Factors Associated with Decline of C-peptide in a Cohort of Young Children Diagnosed with Type 1
Diabetes

Andrea K. Steck<sup>1</sup>, Xiang Liu<sup>2</sup>, Jeffrey P. Krischer<sup>2</sup>, Michael J. Haller<sup>3</sup>, Riitta Veijola<sup>4</sup>, Markus Lundgren<sup>5</sup>, Simi Ahmed<sup>6</sup>, Beena Akolkar<sup>7</sup>, Jorma Toppari<sup>8</sup>, William A. Hagopian<sup>9</sup>, Marian J. Rewers<sup>1</sup> and Helena Elding Larsson<sup>5</sup>

<sup>1</sup>Barbara Davis Center for Diabetes, University of Colorado School of Medicine, Aurora, CO, USA

<sup>2</sup>Health Informatics Institute, University of South Florida, Tampa, FL, USA

<sup>3</sup>Department of Pediatrics, University of Florida, Gainesville, FL, USA

<sup>4</sup>Department of Pediatrics, PEDEGO Research Unit, MRC Oulu, Oulu University Hospital and University of Oulu, Oulu, Finland

<sup>5</sup>Department of Clinical Sciences, Lund University CRC, Skåne University Hospital, Malmö, Sweden

<sup>6</sup>Immunology of T1D, JDRF International, New York, NY, USA

<sup>7</sup>Division of Diabetes, Endocrinology & Metabolism, National Institute of Diabetes, Digestive, & Kidney Diseases, National Institutes of Health, Bethesda, Maryland, USA

<sup>8</sup>Institute of Biomedicine, Research Centre for Integrative Physiology and Pharmacology, University of Turku, and Department of Pediatrics, Turku University Hospital, Turku, Finland

<sup>9</sup>Pacific Diabetes Research Institute, Seattle, WA, USA

© The Author(s) 2020. Published by Oxford University Press on behalf of the Endocrine Society. All rights reserved. For permissions, please e-mail: journals.permissions@oup.com jc.2020-01175 See https://academic.oup.com/endocrinesociety/pages/Author\_Guidelines for Accepted Manuscript disclaimer and additional information.

Corresponding Author and person to whom reprint requests should be addressed:

Andrea Steck, MD

Barbara Davis Center for Childhood Diabetes, University of Colorado School of Medicine

1775 Aurora Court, A140, Aurora, CO 80045-6511, USA

Email: andrea.steck@cuanschutz.edu

Phone: 303-724-6769; Fax: 303-724-6779

**Funding Information:** 

The TEDDY Study is funded by U01 DK63829, U01 DK63861, U01 DK63821, U01 DK63863, U01

DK63790, UC4 DK63829, UC4 DK63861, UC4 DK63821, UC4 DK63863, UC4 DK95300, UC4 DK100238,

UC4 DK106955, UC4 DK112243, UC4 DK117483, and Contract No. HHSN267200700014C from the

National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK), National Institute of

Allergy and Infectious Diseases (NIAID), Eunice Kennedy Shriver National Institute of Child Health

and Human Development (NICHD), National Institute of Environmental Health Sciences (NIEHS),

Centers for Disease Control and Prevention (CDC), and JDRF. This work supported in part by the

NIH/NCATS Clinical and Translational Science Awards to the University of Florida (UL1 TR000064)

and the University of Colorado (UL1 TR002535). The JDRF Follow-up study is funded by grant number

17-2011-274 from the Juvenile Diabetes Research Foundation (JDRF).

**Disclosure Summary**: The authors have no relevant conflict of interest to disclose.

2

## **ABSTRACT**

**Context:** Understanding factors involved in the rate of C-peptide decline is needed to tailor therapies for type 1 diabetes (T1D).

**Objective:** Evaluate factors associated with rate of C-peptide decline after T1D diagnosis in young children.

Design: Observational study.

Setting: Academic centers.

**Participants:** 57 participants in The Environmental Determinants of Diabetes in the Young (TEDDY) enrolled at 3 months of age and followed until T1D and 56 age-matched children diagnosed with T1D in the community.

**Intervention:** A mixed meal tolerance test was used to measure the area under the curve (AUC) C-peptide at 1, 3, 6, 12 and 24 months post-diagnosis.

**Outcome:** Factors associated with rate of C-peptide decline during the first 2 years post-diagnosis were evaluated using mixed effects models adjusting for age at diagnosis and baseline C-peptide.

**Results:** Adjusted slopes of AUC C-peptide decline did not differ between TEDDY subjects and community controls (p=0.21), although the former had higher C-peptide baseline levels. In univariate analyses combining both groups (n=113), younger age, higher weight and BMI z-scores, female sex, increased number of islet autoantibodies, and IA-2A or ZnT8A positivity at baseline were associated

with higher rate of C-peptide loss. Younger age, female sex and higher weight z-score remained significant in multivariate analysis (all p<0.02). At three months after diagnosis, higher HbA1c became an additional independent factor associated with higher rate of C-peptide decline (p<0.01).

**Conclusion:** Younger age at diagnosis, female sex, higher weight z-score, and HbA1c were associated with higher rate of C-peptide decline after T1D diagnosis in young children.

Keywords: C-peptide, pediatric type 1 diabetes, new onset, beta cell decline, risk factors

### **INTRODUCTION**

Type 1 diabetes is an autoimmune disease resulting from the progressive immune-mediated destruction of beta cells. Clinical trials employing immunotherapies in type 1 diabetes have sought to prevent beta cell loss at each of stage 1, 2 and 3 of the disease, i.e. both before and after clinical diagnosis(1). Preservation of beta cell function has been associated with lower risk of hypoglycemia and lower risk of long-term complications such as microalbuminuria and retinopathy(2,3). Notably, clinical trials using rituximab (anti-CD20), abatacept (CTLA4-lg), alefacept (anti-CD2), teplizumab (anti-CD3) and low dose anti-thymocyte globulin (ATG)(4-8) have demonstrated transient preservation of C-peptide in new onset type 1 diabetes. In addition, teplizumab recently demonstrated the ability to delay, by more than 2 years, progression from Stage 2 to Stage 3 type 1 diabetes in high-risk subjects(9). In double islet autoantibody positive subjects with first phase insulin below threshold, oral insulin immunomodulation delayed progression to stage 3 by two years(10). Improved understanding of the factors involved in the rate of C-peptide decline could help tailor immunomodulatory therapies to specific groups or subjects and increase the success rates of these clinical trials(11).

An older age and a higher BMI have been previously associated with higher C-peptide levels at diagnosis of type 1 diabetes(12,13). The decline in stimulated C-peptide during the first year after the diagnosis has been highly variable from 0% to 58% (14,15) and factors involved in this variability are still only partially understood. While older age, lower HbA1c and higher BMI at diagnosis have been reported to predict a slower loss of C-peptide(16) (17,18), the SEARCH study showed a progressive decline in beta cell function independent of age, sex, HbA1c and BMI(19).

Children participating in prospective studies such as The Environmental Determinants of Diabetes in the Young (TEDDY) typically have a less severe clinical presentation at diabetes onset, with less diabetic ketoacidosis and diabetes symptoms(20,21). We have previously shown that children diagnosed through the TEDDY study have higher C-peptide levels at onset compared to community-diagnosed children, and their C-peptide levels stayed higher throughout the first 12 months following the onset of diabetes(22). The goal of the current study was to evaluate factors associated with the rate of C-peptide decline in the first 2 years after the diagnosis of clinical type 1 diabetes in a cohort of young children.

### **MATERIALS AND METHODS**

# **Study Population:**

From September 2004 to February 2010, the TEDDY study identified 8676 infants at increased risk for type 1 diabetes through genetic screening for diabetes-susceptible HLA DR-DQ genotypes at sites in Sweden, Finland, Germany, Colorado, Washington State, and Florida/Georgia. Those enrolled are followed prospectively from birth to 15 years of age, with study visits beginning at 3 months of age, then every 3 months until 4 years of age, then every 6 months thereafter. Children positive for islet autoantibodies are followed every 3 months. The details of screening and follow-up have been previously published(23,24). The JDRF Follow-up study recruited TEDDY children diagnosed with type 1 diabetes from January 2012 to December 2016; a total of 161 TEDDY children were diagnosed with type 1 diabetes during that time. For this study, 70 TEDDY children and 60 age-matched children diagnosed with type 1 diabetes in the community were enrolled by December 2016. Among the 130 enrolled subjects, 113 subjects had complete C-peptide baseline data available and were therefore included in the analysis. Control subjects from the community were matched to TEDDY subjects by age of diabetes diagnosis within one year and were required to have at least one positive islet

autoantibody at diagnosis. Type 1 diabetes was defined according to American Diabetes Association criteria for diagnosis(25).

After diagnosis of type 1 diabetes, all participants had visits with HbA1c and a Mixed Meal Tolerance Test (MMTT) within one month of onset, then at 3, 6, and 12 months after diagnosis, and bi-annually thereafter. The primary outcome measure was the area under the curve (AUC) for serum C-peptide in response to a 2-hour MMTT. The goal was to follow all subjects for at least 2 years after diagnosis or until the loss of detectable endogenous C-peptide. Parents (or legal guardian) of the subjects provided written informed consent, and the children assent when applicable. The study was approved by the ethical review boards of all participating institutions.

## Study visits:

Subjects came in fasting for MMTT, which consisted of a standardized liquid meal, Boost® High Protein (Nestle Health Care Nutrition, Inc.) given at 6 ml/kg to a maximum of 360 ml. HbA1c was measured by a Tosoh G8 HPLC Analyzer (Tosoh Bioscience Inc., San Francisco, CA) at the Diabetes Diagnostic Laboratory at the University of Missouri, Columbia. C-peptide (ng/ml) was measured using Tosoh reagents on a TOSOH 2000 autoanalyzer (Tosoh Bioscience Inc., San Francisco, CA) at the Northwest Lipid Research Laboratories at the University of Washington.

### **Islet Autoantibodies:**

Autoantibodies to GAD65, IA-2, and ZnT8 were measured in two reference laboratories by standard radiobinding assays(26). For sites in the United States, all serum samples were assayed at the Barbara Davis Center for Diabetes at the University of Colorado. In Europe, all sera were assayed at the University of Bristol, United Kingdom. Both laboratories have previously shown high assay

sensitivity and specificity, as well as concordance(27). Positive samples were reanalyzed in the other laboratory for confirmation (US samples in Europe and vice versa).

## **Statistical Analysis:**

Data were analyzed using the Statistical Analysis System software (version 9.4; SAS Institute, Cary, NC). C-peptide was measured at time points 0, 15, 30, 60, 90 and 120 minutes. These timed values were combined using the trapezoidal rule to approximate the AUC; the reported value is the AUC divided by 120 minutes, which is an estimate of the mean of the C-Peptide level over the 2-hour period. Insulin-dose adjusted HbA1c (IDAA1C), an alternate measure of residual beta-cell function(28), was calculated as HbA1c (%) + [4 x insulin dose (units/kg/day)].

Mixed-effects models were used to analyze the longitudinal data of C-peptide AUC, where AUC C-peptide values were normalized using a log-transformation on the value in the unit of ng/mL plus 1(log (x+1)). The matching of case-control was modeled as a random effect and within-subject correlation was modeled using a first-order autoregressive structure. The time from diagnosis (months) was a covariate in the models and its corresponding coefficient was interpreted as the slope of C-peptide AUC change from diagnosis (i.e., the negative value of the rate of C-peptide AUC loss). The effect of a potential factor on the rate of C-peptide AUC loss was modeled by examining an interaction term between the factor and the time from diagnosis. All the analyses were adjusted for age at diagnosis (years), C-peptide AUC at baseline and the factor being examined. C-peptide AUC at baseline and autoantibody status (number of positive autoantibodies) were the measure from the baseline visit or from the 3-month visit if the baseline data was missing.

The effect of each of the potential factors on the rate of C-peptide AUC loss was examined individually. Factors with p<0.05 for their effects on the slope were further analyzed in multivariate analysis using backward elimination procedure. In the backward elimination procedure, factors

having effects on the rate of C-peptide AUC loss with p>0.10 were eliminated. All participants with missing data specified as part of a particular analysis were omitted from that analysis. Two-tailed p-values <0.05 were considered to be statistically significant.

### **RESULTS**

Characteristics at diagnosis of diabetes of the 57 TEDDY and 56 community control children are described in Table 1. Mean age of diagnosis was 6.5 ± 1.8 years in these children and did not differ between TEDDY cases and community controls. As expected, TEDDY children had a higher frequency of the high-risk HLA DR3/4 genotype (56% vs 18%, p<0.001). None of the TEDDY children presented with diabetic ketoacidosis (DKA) at diagnosis, while 16% of the community children did (p=0.001), with an overall low frequency of DKA for this cohort of young children. The low rate of DKA in the community group could potentially be explained by the willingness of more medically-aware or committed community control families to enroll in this intensive follow-up study with multiple MMTTs during the first year post-diagnosis. The mean weight and BMI z-scores were higher at diagnosis in TEDDY children vs community controls (0.6 vs 0.1, p=0.02 and 0.2 vs -0.4, p=0.01 respectively). TEDDY children had a lower HbA1c at diagnosis and higher C-peptide AUC at baseline than community controls (6.9% (52 mmol/mol) vs 10.2% (88 mmol/mol), p<0.001 and 1.7 ng/mL vs 1.3 ng/mL, p=0.007 respectively).

Individual trajectories of AUC C-peptide over time in TEDDY children and community controls are shown in Figure 1. The slope of C-peptide AUC change was -0.030 per month overall (95% CI: -0.033 - 0.028) in log scale, adjusted for age at diagnosis and AUC C-peptide at baseline. Adjusted slopes of C-peptide AUC change did not differ between TEDDY subjects and community controls (slope=-0.032 vs. -0.029 per month respectively, p=0.208).

The following factors were examined for association with rate of C-peptide loss: sex, first-degree relative with type 1 diabetes, HLA DR3-DQ2/DR4-DQ8 genotype, age at diagnosis, presence of DKA and/or diabetes symptoms at diagnosis as well as HbA1c, weight, height and BMI z-scores at diagnosis. Additional factors from the baseline and 3 months visits included autoantibody status (IA-2A, GADA and ZnT8A), C-peptide, HbA1c and insulin dose-adjusted HbA1c. In univariate analyses at baseline, C-peptide, female gender, younger age, higher weight and BMI z-scores as well as increased number of autoantibodies, presence of IA-2A or ZnT8A autoantibodies were all associated with higher rate of C-peptide loss; only factors that were significantly associated with higher rate of C-peptide loss are shown in Table 2. Three of these eight factors remained significant in multivariate analysis adjusted for C-peptide at baseline: younger age at diagnosis, female gender and higher weight z-score (all p < 0.02, Table 3).

The same univariate and multivariate analyses at 3 months after diagnosis are shown in Tables 4 and 5. At that time point, C-peptide, female gender, younger age, higher weight and BMI, increased number of autoantibodies, IA-2A positivity and ZnT8A positivity as well as higher HbA1c were all associated with higher rate of C-peptide loss (Table 4). In multivariate adjusted analyses, higher HbA1c, in addition to younger age, female gender and higher weight, was also associated with higher rate of C-peptide decline (all p < 0.02, Table 5).

### DISCUSSION

In this international cohort of children diagnosed with type 1 diabetes at a young age (mean age 6.5 years), we have analyzed factors involved in rate of C-peptide decline during the first 2 years after the onset of disease. We have previously shown that children diagnosed through the TEDDY study have higher C-peptide levels at onset compared to community-diagnosed children and that C-peptide levels stay higher throughout the first 12 months following the onset of type 1 diabetes(22).

Here we found that rates of C-peptide AUC loss (slope), adjusted for age at diagnosis and C-peptide AUC at baseline, did not differ between TEDDY subjects and community controls, and that younger age at diagnosis, female sex, higher weight z-score and higher HbA1c were associated with faster decline of C-peptide over the first 2 years after diagnosis.

While age and BMI have been associated with C-peptide levels at diagnosis(13,17,18,29), rate of Cpeptide decline after diagnosis has been highly variable in previous studies and factors involved in rate of C-peptide decline have not been consistent across studies. Data from 481 individuals with recent onset type 1 diabetes enrolled in TrialNet studies showed that age at diagnosis and baseline C-peptide were significant predictors of rate of C-peptide loss (11). A study looking at rate of stimulated C-peptide decline in 446 children with new onset type 1 diabetes from Scandinavia, Europe and North America between 1982 and 2009 found that both initial C-peptide and rate of Cpeptide decline seemed to have increased over this 27-year time period(30). In that study, younger age, positivity for GADA, IAA, or both, but not BMI z-score, gender nor initial C-peptide, were associated with faster rate of C-peptide decline during the 15 months after diagnosis. Of note, mean age varied from 9 years to 12.8 years for the 5 cohorts included, while this cohort of children was younger (mean age of 6.5 years at diagnosis). After adjusting for age at diagnosis and C-peptide AUC at baseline, IA-2A positivity and ZnT8A positivity in this study were associated with higher rate of Cpeptide loss in univariate analyses (< 0.01), with p=0.06 in multivariate analyses. Islet autoantibody positivity and levels have been shown to be associated with rate of progression to type 1 diabetes in children followed in prospective birth cohort studies such as TEDDY and the Diabetes Autoimmunity Study in the Young (DAISY) (31,32). Therefore, it seems likely that ongoing autoimmunity as noted by autoantibody positivity would also play a role in rate of beta-cell loss after diagnosis.

Preservation of C-peptide is known to be associated with lower risk of hypoglycemia and lower risk of long-term microvascular complications(2,3). More recently, a study showed that diabetic ketoacidosis (DKA) at diagnosis of type 1 diabetes in children predicts persistently elevated HbA1c levels and poor long-term glycemic control independent of demographic and socioeconomic factors(33). In our current study, both higher HbA1c and higher weight z-score were associated with higher rate of C-peptide decline over the first 2 years after diagnosis, suggesting that metabolic factors could also play a role in preservation of beta-cell function. Diagnosis at a young age and poor metabolic control (higher HbA1c) could imply a more severe autoimmune process leading to early presentation of the disease in those at risk. While some studies have reported preserved beta cell function with higher BMI close to diabetes onset(16), other studies have shown greater C-peptide decline over 1 year post-diagnosis with higher BMI(18,34) or no effect of BMI on the rate of Cpeptide decline post-diagnosis(17,19). At diagnosis of diabetes in subjects < 18 years old, HbA1c levels have been shown to inversely correlate with AUC C-peptide as well as measures of the timing of C-peptide responses in OGTT, including peak C-peptide and early and late C-peptide responses(35). These various metabolic measures are obviously intertwined, but may help develop endpoints for beta cell preservation trials and tailor therapies to specific individuals or groups of individuals.

Limitations of this study include a relatively small number of subjects diagnosed with type 1 diabetes with follow-up limited to 2 years after diagnosis. This could influence the results of some of the factors that were borderline, such as IA-2A and ZnT8A positivity. In addition, as baseline visits happened about a month after diagnosis, IAA levels were not available for community control subjects and therefore could not be analyzed and were not included in the analysis of either group. Although the rate of C-peptide decline was not statistically different between the two groups, there are important differences between the groups, such as higher type 1 diabetes genetic load and earlier diagnosis in TEDDY subjects. Therefore, it is possible that known or unknown factors that regulate beta-cell function decline were different between these two groups.

In summary, this study of young children diagnosed with type 1 diabetes shows that adjusted rates of C-peptide loss were similar between TEDDY subjects and community controls. Younger age at diagnosis, female gender, higher weight z-score and higher HbA1c were associated with higher rate of C-peptide decline over the first 2 years after diagnosis, while positivity for IA-2A and ZnT8A autoantibodies might also contribute to C-peptide decline in young children.

### **ACKNOWLEDGMENTS**

Author Contributions: A.K.S designed the study, wrote manuscript, contributed to discussion, X.L. researched data, reviewed/edited manuscript, J.K. designed the study, contributed to discussion, reviewed/edited manuscript, M.J.H. reviewed/edited manuscript, R.V. contributed to discussion, reviewed/edited manuscript, M.L. contributed to discussion, reviewed/edited manuscript, S.A. contributed to discussion, reviewed/edited manuscript, B.A. contributed to discussion, reviewed/edited manuscript, J.T. contributed to discussion, reviewed/edited manuscript, W.A.H reviewed/edited manuscript, M.J.R designed the study, contributed to discussion, reviewed/edited manuscript, H.E.L. contributed to discussion, reviewed/edited manuscript.

The authors have no relevant conflict of interest to disclose. AKS takes full responsibility for the contents of the article.

The TEDDY Study is funded by U01 DK63829, U01 DK63861, U01 DK63821, U01 DK63821, U01 DK63863, U01 DK63790, UC4 DK63829, UC4 DK63861, UC4 DK63821, UC4 DK63863, UC4 DK95300, UC4 DK100238, UC4 DK106955, UC4 DK112243, UC4 DK117483, and Contract No. HHSN267200700014C from the National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK), National Institute of Allergy and Infectious Diseases (NIAID), Eunice Kennedy Shriver National Institute of Child Health and Human Development (NICHD), National Institute of Environmental Health Sciences (NIEHS), Centers for Disease Control and Prevention (CDC), and JDRF. This work supported in part by the NIH/NCATS Clinical and Translational Science Awards to the University of Florida (UL1 TR000064) and the University of Colorado (UL1 TR002535).

The JDRF Follow-up study is funded by grant number 17-2011-274 from the Juvenile Diabetes Research Foundation (JDRF).

## **The TEDDY Study Group**

Colorado Clinical Center: Marian Rewers, M.D., Ph.D., Pl. 1,4,5,6,9,10, Aaron Barbour, Kimberly Bautista 11, Judith Baxter 8,911, Daniel Felipe-Morales, Kimberly Driscoll, Ph.D. 8, Brigitte I. Frohnert, M.D. 2,13, Marisa Stahl, M.D. 12, Patricia Gesualdo 2,6,11,13, Michelle Hoffman 11,12,13, Rachel Karban 11, Edwin Liu, M.D. 12, Jill Norris, Ph.D. 2,3,11, Stesha Peacock, Hanan Shorrosh, Andrea Steck, M.D. 3,13, Megan Stern, Erica Villegas 2, Kathleen Waugh 6,7,11. University of Colorado, Anschutz Medical Campus, Barbara Davis Center for Childhood Diabetes.

Finland Clinical Center: Jorma Toppari, M.D., Ph.D., Pl<sup>x1,4,10,13</sup>, Olli G. Simell, M.D., Ph.D., Annika Adamsson, Ph.D.<sup>11</sup>, Suvi Ahonen\*<sup>±§</sup>, Mari Åkerlund\*<sup>±§</sup>, Leena Hakola\*, Anne Hekkala, M.D.<sup>μμ</sup>, Henna Holappa<sup>μμ</sup>, Heikki Hyöty, M.D., Ph.D.\*<sup>±6</sup>, Anni Ikonen<sup>μμ</sup>, Jorma Ilonen, M.D., Ph.D.<sup>½13</sup>, Sinikka Jäminki\*<sup>±</sup>, Sanna Jokipuuˆ, Leena Karlssonˆ, Jukka Kero M.D., Ph.D.<sup>½↑</sup>, Miia Kähönen<sup>μμ11,13</sup>, Mikael Knip, M.D., Ph.D.\*<sup>±5</sup>, Minna-Liisa Koivikko<sup>μμ</sup>, Merja Koskinen\*<sup>±</sup>, Mirva Koreasalo\*<sup>±§2</sup>, Kalle Kurppa, M.D., Ph.D.\*<sup>±12</sup>, Jarita Kytölä\*<sup>±</sup>, Tiina Latva-aho<sup>μμ</sup>, Katri Lindfors, Ph.D.\*<sup>12</sup>, Maria Lönnrot, M.D., Ph.D.\*<sup>±6</sup>, Elina Mäntymäkiˆ, Markus Mattila\*, Maija Miettinen<sup>§2</sup>, Katja Multasuo<sup>μμ</sup>, Teija Mykkänen<sup>μμ</sup>, Tiina Niininen<sup>±\*11</sup>, Sari Niinistö<sup>±§2</sup>, Mia Nyblom\*<sup>±</sup>, Sami Oikarinen, Ph.D.\*<sup>±</sup>, Paula Ollikainen<sup>μμ</sup>, Zhian Othmaniˆ, Sirpa Pohjola <sup>μμ</sup>, Petra Rajalaˆ, Jenna Rautanen<sup>±§</sup>, Anne Riikonen\*<sup>±§2</sup>, Eija Riskiˆ, Miia Pekkola\*<sup>±</sup>, Minna Romoˆ, Satu Ruohonenˆ, Satu Simell, M.D., Ph.D.<sup>½12</sup>, Maija Sjöbergˆ, Aino Stenius<sup>μμ11</sup>, Päivi Tossavainen, M.D.<sup>μμ</sup>, Mari Vähä-Mäkilä<sup>¥</sup>, Sini Vainionpäaˆ, Eeva Varjonen<sup>111</sup>, Riitta Veijola, M.D., Ph.D.<sup>μμ13</sup>, Irene Viinikangas<sup>μμ</sup>, Suvi M. Virtanen, M.D., Ph.D.\*<sup>±§2</sup>. <sup>½</sup>University of Turku, \*Tampere University, <sup>μ</sup>University of Oulu, ˆTurku University Hospital, Hospital District of Southwest Finland, <sup>‡</sup>Tampere University Hospital, <sup>§</sup>Oulu University Hospital, §National Institute for Health and Welfare, Finland, <sup>†</sup>University of Kuopio.

Georgia/Florida Clinical Center: Jin-Xiong She, Ph.D., Pl<sup>1,3,4,10</sup>, Desmond Schatz, M.D.\*<sup>4,5,7,8</sup>, Diane Hopkins<sup>11</sup>, Leigh Steed<sup>11,12,13</sup>, Jennifer Bryant<sup>11</sup>, Katherine Silvis<sup>2</sup>, Michael Haller, M.D.\*<sup>13</sup>, Melissa Gardiner<sup>11</sup>, Richard McIndoe, Ph.D., Ashok Sharma, Stephen W. Anderson, M.D.<sup>^</sup>, Laura Jacobsen, M.D.\*<sup>13</sup>, John Marks, DHSc.\*<sup>11,13</sup>, P.D. Towe\*. Center for Biotechnology and Genomic Medicine, Augusta University. \*University of Florida, ^Pediatric Endocrine Associates, Atlanta.

Germany Clinical Center: Anette G. Ziegler, M.D., Pl<sup>1,3,4,10</sup>, Ezio Bonifacio Ph.D.\*<sup>5</sup>, Cigdem Gezginci, Anja Heublein, Eva Hohoff<sup>¥2</sup>, Sandra Hummel, Ph.D.<sup>2</sup>, Annette Knopff<sup>7</sup>, Charlotte Koch, Sibylle Koletzko, M.D.<sup>¶12</sup>, Claudia Ramminger<sup>11</sup>, Roswith Roth, Ph.D.<sup>8</sup>, Jennifer Schmidt, Marlon Scholz, Joanna Stock<sup>8,11,13</sup>, Katharina Warncke, M.D.<sup>13</sup>, Lorena Wendel, Christiane Winkler, Ph.D.<sup>2,11</sup>. Forschergruppe Diabetes e.V. and Institute of Diabetes Research, Helmholtz Zentrum München, Forschergruppe Diabetes, and Klinikum rechts der Isar, Technische Universität München. \*Center for Regenerative Therapies, TU Dresden, <sup>¶</sup>Dr. von Hauner Children's Hospital, Department of Gastroenterology, Ludwig Maximillians University Munich, <sup>¥</sup>University of Bonn, Department of Nutritional Epidemiology.

Sweden Clinical Center: Åke Lernmark, Ph.D., Pl. 1,3,4,5,6,8,9,10, Daniel Agardh, M.D., Ph.D. 6,12, Carin Andrén Aronsson, Ph.D. 2,11,12, Maria Ask, Rasmus Bennet, Corrado Cilio, Ph.D., M.D. 5,6, Susanne Dahlberg, Helene Engqvist, Emelie Ericson-Hallström, Annika Björne Fors, Lina Fransson, Thomas Gard, Monika Hansen, Hanna Jisser, Fredrik Johansen, Berglind Jonsdottir, M.D., Ph.D. 11, Helena Elding Larsson, M.D., Ph.D. 6,13, Marielle Lindström, Markus Lundgren, M.D., Ph.D. 13, Marlena Maziarz, Ph.D., Maria Månsson-Martinez, Jessica Melin 11, Zeliha Mestan, Caroline Nilsson, Karin Ottosson, Kobra Rahmati, Anita Ramelius, Falastin Salami, Anette Sjöberg, Birgitta Sjöberg, Carina Törn, Ph.D. 3, Åsa Wimar 13. Lund University.

<u>Washington Clinical Center:</u> William A. Hagopian, M.D., Ph.D., Pl<sup>1,3,4,5,6,7,10,12,13</sup>, Michael Killian<sup>6,7,11,12</sup>, Claire Cowen Crouch<sup>11,13</sup>, Jennifer Skidmore<sup>2</sup>, Masumeh Chavoshi, Arlene Meyer, Jocelyn Meyer, Denise Mulenga<sup>11</sup>, Nole Powell, Jared Radtke, Matei Romancik, Shreya Roy, Davey Schmitt, Sarah Zink. Pacific Northwest Research Institute.

<u>Pennsylvania Satellite Center:</u> Dorothy Becker, M.D., Margaret Franciscus, MaryEllen Dalmagro-Elias Smith<sup>2</sup>, Ashi Daftary, M.D., Mary Beth Klein, Chrystal Yates. Children's Hospital of Pittsburgh of UPMC.

Data Coordinating Center: Jeffrey P. Krischer, Ph.D.,Pl<sup>1,4,5,9,10</sup>, Sarah Austin-Gonzalez, Maryouri Avendano, Sandra Baethke, Brant Burkhardt, Ph.D.<sup>5,6</sup>, Martha Butterworth<sup>2</sup>, Joanna Clasen, David Cuthbertson, Christopher Eberhard, Steven Fiske<sup>8</sup>, Jennifer Garmeson, Veena Gowda, Kathleen Heyman, Belinda Hsiao, Christina Karges, Francisco Perez Laras, Qian Li, Ph.D.<sup>2,3</sup>, Shu Liu, Xiang Liu, Ph.D.<sup>2,3,8,13</sup>, Kristian Lynch, Ph.D.<sup>5,6,8</sup>, Colleen Maguire, Jamie Malloy, Cristina McCarthy<sup>11</sup>, Hemang Parikh, Ph.D.<sup>3</sup>, Cassandra Remedios, Chris Shaffer, Laura Smith, Ph.D.<sup>8,11</sup>, Susan Smith<sup>11</sup>, Noah Sulman, Ph.D., Roy Tamura, Ph.D.<sup>1,2,11,12,13</sup>, Dena Tewey, Michael Toth, Ulla Uusitalo, Ph.D.<sup>2</sup>, Kendra Vehik, Ph.D.<sup>4,5,6,8,13</sup>, Ponni Vijayakandipan, Jimin Yang, Ph.D., R.D.<sup>2</sup>. Past staff: Michael Abbondondolo, Lori Ballard, Rasheedah Brown, Stephen Dankyi, David Hadley, Ph.D., Hye-Seung Lee, Ph.D., Wendy McLeod, Aubrie Merrell, Steven Meulemans, Ryan Quigley. University of South Florida.

<u>Autoantibody Reference Laboratories:</u> Liping Yu, M.D.<sup>5</sup>, Dongmei Miao, M.D.<sup>^</sup>, Polly Bingley, M.D., FRCP\*<sup>5</sup>, Alistair Williams\*, Kyla Chandler\*, Ilana Kelland\*, Yassin Ben Khoud\*, Huma Zahid\*, Matthew Randell \*. Barbara Davis Center for Childhood Diabetes, University of Colorado Denver, \*Bristol Medical School, University of Bristol, UK.

**HbA1c Laboratory:** Randie R. Little, Ph.D., Curt Rohlfing. Diabetes Diagnostic Laboratory, Dept. of Pathology, University of Missouri School of Medicine.

<u>HLA Reference Laboratory:</u> William Hagopian<sup>3</sup>, MD, PhD, Masumeh Chavoshi, Jared Radtke, Sarah Zink. Pacific Northwest Research Institute, Seattle WA. (Previously Henry Erlich, Ph.D.<sup>3</sup>, Steven J. Mack, Ph.D., Anna Lisa Fear. Center for Genetics, Children's Hospital Oakland Research Institute.)

<u>OGTT/MMTT Laboratory:</u> Santica M. Marcovina, Ph.D., Sc.D., Andrew N. Hoofnagle, M.D., Ph.D., Northwest Lipid Metabolism and Diabetes Research Laboratories, University of Washington.

**Repository:** Sandra Ke, Niveen Mulholland, Ph.D. NIDDK Biosample Repository at Fisher BioServices.

<u>Project scientist:</u> Beena Akolkar, Ph.D.<sup>1,3,4,5,6,7,9,10</sup>. National Institutes of Diabetes and Digestive and Kidney Diseases.

<u>Other contributors:</u> Kasia Bourcier, Ph.D.<sup>5</sup>, National Institutes of Allergy and Infectious Diseases. Thomas Briese, Ph.D.<sup>6</sup>, Columbia University. Suzanne Bennett Johnson, Ph.D.<sup>8,11</sup>, Florida State University. Eric Triplett, Ph.D.<sup>6</sup>, University of Florida.

### **Committees:**

<sup>1</sup>Ancillary Studies, <sup>2</sup>Diet, <sup>3</sup>Genetics, <sup>4</sup>Human Subjects/Publicity/Publications, <sup>5</sup>Immune Markers, <sup>6</sup>Infectious Agents, <sup>7</sup>Laboratory Implementation, <sup>8</sup>Psychosocial, <sup>9</sup>Quality Assurance, <sup>10</sup>Steering, <sup>11</sup>Study Coordinators, <sup>12</sup>Celiac Disease, <sup>13</sup>Clinical Implementation.

#### References

- 1. Insel RA, Dunne JL, Atkinson MA, Chiang JL, Dabelea D, Gottlieb PA, Greenbaum CJ, Herold KC, Krischer JP, Lernmark A, Ratner RE, Rewers MJ, Schatz DA, Skyler JS, Sosenko JM, Ziegler AG. Staging Presymptomatic Type 1 Diabetes: A Scientific Statement of JDRF, the Endocrine Society, and the American Diabetes Association. Diabetes Care 2015; 38:1964-1974
- 2. Steffes MW, Sibley S, Jackson M, Thomas W. Beta-cell function and the development of diabetes-related complications in the diabetes control and complications trial. Diabetes Care 2003; 26:832-836
- 3. Lachin JM, McGee P, Palmer JP, Group DER. Impact of C-peptide preservation on metabolic and clinical outcomes in the Diabetes Control and Complications Trial. Diabetes 2014; 63:739-748
- 4. Pescovitz MD, Greenbaum CJ, Krause-Steinrauf H, Becker DJ, Gitelman SE, Goland R, Gottlieb PA, Marks JB, McGee PF, Moran AM, Raskin P, Rodriguez H, Schatz DA, Wherrett D, Wilson DM, Lachin JM, Skyler JS. Rituximab, B-lymphocyte depletion, and preservation of beta-cell function. N Engl J Med 2009; 361:2143-2152
- 5. Orban T, Bundy B, Becker DJ, DiMeglio LA, Gitelman SE, Goland R, Gottlieb PA, Greenbaum CJ, Marks JB, Monzavi R, Moran A, Raskin P, Rodriguez H, Russell WE, Schatz D, Wherrett D, Wilson DM, Krischer JP, Skyler JS, Type 1 Diabetes TrialNet Abatacept Study G. Co-stimulation modulation with abatacept in patients with recent-onset type 1 diabetes: a randomised, double-blind, placebo-controlled trial. Lancet 2011; 378:412-419
- 6. Rigby MR, Harris KM, Pinckney A, DiMeglio LA, Rendell MS, Felner EI, Dostou JM, Gitelman SE, Griffin KJ, Tsalikian E, Gottlieb PA, Greenbaum CJ, Sherry NA, Moore WV, Monzavi R, Willi SM, Raskin P, Keyes-Elstein L, Long SA, Kanaparthi S, Lim N, Phippard D, Soppe CL, Fitzgibbon ML, McNamara J, Nepom GT, Ehlers MR. Alefacept provides sustained clinical and immunological effects in new-onset type 1 diabetes patients. J Clin Invest 2015; 125:3285-3296
- 7. Hagopian W, Ferry RJ, Jr., Sherry N, Carlin D, Bonvini E, Johnson S, Stein KE, Koenig S, Daifotis AG, Herold KC, Ludvigsson J, Protege Trial I. Teplizumab preserves C-peptide in recent-onset type 1 diabetes: two-year results from the randomized, placebo-controlled Protege trial. Diabetes 2013; 62:3901-3908
- 8. Haller MJ, Long SA, Blanchfield JL, Schatz DA, Skyler JS, Krischer JP, Bundy BN, Geyer SM, Warnock MV, Miller JL, Atkinson MA, Becker DJ, Baidal DA, DiMeglio LA, Gitelman SE, Goland R, Gottlieb PA, Herold KC, Marks JB, Moran A, Rodriguez H, Russell WE, Wilson DM, Greenbaum CJ, Type 1 Diabetes TrialNet ATGGSG. Low-Dose Anti-Thymocyte Globulin Preserves C-Peptide, Reduces HbA1c, and Increases Regulatory to Conventional T-Cell Ratios in New-Onset Type 1 Diabetes: Two-Year Clinical Trial Data. Diabetes 2019; 68:1267-1276
- 9. Herold KC, Bundy BN, Long SA, Bluestone JA, DiMeglio LA, Dufort MJ, Gitelman SE, Gottlieb PA, Krischer JP, Linsley PS, Marks JB, Moore W, Moran A, Rodriguez H, Russell WE, Schatz D, Skyler JS, Tsalikian E, Wherrett DK, Ziegler AG, Greenbaum CJ, Type 1 Diabetes TrialNet Study G. An Anti-CD3 Antibody, Teplizumab, in Relatives at Risk for Type 1 Diabetes. N Engl J Med 2019; 381:603-613
- **10.** Writing Committee for the Type 1 Diabetes TrialNet Oral Insulin Study G, Krischer JP, Schatz DA, Bundy B, Skyler JS, Greenbaum CJ. Effect of Oral Insulin on

- Prevention of Diabetes in Relatives of Patients With Type 1 Diabetes: A Randomized Clinical Trial. JAMA 2017; 318:1891-1902
- **11.** Bundy BN, Krischer JP, Type 1 Diabetes TrialNet Study G. A model-based approach to sample size estimation in recent onset type 1 diabetes. Diabetes Metab Res Rev 2016; 32:827-834
- 12. Xu P, Qian X, Schatz DA, Cuthbertson D, Krischer JP, Group DPTS. Distribution of C-peptide and its determinants in North American children at risk for type 1 diabetes. Diabetes Care 2014; 37:1959-1965
- 13. Sosenko JM, Geyer S, Skyler JS, Rafkin LE, Ismail HM, Libman IM, Liu YF, DiMeglio LA, Evans-Molina C, Palmer JP. The influence of body mass index and age on C-peptide at the diagnosis of type 1 diabetes in children who participated in the diabetes prevention trial-type 1. Pediatr Diabetes 2018; 19:403-409
- **14.** Palmer JP. C-peptide in the natural history of type 1 diabetes. Diabetes Metab Res Rev 2009; 25:325-328
- **15.** Brown RJ, Sinaii N, Rother KI. Too much glucagon, too little insulin: time course of pancreatic islet dysfunction in new-onset type 1 diabetes. Diabetes Care 2008; 31:1403-1404
- **16.** Greenbaum CJ, Anderson AM, Dolan LM, Mayer-Davis EJ, Dabelea D, Imperatore G, Marcovina S, Pihoker C, Group SS. Preservation of beta-cell function in autoantibody-positive youth with diabetes. Diabetes Care 2009; 32:1839-1844
- 17. Hao W, Gitelman S, DiMeglio LA, Boulware D, Greenbaum CJ, Type 1 Diabetes TrialNet Study G. Fall in C-Peptide During First 4 Years From Diagnosis of Type 1 Diabetes: Variable Relation to Age, HbA1c, and Insulin Dose. Diabetes Care 2016; 39:1664-1670
- 18. Ludvigsson J, Carlsson A, Deli A, Forsander G, Ivarsson SA, Kockum I, Lindblad B, Marcus C, Lernmark A, Samuelsson U. Decline of C-peptide during the first year after diagnosis of Type 1 diabetes in children and adolescents. Diabetes Res Clin Pract 2013; 100:203-209
- 19. Dabelea D, Mayer-Davis EJ, Andrews JS, Dolan LM, Pihoker C, Hamman RF, Greenbaum C, Marcovina S, Fujimoto W, Linder B, Imperatore G, D'Agostino R, Jr. Clinical evolution of beta cell function in youth with diabetes: the SEARCH for Diabetes in Youth study. Diabetologia 2012; 55:3359-3368
- 20. Elding Larsson H, Vehik K, Bell R, Dabelea D, Dolan L, Pihoker C, Knip M, Veijola R, Lindblad B, Samuelsson U, Holl R, Haller MJ, Group TS, Group SS, Swediabkids Study G, Group DPVS, Finnish Diabetes Registry Study G. Reduced prevalence of diabetic ketoacidosis at diagnosis of type 1 diabetes in young children participating in longitudinal follow-up. Diabetes Care 2011; 34:2347-2352
- **21.** Elding LH, Vehik K, Gesualdo P, Akolkar B, Hagopian W, Krischer J, Lernmark A, Rewers M, Simell O, She JX, Ziegler A, Haller MJ. Children followed in the TEDDY study are diagnosed with type 1 diabetes at an early stage of disease. Pediatr Diabetes 2014; 15:118-126
- 22. Steck AK, Larsson HE, Liu X, Veijola R, Toppari J, Hagopian WA, Haller MJ, Ahmed S, Akolkar B, Lernmark A, Rewers MJ, Krischer JP, and the TSG. Residual beta-cell function in diabetes children followed and diagnosed in the TEDDY study compared to community controls. Pediatr Diabetes 2017; 18:794-802
- **23.** Kiviniemi M, Hermann R, Nurmi J, Ziegler AG, Knip M, Simell O, Veijola R, Lovgren T, Ilonen J. A high-throughput population screening system for the estimation of genetic risk for type 1 diabetes: an application for the TEDDY (the Environmental Determinants of Diabetes in the Young) study. Diabetes Technol Ther 2007; 9:460-472

- 24. Hagopian WA, Erlich H, Lernmark A, Rewers M, Ziegler AG, Simell O, Akolkar B, Vogt R, Jr., Blair A, Ilonen J, Krischer J, She J. The Environmental Determinants of Diabetes in the Young (TEDDY): genetic criteria and international diabetes risk screening of 421 000 infants. Pediatr Diabetes 2011; 12:733-743
- **25.** Diagnosis and classification of diabetes mellitus. Diabetes Care 2014; 37 Suppl 1:S81-S90
- 26. Bonifacio E, Yu L, Williams AK, Eisenbarth GS, Bingley PJ, Marcovina SM, Adler K, Ziegler AG, Mueller PW, Schatz DA, Krischer JP, Steffes MW, Akolkar B. Harmonization of glutamic acid decarboxylase and islet antigen-2 autoantibody assays for national institute of diabetes and digestive and kidney diseases consortia. J Clin Endocrinol Metab 2010; 95:3360-3367
- 27. Torn C, Mueller PW, Schlosser M, Bonifacio E, Bingley PJ. Diabetes Antibody Standardization Program: evaluation of assays for autoantibodies to glutamic acid decarboxylase and islet antigen-2. Diabetologia 2008; 51:846-852
- 28. Mortensen HB, Hougaard P, Swift P, Hansen L, Holl RW, Hoey H, Bjoerndalen H, de Beaufort C, Chiarelli F, Danne T, Schoenle EJ, Aman J, Hvidoere Study Group on Childhood D. New definition for the partial remission period in children and adolescents with type 1 diabetes. Diabetes Care 2009; 32:1384-1390
- **29.** Wallensteen M, Dahlquist G, Persson B, Landin-Olsson M, Lernmark A, Sundkvist G, Thalme B. Factors influencing the magnitude, duration, and rate of fall of B-cell function in Type I (insulin-dependent) diabetic children followed for two years from their clinical diagnosis. Diabetologia 1988; 31:664-669
- 30. Max Andersen ML, Nielsen LB, Svensson J, Porksen S, Hougaard P, Beam C, Greenbaum C, Becker D, Petersen JS, Hansen L, Mortensen HB. Disease progression among 446 children with newly diagnosed type 1 diabetes located in Scandinavia, Europe, and North America during the last 27 yr. Pediatr Diabetes 2014; 15:345-354
- 31. Steck AK, Vehik K, Bonifacio E, Lernmark A, Ziegler AG, Hagopian WA, She J, Simell O, Akolkar B, Krischer J, Schatz D, Rewers MJ. Predictors of Progression From the Appearance of Islet Autoantibodies to Early Childhood Diabetes: The Environmental Determinants of Diabetes in the Young (TEDDY). Diabetes Care 2015; 38:808-813
- 32. Steck AK, Johnson K, Barriga KJ, Miao D, Yu L, Hutton JC, Eisenbarth GS, Rewers MJ. Age of Islet Autoantibody Appearance and Mean Levels of Insulin, but Not GAD or IA-2 Autoantibodies, Predict Age of Diagnosis of Type 1 Diabetes: Diabetes Autoimmunity Study in the Young. Diabetes Care 2011; 34:1397-1399
- **33.** Duca LM, Wang B, Rewers M, Rewers A. Diabetic Ketoacidosis at Diagnosis of Type 1 Diabetes Predicts Poor Long-term Glycemic Control. Diabetes Care 2017; 40:1249-1255
- **34.** Lauria A, Barker A, Schloot N, Hosszufalusi N, Ludvigsson J, Mathieu C, Mauricio D, Nordwall M, Van der Schueren B, Mandrup-Poulsen T, Scherbaum WA, Weets I, Gorus FK, Wareham N, Leslie RD, Pozzilli P. BMI is an important driver of beta-cell loss in type 1 diabetes upon diagnosis in 10 to 18-year-old children. Eur J Endocrinol 2015; 172:107-113
- 35. Ismail HM, Evans-Molina C, DiMeglio LA, Becker DJ, Libman I, Sims EK, Boulware D, Herold KC, Rafkin L, Skyler J, Cleves MA, Palmer J, Sosenko JM, Type 1 Diabetes Trial N, Diabetes Prevention Trial-Type-1 Study G. Associations of HbA1c with the timing of C-peptide responses during the oral glucose tolerance test at the diagnosis of type 1 diabetes. Pediatr Diabetes 2019; 20:408-413

**Table 1: Characteristics of Study Participants** 

	TEDDY Case (N=57)	Control (N=56)	P value*
Female, N (%)	25 (43.9)	30 (53.6)	0.349
FDR with T1D+, N (%)	9 (15.8)	5 (8.9)	0.393
HLA DR3/4, DQB1*0302, N (%)	32 (56.1)	10 (17.9)	<0.001
Age at Diagnosis (years)	6.4 <u>+</u> 1.8	6.7 <u>+</u> 1.9	0.378
Diabetic ketoacidosis, N (%)	0 (0.0)	9 (16.1)	0.001
Weight z-score at diagnosis	0.6 ± 0.9	0.1 ± 0.9	0.021
Height z-score at diagnosis‡	0.8 ± 1.0	0.6 ± 1.0	0.208
BMI z-score at diagnosis‡	0.2 ± 1.1	-0.4 ± 1.2	0.010
HbA1c at diagnosis# (%)	6.9 <u>+</u> 1.5	10.2 <u>+</u> 2.3	<0.001
HbA1c at diagnosis (mmol/mol)	52 <u>+</u> 16	88 <u>+</u> 25	
Positive autoantibodies§, N (%)			1
0-1 Ab	14 (24.6)	13 (23.2)	
≥2 Ab	43 (75.4)	43 (76.8)	
AUC¶ C-Peptide at baseline§ (ng/mL)	1.7 ± 0.8	1.3 ± 0.6	0.007

Means ± standard deviations are shown unless specified otherwise

Weight, height and BMI were converted to SD units (Z scores) using CDC growth chart

- \* Wilcoxon rank-sum test for continuous variables and Fisher's exact test for proportions
- † FDR with T1D= first-degree relative with type 1 diabetes
- ‡ Eight subjects with missing information were omitted from the analysis
- # One subject with missing HbA1c was omitted from the analysis
- § data from baseline visit and if missing from 3-month visit
- || Ab= autoantibodies
- ¶ AUC= area under the curve

Table 2: Factors associated with slope of AUC C-peptide change (Univariate baseline analyses)

Factor	Estimated effect (95% CI) on the slope of AUC C-peptide (ng/mL/month)	P-value
C peptide AUC (per 1 unit log)	-0.011 (-0.021, -0.001)	0.037
Female sex	-0.006 (-0.011, -0.002)	0.010
Age at diagnosis (per year)	0.002 (0.000, 0.003)	0.008
Weight z-score at Dx (per SD)	-0.005 (-0.008, -0.002)	0.001
BMI z-score at Dx (per SD)*	-0.003 (-0.005, -0.001)	0.013
# of positive autoantibodies	-0.006 (-0.009, -0.003)	<0.001
IA-2A positive	-0.009 (-0.015, -0.003)	0.004
ZnT8A positive	-0.007 (-0.012, -0.002)	0.009

Analyses were adjusted for age at diagnosis (years) and C-peptide AUC (area under the curve) at baseline.

<sup>\*</sup> Eight subjects with missing BMI z-score were omitted from the analysis

Table 3: Factors associated with slope of AUC C-peptide change (Multivariate baseline analyses)

Factor	Estimated effect (95% CI) on the slope of AUC C-peptide (ng/mL/month)	P-value
Female	-0.006 (-0.011, -0.001)	0.014
Age at diagnosis (per year)	0.002 (0.001, 0.003)	0.005
Weight z-score at Dx (per SD)	-0.003 (-0.006, -0.001)	0.014
IA-2A positive	-0.006 (-0.012, 0.000)	0.061
ZnT8A positive	-0.005 (-0.010, 0.000)	0.065

Analyses were adjusted for age at diagnosis (years) and C-peptide AUC (area under the curve) at baseline.

Table 4: Factors associated with slope of AUC C-peptide change (Univariate 3 month visit analyses)

Factor	Estimated effect (95% CI) on the slope of AUC C-peptide (ng/mL/month)	P-value
C peptide AUC (per 1 unit log)	-0.011 (-0.021, -0.001)	0.037
Female	-0.006 (-0.011, -0.002)	0.010
Age at diagnosis (per year)	0.002 (0.000, 0.003)	0.008
Weight z-score (per SD)*	-0.006 (-0.009, -0.002)	0.001
BMI z-score (per SD)*	-0.005 (-0.008, -0.002)	0.002
# of positive autoantibodies †	-0.006 (-0.009, -0.003)	<0.001
IA-2A positive ‡	-0.010 (-0.016, -0.003)	0.002
ZnT8A positive ‡	-0.006 (-0.012, -0.001)	0.018
HbA1c (per %) §	-0.003 (-0.006, -0.001)	0.018

Analyses were adjusted for age at diagnosis (years) and C-peptide AUC (area under the curve) at baseline.

- \* Three subjects with missing information were omitted from the analysis
- † Six subjects with missing information were omitted from the analysis
- ‡ Four subjects with missing information were omitted from the analysis
- § Three subjects with missing HbA1c were omitted from the analysis

Table 5: Factors associated with slope of AUC C-peptide change (Multivariate 3 month visit analyses)

	Estimated effect (95% CI) on the slope of AUC	
Factor	C-peptide (ng/mL/month)	P-value
Female	-0.006 (-0.010, -0.001)	0.017
Age at diagnosis (per year)	0.002 (0.001, 0.003)	0.001
Weight z-score (per SD)	-0.005 (-0.008, -0.001)	0.009
HbA1c (per %)	-0.004 (-0.007, -0.001)	0.005
IA-2A positive	-0.006 (-0.012, 0.000)	0.051
ZnT8A positive	-0.005 (-0.010, 0.000)	0.052

Analyses were adjusted for age at diagnosis (years) and C-peptide AUC (area under the curve) at baseline.

Five subjects with missing information on HbA1c, IA-2A or ZnT8A status were omitted from the analysis.

# **Figure legends**

Figure 1: Individual trajectories of AUC C-peptide (log-transformed) over time in TEDDY children (A) and community controls (B)

AUC= area under the curve



