brine, dried over Na₂SO₄, and evaporated to dryness. The crude product was purified on a silica gel column eluting with 5% MeOH in DCM. Compound **1** was obtained as white solid in 67% yield (1.40 g). 1 H-NMR (500 MHz, CDCl₃) δ 8.41 (s, 1H, H2), 8.35 (s, 1H, H8), 6.83 (br. s, 2H, NH₂), 6.29 (s, 1H, H1′), 6.22 (br. s, 1H, OH), 4.62 (br. s, 1H, OH), 4.12-4.20 (m, 3H, H3′, H4′, H5′), 3.97 (d, J = 10.2 Hz, 1H, H5′′), 1.05 (s, 3H, 2′-Me), 0.94 (s, 9H, Me₃CSi), 0.15 (s, 3H, MeSi), 0.14 (s, 3H, MeSi). 13 C-NMR (126 MHz, CDCl₃) δ 155.82 (C5), 152.66 (C8), 148.98 (C4), 138.80 (C2), 119.50 (C6), 91.38 (C1′), 83.67 (C4′), 79.21 (C2′), 72.98 (C3′), 61.57 (C5′), 26.06 (Me_3 CSi), 20.25 (Me₃CSi), 18.63 (2′-Me), 5.24 (MeSi), -5.39 (MeSi). HR-ESI-MS:). HR-ESI-MS: [M+H]⁺ obsd. 396.2041, calcd. 396.2062.

5'-O-(tert-Butyldimethylsilyl)- N^6 -(4-methoxytrityl)- 2'-C-methyladenosine (2). Compound 1 (3.54 mmol; 1.40 g) was dried over P_2O_5 for one day and dissolved in dry pyridine (10 mL). 4-Methoxytrityl chloride (3.92 mmol; 1.21 g) was added and the reaction mixture was stirred on an oil bath at 51 °C for 17 hours. The reaction was quenched with MeOH and the mixture was evaporated to dryness. The residue was purified on a silica gel column eluting with 50% EtOAc in petroleum ether. Compound 2 was obtained as white solid in 89% yield (2.11 g). 1 H-NMR (500 MHz, CDCl₃) δ 8.22 (s, 1H, H2), 7.98 (s, 1H, H8), 7.22-7.34 (m, 12H, MMTr), 7.05 (s, 1H, NH), 6.79 (d, J = 9.0 Hz, 2H, MMTr), 6.10 (s, 1H, H1'), 4.05-4.11 (m, 3H, H3', H4', H5'), 3.93 (dd, J = 8.7 and J = 3.6 Hz, 1H, H5''), 3.77 (s, 3H, OMe), 1.01 (s, 3H, Me2'), 0.95 (s, 9H, Me₃CSi), 0.14 (s, 3H, MeSi), 0.14 (s, 3H, MeSi). 13 C-NMR (126 MHz, CDCl₃) δ 158.34 (MMTr), 154.29 (C5), 151.75 (C8), 147.83 (C4), 145.04 (MMTr), 138.11 (C2), 137.07 (MMTr),

130.25, 128.92, 127.89, 126.91 (MMTr), 120.98 (C6), 113.15 (MMTr), 91.42 (C1′), 84.32 (C4′), 78.75 (C2′), 73.64 (C3′), 71.18 (MMTr), 61.90 (C5′), 55.23 (MeO of MMTr), 26.07, 26.01, 25.94 (Me_3 CSi), 20.33 (Me2′), 18.61 (Me_3 CSi), -5.26, -5.36 (MeSi). HR-ESI-MS: [M+H]⁺ obsd. 668.3268, calcd. 668.3263, [M+Na]⁺ obsd. 690.3084, calcd. 690.3082.

5'-O-(tert-Butyldimethylsilyl)-2',3'-di-O-levulinoyl-N⁶-(4-methoxytrityl)-2'-C-methyl-

adenosine (3). Levulinic acid (16.36 mmol; 1.90 g) was dissolved in dry 1,4-dioxane (20 mL) and cooled on an ice bath. Dicyclohexylcarbodiimide (DCC; 8.58 mmol; 1.77 g) was added portion wise within an hour. The reaction mixture was filtered and the precipitate was washed with dry 1,4-dioxane (10 mL). Compound 2 (3.16 mmol; 2.11 g) was coevaporated with dry pyridine and dissolved in the same solvent (10 mL) and the levulinic anhydride in 1,4-dioxane was added together with a catalytic amount of 4-dimethylaminopyridine (DMAP). The reaction mixture was stirred at r.t. overnight, heated on an oil bath at 54 °C for 5 hours and stirred again over night at r.t. The reaction mixture was evaporated to dryness, the residue was dissolved in DCM and washed with H₂O, saturated aqueous NaHCO₃ and brine. The crude product, dried over Na₂SO₄, was purified twice by silica gel chromatography, first eluting with 2% MeOH in DCM and then with 1% MeOH in DCM. According to the NMR spectrum, a mixture of products containing one or two levulinoyl groups was obtained, the singly acylated product predominating. The mixture of products was subjected to another levulinovlation. Levulinic acid (11.19 mmol; 1.30 g) was dissolved in dry DCM (20 mL) and DCC (5.48 mmol; 1.13 g) was added in one portion. The reaction mixture was stirred at r.t. overnight and filtered. The precipitate was washed with dry DCM (5 mL) and the DCM phases were combined. The filtrate and a catalytic amount of DMAP were added to the mixture (2.11 g) in dry DCM (5 mL). Owing to sluggish progress of the reaction, dry Et₃N (0.50 mL) and some DMAP were after one day added to the reaction mixture. The reaction was completed in 4 days. The reaction mixture was washed with H₂O, saturated aqueous NaHCO₃ and brine, dried over Na₂SO₄ and evaporated to dryness. The crude product was purified on a silica gel column eluting with 1% MeOH in DCM. Compound 3 was obtained as yellowish solid in 72% yield (1.98 g). ¹H-NMR (400 MHz, CDCl₃) δ 8.09 (s, 1H, H2), 8.07 (s, 1H, H8), 7.22-7.36 (m, 12H, MMTr), 6.93 (s, 1H, NH), 6.80 (d, J =11.2 Hz, 2H, MMTr), 6.38 (s, 1H, H1 $^{\circ}$), 5.59 (d, J = 6.8 Hz, 1H, H3 $^{\circ}$), 4.14 (m, 1H, H4 $^{\circ}$), 4.01 (dd, J = 11.7 and 2.4 Hz, 1H, H5'), 3.96 (dd, J = 11.6 and 3.2 Hz, 1H, H5''), 3.78 (s, 3H, Me of the state of the sMMTr), 2.59-2.79 (m, 8H, CH₂ of Lev), 2.19 (s, 3H, Me of Lev), 2.08 (s, 3H, Me of Lev), 1.41 (s, 3H, Me2'), 0.93 (s, 9H, Me₃CSi), 0.12 (s, 6H, MeSi). ¹³C-NMR (101 MHz, CDCl₃) δ 206.34, 206.23 (CH₂COCH3 Lev), 171.73, 171.10 (OCOCH₂ Lev), 158.32 (MMTr), 154.14 (C6), 152.52 (C2), 148.68 (C4), 145.20 (MMTr), 138.54 (C8), 137.23 (MMTr), 130.23, 128.91, 127.87, 126.85 (MMTr), 120.85 (C5), 113.15 (MMTr), 87.87 (C1'), 84.69 (C2'), 82.89 (C4'), 74.26 (C3´), 71.02 (MMTr), 62.16 (C5´), 55.22 (MeO of MMTr), 37.86, 37.74 (MeCOCH₂ Lev), 29.76, 29.65 (Me of Lev), 28.54, 27.72 (CH₂COO Lev), 25.99 (Me₃CSi), 18.52 (Me₃CSi), 17.04 (Me2⁻), -5.43, -5.45 (MeSi). HR-ESI-MS: [M+H]⁺ obsd. 864.4001, calcd. 864,3998.

2',3'-Di-O-levulinoyl- N^6 -(4-methoxytrityl)-2'-C-methyladenosine (4). Compound 3 (2.29 mmol; 1.98 g) was dissolved in a mixture of THF (32 mL) and AcOH (6 mL) and

tetrabutylammonium fluoride hydrate (4.63 mmol; 1.21 g) was added. The reaction mixture was stirred at r.t for 42 hours and evaporated to dryness. The residue was dissolved in EtOAc, washed with $\rm H_2O$, saturated aqueous NaHCO₃ and brine. The crude product dried over Na₂SO₄ was purified on a silica gel column eluting with 3% MeOH in DCM. Compound 4 was obtained as white solid in 80% yield (1.37 g). 1 H-NMR (500 MHz, CDCl₃) δ 8.00 (s, 1H, H2), 7.90 (s, 1H, H8), 7.21-7.33 (m, 12H, MMTr), 7.01 (s, 1H, NH), 6.80 (d, J = 10.1 Hz, 2H, MMTr), 6.44 (s, 1H, H1′), 5.83 (d, J = 6.8 Hz, 1H, H3′), 5.62 (m, 1H, OH5′), 4.30 (m, 1H, H4′), 4.09 (dd, J = 12.7 and 1.3 Hz, 1H, H5′), 3.78 (s, 3H, Me of MMTr), 3.76 (m, 1H, H5′′), 2.62-2.84 (m, 8H, CH₂ of Lev), 2.21 (s, 3H, Me of Lev), 2.19 (s, 3H, Me of Lev), 1.31 (s, 3H, Me2′). 13 C-NMR (126 MHz, CDCl₃) δ 206.49, 206.28 (CH₂COMe Lev), 172.21, 171.78 (OCOCH₂ Lev), 158.37 (MMTr), 154.51 (C6), 152.18 (C2), 147.69 (C4), 144.99, 144.97 (MMTr), 139.66 (C8), 136.95 (MMTr), 130.20, 128.86, 127.92, 126.94 (MMTr), 121695 (C5), 113.18 (MMTr), 91.50 (C1′), 86.78 (C2′), 81.36 (C4′), 73.69 (C3′), 71.16 (MMTr), 60.28 (C5′), 55.22 (MeO of MMTr), 37.89, 37.85 (MeCOCH₂ Lev), 29.81, 29.78 (Me of Lev), 28.71, 27.67 (CH₂COO Lev), 18.47 (Me2′). HR-ESI-MS: [M+H] $^+$ obsd. 750.3109, calcd. 750.3134.

 $5'-O-(tert-Butvldimethylsilvl)-N^2-(4-methoxytrityl)-2'-C-methylguanosine (5). 2'-C-Methyl$ guanosine (12.28 mmol; 3.65 g) was dried over P₂O₅ for two days, dissolved in dry pyridine (30 mL) and tert-butyldimethylsilyl chloride (12.27 mmol; 1.85 g) was added. The reaction mixture was stirred at r.t. for 24 hours and evaporated to dryness. The residue was coevaporated three times with dry pyridine and dissolved in the same solvent (15 mL). 4-Methoxytrityl chloride (12.27 mmol; 3.79 g) was added and the reaction mixture was heated on an oil bath at 57 °C for 20 hours and evaporated to dryness. The residue was dissolved in DCM, washed with saturated aqueous NaHCO₃ and brine, dried over Na₂SO₄, and evaporated to dryness. The crude product was purified three times on a silica gel column: first with 8% MeOH in DCM, then with 4% MeOH in DCM and finally by using gradient elution from 1% to 4% MeOH in DCM. Compound 5 was obtained as white solid in 82% yield (6.88 g). ¹H-NMR (500 MHz, CDCl₃) δ 7.74 (s, 1H, H8), 7.11-7.36 (m, 13H, MMTr, NH), 6.79 (d, J = 8.7 Hz, 2H, MMTr), 5.37 (s, 1H, H1'), 3.85-4.02 (m, 5H, H3', H4', H5', H5'', OH), 3.70 (s, 3H, OMe), 2.89 (br. s, 1H, OH), 0.92 (s, 9H, Me₃CSi), 0.54 (s, 3H, Me²), 0.11 (s, 6H, MeSi). ¹³C-NMR (126 MHz, CDCl₃) δ 158.40 (MMTr), 158.26 (C6), 151.38 (C4), 149.89 (C2), 144.73, 144.61 (MMTr), 136.33 (MMTr), 135.44 (C8), 130.29, 128.82, 128.71, 127.87, 126.83 (MMTr), 117.25 (C5), 113.17 (MMTr), 90.64 (C1'), 83.36 (C4'), 78.69 (C2'), 72.86 (C3'), 70.52 (MMTr), 62.15 (C5'), 55.15 (MeO of MMTr), 26.07 (*Me*₃CSi), 19.77 (Me₂'), 18.55 (Me₃CSi), -5.30, -5.38 (MeSi). HR-ESI-MS: [M+H]⁺ obsd. 684.3213, calcd. 684.3212, [M+Na]⁺ obsd. 706.3054, calcd. 706.3031.

 N^2 -(4-Methoxytrityl)-3'-O-levulinoyl-2'-C-methylguanosine (6a). Levulinic acid (4.82 mmol; 0.56 g) was dissolved in dry DCM (20 mL), DCC (2.90 mmol; 0.60 g) was added and the reaction mixture was stirred at r.t. for 4 hours. Precipitated dicyclohexylurea was removed by filtration and compound 5 (1.94 mmol; 1.33 g) in DCM (2 mL) and a catalytic amount of DMAP and Et₃N (0.80 ml) were added. The reaction mixture was stirred at r.t. over night. The mixture was washed with H₂O and brine. The organic layer was dried over Na₂SO₄ and evaporated to

dryness. The crude product was purified on a silica gel column eluting with 1 to 3% MeOH in DCM. According to NMR spectroscopy, the main product bore only one levulinoyl group. The product was used as such for the next step. Accordingly, the levulinoylated nucleoside was dissolved in a mixture of dry THF (20 mL) and acetic acid (4 mL). Tetrabutylammonium fluoride hydrate (2.95 mmol, 0.77 g) was coevaporated from dry THF and added to the reaction mixture in the same solvent (1 mL). The mixture was evaporated to dryness after 21 hours stirring at r.t. The residue was equilibrated between EtOAc and saturated aqueous NaHCO3. The organic layer was washed with brine, dried over Na₂SO₄ and evaporated to dryness. The crude product was purified on a silica gel column eluting with 5% MeOH in DCM. The product was obtained as white solid in 50% yield (0.64 g). ¹H-NMR (500 MHz, CDCl₃) δ 7.76 (s, 1H, H8), 7.17-7.34 (m, 13H, MMTr, NH), 6.78 (d, J = 8.8 Hz, 2H, MMTr), 5.52 (s, 1H, H1'), 5.22 (m, 1H, H3'), 4.16 (m, 1H, H4'), 4.02 (m, 1H, H5'), 3.74 (s, 3H, OMe), 3.70 (m, 1H, H5''), 2.80-2.83 (m, 2H, MeCOCH₂CH₂ Lev), 2.51-2.65 (m, 2H, CH₂CH₂COO Lev), 2.20 (s, 3H, Me of Lev), 0.73 (s, 3H, Me2'). ¹³C-NMR (126 MHz, CDCl₃) δ 207.16 (CH₂COCH3 Lev), 172.35 (OCOCH₂ Lev), 158.15 (MMTr), 151.56 (C6), 149.79 (C4), 143.38 (MMTr), 135.86 (MMTr), 135.71 (C8), 130.13, 128.65, 128.22, 127.11 (MMTr), 117.67 (C5), 113.47 (MMTr), 80.60 (C4´), 80.13 (C2´), 73.62 (C3´), 70.54 (MMTr), 60.44 (C5´), 55.23 (MeO of MMTr), 38.23 (MeCOCH₂ Lev), 29.77 (Me of Lev), 27.97 (CH₂COO Lev), 20.80 (Me2'). HR-ESI-MS: [M+H]⁺ obsd. 668.2713, calcd. 668.2715, [M+Na]⁺ obsd. 690.2553, calcd. 690.2534.

 N^2 -(4-Methoxytrityl)-3'-O-benzoyl-2'-C-methylguanosine (6b). Compound 5 (1.74 mmol; 1.19 g) was coevaporated three times with dry pyridine and the residue was dissolved in the same solvent (5.0 mL). The solution was cooled on an ice bath and benzoyl chloride (4.39 mmol; 0.22 mL) was added. The solution was stirred at r.t. overnight and evaporated to dryness. The residue was dissolved in DCM (5.0 mL) and washed with brine. The organic phase was dried over Na₂SO₄ and evaporated to dryness. The crude product was isolated by Silica gel chromatography eluting with DCM containing MeOH from 5% to 8% MeOH. HR-ESI-MS: [M+H]⁺ obsd. 788.3393, calcd. 788.3474. According to ¹H- and ¹³C-NMR spectroscopic characterization, the desired product, 5'-O-(tert-butyldimethylsilyl)-N²-(4-methoxytrityl)-3'-Obenzoyl-2'-C-methylguanosine, was contaminated with some impurities. In spite of this, the crude product was subjected as such to desilylation that was carried out as described above for compound 6a. The crude product was purified by silica gel chromatography eluting with DCM that contained 5% MeOH. Compound **6b** was obtained as white solid in 53% yield (0.52 g). ¹H-NMR (500 MHz, CDCl₃) δ : 8.05 (dd, 2H, J = 8.45 and 1.35 Hz, Bz), 7.84 (br. s, 1H, H8), 7.60 (m, 1H, Bz), 7.48 (t, 2H, J = 7.90 Hz, Bz), 7.36-7.11 (12H, MMTr), 6.72 (d, J = 8.75 Hz, 2H, MMTr), 5.44 (s, 1H, H1'), 5.28 (m, 1H, H3'), 4.24 (m, 1H, H4'), 4.05 (m, 1H, H5''), 3.76-3.72 (m, 2H, H5', OH), 3.68 (s, 3H, OMe), 2.25 (br s, 1H, N²H), 0.73 (s, 3H, 2'-Me). ¹³C-NMR (126 MHz, CDCl₃) δ: 165.92 (C=O Bz), 158.21 (C6), 151.45 (C4), 149.89 (C2), 144.82 (C8), 158.1, 144.9, 133.74, 130.28, 129.89, 129.00, 128.87 128.77, 128.60, 127.86, 127.81 and 126.75 (MMTr and Bz), 117.26 (C5), 113.17 (MMTr), 90.81 (C1'), 80.98 (C4'), 79.29 (MMTr), 74.00

(C3'), 70.76 (C2'), 60.59 (C5'), 55.19 (MeO of MMTr), 20.55 (2'-CH₃). HR-ESI-MS: [M+H]⁺ obsd. 674.2603, calcd. 674.2609.

5′-O-(4-Methoxytrityl)-2′-C-methyluridine (7). 2′-C-Methyluridine (0.79 g; 3.05 mmol) was dissolved in dry pyridine (20 mL) under argon. 4-Methoxytrityl chloride (1.03 g; 3.36 mmol) was added and the mixture was stirred at 50 °C for 20 hours. The reaction was quenched with saturated aq NaHCO₃ (50 mL) and subjected to DCM (3 × 100 mL) workup. The organic phase was dried over Na₂SO₄ and evaporated to dryness. The residue was purified by Silica gel chromatography using DCM containing 5% MeOH as eluent. Compound **7** was obtained as solid in 92% yield (1.50 g). 1 H-NMR (500 MHz, CDCl₃) δ : 8.16 (d, 1H, J = 8.0 Hz, H6), 7.41 (m, 4H, MMTr), 7.26 (m, 6H, MMTr), 7.19 (m, 2H, MMTr), 6.86 (d, 2H, J = 9.0 Hz, MMTr), 6.09 (s, 1H, H1′), 5.23 (d, 1H, J = 8.0 Hz, H5), 4.05-4.17 (m, 2H, H3′&H4′), 3.81-3.84 (m, 4H, MeO-MMTr & H5′), 3.61 (br s, 2H, 2′-&3′-OH); 3.23 (d, 1H, J = 12.5 Hz, H5′′), 1.36 (s, 3H, CH₃). 13 C-NMR (100 MHz, CD₃OD) δ : 164.64 (C4), 158.76 (MMTr), 151.02 (C2), 144.52 (MMTr), 141.12 (C6), 135.35 (MMTr), 130.18, 128.10, 127.34, 126.46 and 112.64 (MMTr), 102.94 (C5), 91.67 (C1′), 87.66 (MMTr), 82.44 (C2′), 78.66 (C4′), 71.94 (C3′), 59.05 (C5′), 54.32 (OMe), 18.80 (Me). HR-ESI-MS: [M+Na]⁺ obsd. 553.1921, calcd. 553.1945.

2',3'-di-O-Benzoyl-5'-O-(4-methoxytrityl)-2'-C-methyluridine (8). 5'-O-(4-Methoxytrityl)-2'-C-methyluridine (1.50 g; 2.87 mmol) (7) was dissolved in dry pyridine (60 mL). Benzoyl chloride (0.83 mL, 7.17 mmol) was added and the mixture was stirred overnight at room temperature. The reaction was quenched with cold water and the mixture was evaporated to dryness. The residue was dissolved in DCM and washed with water (3 \times 100 mL). The organic phase was dried over Na₂SO₄ and evaporated to dryness. The residue was purified by Silica gel chromatography using DCM containing 10% EtOAc as eluent. Compound 8 was obtained as solid in 77% yield (1.63 g). ¹H-NMR (400 MHz, CDCl₃) δ : 8.29 (d, 1H, J = 8.4 Hz, H6), 8.07 (d, 2H, J = 7.6 Hz, Bz), 7.90 (d, 2H, J = 7.6 Hz, Bz), 7.64 (t, 2H, J = 7.2 Hz, Bz), 7.51-7.42 (m, 8H, Bz and MMTr), 7.33-7.17 (m, 8H, Bz and MMTr), 6.80 (d, 2H, J = 8.8 Hz, MMTr), 6.07 (s, 1H, H1 $^{\circ}$), 5.77 (d, 1H, J = 8.4 Hz, H5), 5.45 (d, 1H, J = 9.2 Hz, H3 $^{\circ}$), 4.45 (d, 1H, J = 8.8 Hz, H4 $^{\circ}$), 3.74 (s, 3H, OCH₃), 3.63 (d, 1H, J = 11.2 Hz, H5^{-/-}), 3.46 (d, 1H, J = 9.6 Hz, H5^{-/-}), 2.88 (br s, 1H, NH), 1.36 (s, 3H, CH₃). ¹³C-NMR (100 MHz, CDCl₃) δ: 168.30 (C=O Bz), 165.43 (C4), 161.78 (C=O Bz), 158.80 (MMTr), 149.75 (C2), 143.71, 143.49 (MMTr), 139.79 (C6), 135.15, 134.35, 134.17, 133.88, 131.39, 130.52, 130.39, 129.94, 129.20, 128.72, 128.69, 128.44, 128.35, 128.08, 127.88, 127. 30, 127.25, 127.20, 113.34, 113.22 (Bz and MMTr), 102.52 (C5), 91.66 (C1'), 87.55 (spiro C, MMTr), 79.64 (C2'), 79.41 (C4'), 73.39 (C3'), 60.45 (C5'), 55.20 (MeO of MMTr), 21.00 (Me). HR-ESI-MS: [M+Na]⁺ obsd. 761.2450, calcd. 761.2470.

2′,3′-Di-*O*-benzoyl-**2′-***C*-methyluridine (9). 2′,3′-Di-*O*-benzoyl-5′-*O*-(4-methoxytrityl)-2′-*C*-methyluridine (1.88g; 2.54 mmol) (8) was dissolved in 80% (v/v) aq AcOH (36 mL). After stirring overnight at room temperature, the mixture was evaporated to dryness. The residue was dissolved in dichloromethane (100 mL) and washed with water (2 × 100 mL). Organic phase was dried over Na₂SO₄ and evaporated to dryness. The residue was purified by Silica gel chromatography eluting with a 7:3 mixture of DCM and EtOAc. Compound **9** was obtained as

white foam in 78% yield (0.90 g). 1 H-NMR (400 MHz, CDCl₃) δ : 8.21 (d, 1H, J = 8.2 Hz, H6), 8.07 (dd, 2H, J = 8.0 and 1.2 Hz, Bz), 7.91 (dd, 2H, J = 8.0 and 1.3 Hz, Bz), 7.66 (m, 2H, Bz), 7.50 (t, 4H, J = 7.2 Hz, Bz), 6.09 (s, 1H, H1'), 5.91 (d, 1H, J = 8.2 Hz, H5), 5.22 (d, 1H, J = 8.7Hz, H3 $^{\circ}$), 4.32 (d, 1H, J = 8.5 Hz, H4 $^{\circ}$), 4.12 (dd, 1H, J = 13.2 Hz and 1.3 Hz, H5 $^{\circ}$), 3.85 (dd, 1H, J = 13.2 Hz and 2.2 Hz, H5²), 1.38 (s, 3H, CH₃). ¹³C-NMR (100 MHz, CDCl₃) δ : 168.29 (C=O Bz), 166.45 (C4), 161.76 (C=O Bz), 149.78 (C2), 139.49 (C6), 135.18, 134.24, 131.40, 130.40, 129.96, 129.21, 128.80, 128.34 (Bz), 102.66 (C5), 91.55 (C1'), 80.74 (C4'), 79.71 (C2'), 73.50 (C3⁻), 60.36 (C5⁻), 20.82 (CH₃). HR-ESI-MS: [M+Na]⁺ obsd. 489.1241, calcd. 489.1268. Bis[2',3'-di-O-levulinoyl- N^6 -(4-methoxytrityl)-2'-C-methyladenosin-5'-yl] phosphate (10). Compound 4 (1.82 mmol; 1.37 g) was dried on P₂O₅ over two nights and dissolved in dry DCM (6 mL) under nitrogen. Dry Et₃N (9.28 mmol; 1.29 mL) and 1,1-dichloro-N,Ndiisopropylphosphinamine (0.97 mmol; 180 µL) were added and the reaction mixture was stirred at r.t. for 1,5 hours. To complete the reaction, another portion of 1,1-dichloro-N,Ndiisopropylphosphinamine (0.27 mmol; 50 µL) was added and the mixture was stirred for 30 minutes. The reaction mixture was then passed through a short silica gel column eluting with EtOAc/hexane (8:2, v/v) containing 0.5% Et₃N. The solvents were removed under reduced pressure and the residue was co-evaporated two times from dry MeCN. ³¹P-NMR signal at 149.66 ppm verified the formation of the desired dinucleoside phosphoramidite. The product (0.44 mmol; 0.72 g) was dissolved in MeCN (1 mL), and 1-H-tetrazole (5.22 mmol; 11.60 mL of 0.45 mol L⁻¹ solution in MeCN) and H₂O (8.00 mL) were added. The progress of the reaction was followed by ³¹P-NMR spectroscopy. After 75 min, 7 mL of 0.15 mol L⁻¹ solution of I₂ (1.05 mmol) in a 4:2:1 mixture of THF, H₂O and 1,2-lutidine was added. The oxidation was completed in 2.5 hours. The product was isolated by conventional 5% aq NaHSO₃/DCM workup. The crude product was purified on a silica gel column with gradient elution from 5% to 10% MeOH in DCM. Compound 10 was obtained as yellowish solid in 70% yield (0.39 g). ¹H-NMR (500 MHz, $CDCl_3$) δ 8.32 (s, 2H, H8), 8.03 (s, 2H, H2), 7.19-7.35 (m, 24H, MMTr), 6.78 (d, J = 9.0 Hz, 4H, MMTr), 6.36 (s, 2H, H1'), 5.59 (d, J = 5.0 Hz, 2H, H3'), 4.15-4.23 (m, 6H, H4', H5' & H5''), 3.75 (s, 6H, OMe of MMTr), 2.51-2.79 (m, 16H, CH₂ of Lev), 2.17, 2.05 (2s, 12H, Me of Lev), 1.37 (s, 6H, Me2'). ¹³C-NMR (126 MHz, CDCl₃) δ 207.74, 207.05 (CH₂COCH3 Lev), 171.99, 171.08 (OCOCH₂ Lev), 158.11 (MMTr), 153.73 (C6), 152.23 (C2), 148.44 (C4), 144.79 (MMTr), 139.03 (C8), 136.86 (MMTr), 130.06, 128.70, 127.64, 126.70 (MMTr), 119.88 (C5), 112.90 (MMTr), 87.67 (C1´), 84.20 (C2´), 80.94 (C4´), 74.13 (C3´), 70.85 (MMTr), 63.97, 63.95 (C5'), 54.97 (MeO of MMTr), 37.62, 37.45 (MeCOCH₂ Lev), 29.34, 29.29 (Me of Lev), 28.19, 27.60 (CH₂COO Lev), 16.77 (Me2′). ³¹P-NMR (202 MHz, CDCl₃) δ 0.10. HR-ESI-MS: [M-H]⁻ obsd. 1559.5598, calcd. 1559.5606.

Bis(2'-C-methyladenosin-5'-yl) phosphate (11). Compound **10** (0.04 mmol; 0.06 g) was dissolved in DCM (0.50 mL) and hydrazinium acetate (0.32 mmol; 0.03 g) in MeOH (0.2 mL) was added. After 20 hours stirring at r.t., the reaction mixture was evaporated to dryness and the residue was dissolved in 80% aqueous AcOH (1 mL) and stirred at r.t. for 4 hours. The volatiles were removed under reduced pressure and the residue was equilibrated between H₂O and ethyl

acetate. The aqueous layer was separated and evaporated to dryness. The crude product was purified by HPLC on a Sun FireTM Prep C18 column (250×10 mm, 5 μ m, flow rate 3.0 mL min⁻¹), using isocratic elution with 9% buffer A in buffer B (Buffer A: 20 mmol L⁻¹ aq NH₄OAc in 50% MeCN, Buffer B: 20 mmol L⁻¹ aq NH₄OAc). Compound **11** was obtained as white solid in 27% yield (6 mg as an ammonium salt). ¹H-NMR (500 MHz, D₂O) δ 7.99 (s, 2H, H2), 7.96 (s, 2H, H8), 5.87 (s, 2H, H1′), 4.36-4.40 (m, 2H, H5′), 4.22-4.27 (m, 4H, H4′, H5′'), 4.01 (m, 2H, H3′), 0.71 (s, 6H, Me2′). ¹³C NMR (126 MHz, D₂O) δ 154.59 (C6), 152.13 (C2), 147.29 (C4), 139.27 (C8), 117.60 (C5), 90.76 (C1′), 80.62, 80.55 (C4′), 79.18 (C2′), 72.21 (C3′), 65.60, 65.56 (C5′), 18.30 (Me2′). ³¹P-NMR (202 MHz, CDCl₃) δ 0.28. HR-ESI-MS: [M-H]⁻ obsd. 623.1735, calcd. 623.1733.

2-Cyanoethyl bis[3'-O-levulinoyl- N^6 -(4-methoxytrityl)-2'-C-methylguanosin-5'-yl] phosphate (12). Compound 6a (0.48 mmol; 0.32 g) dried over P₂O₅ over night was dissolved in dry DCM (3 mL) under N₂. Dry Et₃N (2.41 mmol; 335 µL) and 1-chloro-1-(2-cyanoethoxy)-N,Ndiisopropylphosphinamine (0.53 mmol; 118 µL) were added. After 1 hour stirring at r.t., the mixture was passed through a short silica gel column eluting with EtOAc that contained 0.5% Et₃N. The solvents were removed under reduced pressure and the residue was co-evaporated from dry MeCN. The residue was dissolved in dry MeCN (2 mL) and compound 6 (0.48 mmol; 0.32 g) and 1-H-tetrazole (0.96 mmol; 2.13 mL of 0.45 mol L⁻¹ solution in MeCN) were added under N₂, and the mixture was stirred for 1.5 hours. The phosphite ester obtained was oxidized as described above for compound 10. The phosphate triester 12 was isolated by conventional 5% aq NaHSO₃/DCM workup. The crude product was purified on a silica gel column eluting with 5 to 10% MeOH in DCM. Compound 12 was obtained as white solid in 21% yield (0.15 g). ¹H-NMR (500 MHz, MeOD) δ 7.82 (s, 1H, H8), 7.81 (s, 1H, H8), 7.19-7.38 (m, 24H, MMTr), 6.86 (m, 4H, MMTr), 5.35 (s, 2H, H1'), 5.15 (m, 2H, H4'), 4.25-4.42 (m, 8H, H3', H5', H5'', CH₂), 3.77 (s, 6H, OMe), 2.86-2.89 (m, 2H, CH₂CN), 2.77-2.84 (m, 4H, MeCOCH₂CH₂ Lev), 2.61-2.64 (m, 4H, CH₂CH₂COO Lev), 2.15 (s, 3H, Me of Lev), 2.13 (s, 3H, Me of Lev), 0.63 (s, 3H, Me2'), 0.63 (s, 3H, Me2'). ¹³C-NMR (126 MHz, MeOD) δ 208.33 (CH₂COCH₃ Lev), 158.48 (MMTr), 157.00 (C6), 151.21 (C4), 149.25 (C2), 144.78, 144.80, 144.81, 144.83 (MMTr), 136.50 (MMTr), 135.25 (C8), 130.05, 128.64, 127.48, 126.50 (MMTr), 120.75 (CN), 112.76 (MMTr), 78.55 (C4´), 77.89 (C2´), 73.13 (C3´), 70.42, 70.45 (MMTr), 66.17 (C5´), 63.00 (OCH₂CH₂CN), 54.37 (MeO of MMTr), 37.30 (MeCOCH₂ Lev), 28.32 (Me of Lev), 27.36 $(CH_2COO Lev)$, 18.96 (Me2') 18.62, 18.57 (OCH_2CH_2CN) . ³¹P-NMR (202 MHz, MeOD) δ -2.24.

Bis(2'-C-methylguanosin-5'-yl) phosphate (13). Compound 12 (0.10 mmol; 0.15 g) was dissolved in 30% methanolic ammonia (25 mL) and the mixture was stirred at r.t. overnight, and evaporated to dryness under reduced pressure. The residue was dissolved in 80% aq AcOH (10 mL) and the mixture was stirred at r.t. for 3 days. The crude product obtained by evaporation to dryness was equilibrated between H₂O and hexane. The aqueous phase was evaporated to dryness and the product was purified by HPLC as described above for compound 11. The product was dissolved in water and passed through an ion-exchange column (Dowex Na⁺-form,

50Wx8, 100-200 mesh) and evaporated to dryness. Compound **13** was obtained as white solid in 18% yield (12 mg). 1 H-NMR (500 MHz, D₂O) δ 7.55 (s, 2H, H8), 5.51 (s, 2H, H1′), 4.18 (m, 2H, H5′), 4.02-4.11 (m, 4H, H4′, H5′′), 3.88 (m, 2H, H5′), 0.65 (s, 6H, 2′-Me). 13 C-NMR (126 MHz, MeOD) δ 163.26 (C6), 90.72, 90.70 (C1′), 80.60, 80.59, 80.52, 80.50 (C4′), 79.25 (C2′), 72.55, 72.53 (C3′), 65.70, 65.66 (C5′), 18.52, 18.50 (2′-Me). 31 P-NMR (202 MHz, D₂O) δ 0.20. HR-ESI-MS: [M-H] obsd. 655.1624, calcd. 655.1631.

2-Cvanoethyl bis(2',3'-di-O-benzoyl-2'-C-methyluridin-5'-vl) phosphate (14). 2',3'-Di-Obenzoyl-2'-C-methyluridine (0.225 g; 0.48 mmol) (9) dried over P₂O₅ overnight was dissolved in dry DCM (5 mL) under argon. Anhydrous triethylamine (0.336 mL; 2.41 mmol) and 1-chloro-1-(2-cyanoethoxy)-N.N-diisopropylphosphinamine (0.118 mL; 0.53 mmol) were added. The mixture was stirred at room temperature for 1 hour. The product was isolated by passing the mixture through a short silica gel column with dry EtOAc containing 0.5% Et₃N. The solvent was removed under reduced pressure and the product was coevaporated twice with dry acetonitrile (2 × 20 mL) to remove the traces of Et₃N. The phosphitylated nucleoside was dissolved in dry MeCN (1 mL) under argon. Nucleoside 9 (0.225 g; 0.48 mmol, dried over P₂O₅ overnight) in dry MeCN (2 mL) and tetrazole (1.92 mmol, 4.2 mL of 0.45 mol L⁻¹ solution in MeCN) were added. The mixture was stirred for 2 hours. The phosphite ester formed was oxidized with I₂ as described above form compound 10 and subjected to 5% ag NaHSO₃/DCM workup. The product was purified by Silica gel chromatography, eluting with DCM containing 5% MeOH. Compound 14 was obtained as solid in 52% yield (0.26 g). ¹H-NMR (500 MHz, CDCl₃) δ : 8.01-7.99 (m, 4H, Bz), 7.87-7.85 (m, 4H, Bz), 7.73 (t, J = 8.0 Hz, 2H, H6), 7.61-7.54 $(m, 4H, Bz), 7.44-7.38 (m, 8H, Bz), 5.99 (s, 2H, 2 \times H1'), 5.91 and 5.90 (d, <math>J = 8.0, 2H, 2 \times H5),$ 5.23-5.29 (m, 2H, $2 \times H3$ ′), 4.51-4.36 (m, 6H, $2 \times H4$ ′, H5′, 5′′), 4.29 (m, 2H, OCH_2), 2.70 (m, 2H, CH₂CN), 1.28 and 1.27 (s, 6H, $2 \times Me$). ¹³C-NMR (100 MHz, CD₃OD) δ : 168.38 (C=O Bz), 165.61 and 165.60 (C4), 161.72 (C=O Bz), 149.70 and 149.66 (C2), 139.25 (C6), 133.54, 131.95, 129.88, 128.62, 128.24, 127.57, 127.46, 126.78 (Bz), 116.67 (CN), 102.73 (C5), 91.71 (C1'), 78.94 (C2'), 78.71 (C4'), 74.25 and 74.15 (C3'), 66.43 and 66.40 (C5'), 62.67 and 62.63 (OCH₂), 20.88 (Me), 19.66 and 19.60 (CH₂CN). ³¹P-NMR (202 MHz, CDCl₃) δ: -1.85. HR-ESI-MS: [M+H]⁺ obsd. 1048.2608, calcd. 1048.2648, [M+Na]⁺ obsd. 1070.2413, calcd. 1070.2468. Bis(2'-C-methyluridin-5'-vl) phosphate (15). Compound 14 (130mg; 0.124 mmol) was dissolved in mathanolic ammonia (2.0 mL; 7 mol L⁻¹) on an ice bath. After stirring overnight (20 h) at room temperatue, the mixture was evaporated to dryness. The product was purified by RP-HPLC on a Lichrochart[®] Prep C18 column (250 x 10 mm, 5 μm, flow rate 3.0 mL min⁻¹) eluting with water that contained 3% MeCN. Compound 15 was obtained as an ammonium salt in 87% yield (62 mg). Finally, the product was converted to sodium salt as described above for compound 13. H-NMR (500 MHz, D₂O) δ : 7.85 (d, J = 8.1 Hz, 2H, H6), 5.97 (s, 2H, H1′), 5.89 (d, J = 8.1 Hz, 2H, H5), 4.30-4.33 (m, 2H, 2 x H5''), 4.13-4.17 (m, 4H, 2 x H5' and 2 x H4'),3.97 (d, J = 9.3 Hz, 2H, H3′), 1.18 (s, 6H, CH₃). ¹³C-NMR (126 MHz, D₂O) δ : 165.80 (C4), 151.41 (C2), 141.36 (C6), 102.35 (C5), 91.56 (C1'), 80.28 and 80.22 (C4'), 78.96 (C2'), 71.97

(C3´), 63.58 and 63.54 (C5´), 18.80 (Me). 31 P-NMR (202 MHz, D₂0) δ : 0.01. HR-ESI-MS: [M-H]⁻ obsd. 577.1186, calcd. 577.1189.

2'-C-Methylguanosin-5'-yl 2'-C-methyluridin-5'-yl phosphate (16). Compound 6b (0.260 g; 0.38 mmol) dried over P₂O₅ overnight was phosphitylated with 1-chloro-1-(2-cyanoethoxy)-N,Ndiisopropylphosphinamine, as described above for its 3'-O-levulinoyl analog 6a. The phosphitylated nucleoside was dissolved in dry MeCN (1 mL) under argon and 2',3'-di-Obenzoyl-2'-C-methyluridine (9, 0.18 g, 0.38 mmol, dried over P₂O₅ overnight) in dry MeCN (2 mL) and tetrazole (1.92 mmol, 4.2 mL of 0.45 mol L⁻¹ solution in MeCN) were added. The mixture was stirred for 2 hours and the phosphite ester formed was then oxidized as described above for compound 10. The product was purified by silica gel chromatography, eluting with 10% MeOH in DCM that contained 0.5% Et₃N. The 2-cyanoethyl group was cleaved during the chromatography and, hence, 2',3'-di-O-benzoyl-2'-C-methyluridin-5'-yl 3'-O-benzoyl-N²-(4methoxytrityl)-2'-C-methylguanosin-5'-yl phosphate was obtained as solid in 63% yield (0.29 g). HR-ESI-MS: [M-H] obsd. 1200.3393, calcd. 1200.3398. The benzoyl groups were then removed with ammoniacal methanol, as described above for compound 15. Silica gel chromatography gave the product in 96% yield (71 mg). ESI-MS: [M-H] obsd. 888.6, calcd. 888.8. The N^2 -(4-methoxytrityl) group was finally removed with aq 80% acetic acid (1.5 mL) at room temperature. The product was purified by HPLC and converted to Na+ salt, as described above for compound 13. The yield was 56% (27 mg). 1 H-NMR (400 MHz, D₂O) δ : 7.93 (s, 1H, H8 Guo), 7.67 (d, J = 8.4 Hz, 1H, H6 Urd), 5.91 (s, 1H, H1ouG '), 5.85 (s, 1H, H1'Urd), 5.70 (d, J = 8.4 Hz, 1H, H5 Urd), 4.10-4.38 (m, 7H, $2 \times \text{H5}''$, $2 \times \text{H5}'$, $2 \times \text{H4}'$ and H3'Guo), 3.87 (d, J= 9.6 Hz, 1H, H3'Urd), 1.13 (s, 3H, Me), 1.01 (s, 3H, Me). 13 C-NMR (100 MHz, D₂O) δ : 165.54 (C4 Urd), 158.69 (C6 Guo), 153.72 (C2 Guo), 151.06 and 150.95 (C4 Guo and C2 Urd), 140.88 (C6 Urd), 137.61 (C8 Guo), 116.09 (C5 Guo), 101.84 (C5 Urd), 91.56 and 91.15 (C1' Urd and Guo), 80.92, 80.84, 79.28 and 79.02 (C4' Urd and Guo), 79.28 and 79.02 (C2' Urd and Guo), 72.85 and 71.95 (C3´ Urd and Guo), 65.23, 65.19 64.24, 64.21 (C5´ Urd and Guo), 18.70 (2Me). ³¹P-NMR (162 MHz, D₂0) δ: 0.24. HR-ESI-MS: [M-H]⁻ obsd. 616.1418, calcd. 616.1410.

3-Acetyloxymethoxy-2,2-bis(ethoxycarbonyl)propyl bis[2',3'-di-*O*-**levulinoyl-***N*⁶-**(4-methoxytrityl)-2'-***C*-**methyladenosin-5'-yl] phosphate (18).** Compound **4** (1.82 mmol; 1.37 g) was dried over P_2O_5 for 2 days and dissolved in dry DCM (6 mL) under N_2 . Dry Et_3N (9.28 mmol; 1.29 ml) and 1,1-dichloro-*N*,*N*-diisopropylphosphinamine (0.97 mmol; 180 μl) were added and the reaction mixture was stirred at r.t. for 1,5 hours. Since according to TLC some unreacted nucleoside was still present, another portion of 1,1-dichloro-*N*,*N*-diisopropylphosphinamine (0.27 mmol; 50 μl) was added and the mixture was stirred for 30 minutes. The mixture was then passed through a short silica gel column eluting with EtOAc/hexane (8:2, ν/ν) that contained 0.5% Et_3N . The solvents were removed under reduced pressure and the residue was coevaporated two times from dry MeCN. ³¹P-NMR spectroscopy exhibited a signal at 149.66 ppm (202 MHz, CD₃CN), consistent with formation of the desired dinucleoside phosphoramidite. The compound (0.65 mmol; 0.77 g) was dissolved in dry MeCN (1 mL) and diethyl 2-acetyloxymethyloxymethyl-2-hydroxymethylmalonate¹ (1.16 mmol; 0.34 g) in dry MeCN (2

mL) and 1-H-tetrazole (1.89 mmol; 4.20 mL of 0.45 mol L⁻¹ solution in MeCN) were added under N₂ and the reaction mixture was stirred at r.t. for 1 hour. Formation of the phosphite ester was accompanied by appearance of a ³¹P-NMR signal at 140.12. Oxidation to phosphate ester was carried out as described above for compound 10. Conventional 5% ag NaHSO₃/DCM workup gave the crude product that was purified twice on a silica gel column; first elution with 3% MeOH in DCM and the second with 3% MeOH in EtOAc. Compound 18 was obtained as white solid in 44% yield (0.38 g). ¹H-NMR (500 MHz, CDCl₃) δ 8.09 (s, 1H, H8), 8.06 (s, H, H2), 8.05 (s, H, H2), 8.02 (s, 1H, H8), 7.19-7.36 (m, 24H, MMTr), 6.98 (s, 1H, NH), 6.95 (s, 1H, NH), 6.77-6.80 (m, 4H, MMTr), 6.41 (s, 1H, H1'), 6.39 (s, 1H, H1'), 5.62-5.68 (m, 2H, H4'), 5.19 (s, 2H, OC H_2 OAc), 4.56 (d, J = 5.4 Hz, 2H, POC H_2 C), 4.39-4.47 (m, 4H, H5' & H5''), 4.30 (m, 2H, H4'), 4.13-4.18 (m, 4H, MeCH₂CO), 4.11 (s, 2H, CH₂OCH₂OAc), 3.77 (s, 3H, OMe of MMTr), 3.76 (s, 3H, OMe of MMTr), 2.69-2.80 (m, 8H, MeCOCH₂CH₂ Lev), 2.53-2.65 (m, 8H, CH₂CH₂COO Lev), 2.17, 2.16 (2s, 6H, Me of Lev), 2.07, 2.06 (2s, 6H, Me of Lev), 2.02 (s, 3H, AcO), 1.37, 1.34 (2s, 6H, Me2'), 1.15-1.21 (m, 6H, MeCH₂O). ¹³C-NMR (126 MHz, CDCl₃) δ 171.91, 171.88 (CH₂COCH₃ Lev), 171.37, 171.32 (OCOCH₂ Lev), 170.28 (CO of Ac), 166.64 (COOEt), 158.30 (MMTr), 154.15 (C6), 152.42 (C2), 148.48, 148.37 (C4), 145.19, 145.15 (MMTr), 139.33, 139.25 (C8), 137.20 (MMTr), 130.23, 128.90, 127.87, 126.85 (MMTr), 121.14, 121.05 (C5), 113.14 (MMTr), 88.78 (OCH₂O), 88.06, 87.96 (C1'), 84.40, 84.08 (MMTr), 80.31, 80.25, 80.16, 80.06 (C4'), 74.62 (C3'), 70.98 (C2'), 67.22, 67.09 (C5'), 65.23, 65.20 (CCH₂OP), 62.12 (MeCH₂O), 58.98, 58.91 (CCH₂OCH₂OAc), 55.21 (MeO of MMTr), 45.23 (-C-), 37.86, 37.76 (MeCOCH₂ Lev), 29.73, 29.62 (Me of Lev), 27.68 (CH₂COO Lev), 20.91 (Me of Ac), 17.22 (Me2'), 13.90 (MeCH₂O). ³¹P-NMR (202 MHz, CDCl₃) δ -1.54. HR-ESI-MS: [M+H]⁺ obsd. 1835.6900, calcd. 1835.6805, [M+Na]⁺ obsd. 1857.6726, calcd. 1857.6624.

3-Acetyloxymethoxy-2,2-bis(ethoxycarbonyl)propyl bis(2´-C-methyladenosin-5´-yl) phosphate (19). Compound 18 (80.23 mmol; 0.34 g) was deprotected and purified as described above for compound 10 to obtain 11. Compound 19 was obtained as white solid in 57% yield (0.12 g). 1 H-NMR (500 MHz, MeOD) δ 8.22 (s, 1H, H2), 8.21 (s, 1H, H2), 8.20 (s, H, H8), 8.19 (s, H, H8), 6.10 (s, 1H, H1´), 6.10 (s, 1H, H1´), 5.22 (d, J = 6.4 Hz, 1H, OC H_2 O), 5.19 (d, J = 6.4 Hz, 1H, OC H_2 O), 4.53-4.58 (m, 6H, POC H_2 C, H5´, H5´´), 4.13-4.29 (m, 10H, H3´, H4´, MeC H_2 COO, C H_2 OCH₂OAc), 2.05 (s, 3H, AcO), 1.20 (t, J = 7.1 Hz, 3H, MeCH₂COO), 0.94 (s, 6H, 2*Me). 13 C-NMR (126 MHz, MeOD) δ 170.66 (CO of Ac), 166.56, 166.55 (COOEt), 155.94 (C6), 152.57 (C2), 148.85 (C4), 139.21 (C8), 118.94 (C5), 91.97 (C1´), 88.35 (OC H_2 O), 80.57, 80.55, 80.51, 80.50 (C4´), 78.54, 78.53 (C3´), 73.10, 73.05 (C2´), 67.56, 67.52, 67.48 (C5´), 66.72 (CC H_2 OCH₂OAc), 65.23, 65.19 (CC H_2 OP), 61.90, 61.89 (MeC H_2 O), 58.84, 58.77 (-C-), 19.50 (Me of Ac), 18.71 (2´-Me), 12.85 (MeCH₂O). 31 P-NMR (202 MHz, MeOD) δ -1.61. HR-ESI-MS: [M+H]⁺ obsd. 899.2970, calcd. 899.2931.

Pivaloyloxymethyl bis(2'-C-methyladenosin-5'-yl) phosphate (20). Compound 10 (0.14 g; 0.09 mmol) was dissolved in N-methylpyrrolidone (1 mL), triethylamine (25 μ L; 0.18 mmol) was added and the reaction mixture was incubated at 60 °C for 40 minutes. Chloromethyl

pivalate (POMCl, 25 µL, 0.18 mmol) was added and the reaction was allowed to proceed at 60 °C for 3 days. One equivalent of Et₃N (13 µL, 0.09 mmol) and POMCl (13 µL, 0.09 mmol) was added and the reaction mixture was stirred for one more day. The mixture was equilibrated between water and EtOAc. The organic layer was washed with brine and evaporated to dryness. The residue was filtered through a short silica gel column eluting with 5% MeOH in DCM. The crude product was dissolved in DCM (600 µL) and hydrazinium acetate (0.044 g, 0.48 mmol) in MeOH (200 µL) was added. The reaction mixture was stirred at r.t. for 18 hours. The unreacted hydrazinium acetate was destroyed with acetone and the reaction mixture was evaporated to dryness. The residue was dissolved in 80% aqueous acetic acid (1 mL) and the mixture was stirred at r.t. for 24 hours. The crude product was purified by HPLC as described above for compound 11. Compound 20 was obtained as white solid in 15% yield (10 mg). ¹H-NMR (500 MHz, MeOD) δ 8.20 (s, 1H, H2), 8.20 (s, 1H, H2), 8.19 (s, 2H, H8), 9.08 (s, 1H, H1'), 6.08 (s, 1H, H1 $^{\circ}$), 5.73 (d, J = 1.1 Hz, 1H, OCH₂O), 5.70 (d, J = 1.1 Hz, 1H, OCH₂O), 4.56-4.60 (m, 4H, H5', H5''), 4.22-4.28 (m, 4H, H3', H4''), 1.19 (CMe_3), 0.93, 0.92 (s, 6H, 2'-Me). 13 C-NMR(126 MHz, MeOD) δ 176.59 (CO), 155.90 (C6), 152.53 (C2), 148.82 (C4), 139.20, 139.15 (C8), 117.51 (C5), 91.94, 91.90 (C1'), 83.05, 83.01 (OCH₂O), 80.49, 80.46, 80.42 (C4'), 78.54 (C2'), 73.05, 72.86 (C3'), 67.53, 67.49, 67.39, 67.34 (C5'), 38.31 (CMe_3), 25.78 (CMe_3), 18.30 (2'-Me). ³¹P-NMR (202 MHz, CDCl₃) δ -2.50. HR-ESI-MS: [M+H]⁺ obsd. 739.2533, calcd. 739.2567, [M+Na]⁺ obsd. 761.2348, calcd. 761.2379.

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