#### TURUN YLIOPISTON JULKAISUJA ANNALES UNIVERSITATIS TURKUENSIS

SARJA - SER. D OSA - TOM. 886 MEDICA - ODONTOLOGICA

# PROGNOSTIC MARKERS AND PROLONGED INTERFERON-α THERAPY IN RENAL CELL CARCINOMA

by

Minna Kankuri-Tammilehto

TURUN YLIOPISTO UNIVERSITY OF TURKU Turku 2009

#### From:

Departments of Oncology and Radiotherapy, and Pathology University of Turku Turku, Finland

#### **Supervisors:**

Professor Eeva Salminen, MD, PhD Department of Oncology and Radiotherapy University of Turku, Turku University Hospital Turku, Finland

Professor Seppo Pyrhönen, MD, PhD Department of Oncology and Radiotherapy University of Turku, Turku University Hospital Turku, Finland

Docent Tarja-Terttu Pelliniemi, MD, PhD Department of Clinical Chemistry University of Turku, Turku University Hospital Turku, Finland

#### **Reviewers:**

Docent Vesa Kataja, MD, PhD Department of Oncology University of Kuopio Vaasa Central Hospital Vaasa, Finland

Docent Anne Räisänen-Sokolowski, MD, PhD Department of Pathology University of Helsinki, Helsinki University Central Hospital Helsinki, Finland

#### **Dissertation Opponent:**

Professor Pirkko-Liisa Kellokumpu-Lehtinen, MD, PhD Department of Oncology and Radiotherapy University of Tampere, Tampere University Hospital Tampere, Finland

ISBN 978-951-29-4159-9 (PRINT) ISBN 978-951-29-4160-5 (PDF) ISSN 0355-9483 Painosalama Oy – Turku, Finland 2009



Now that almost anything seems technically possible, the key issue for the twenty-first century biologist is to identify the right questions to ask.

L. Franks and M. Knowles

#### **ABSTRACT**

Minna Kankuri-Tammilehto

Prognostic Markers and Prolonged Interferon-α Therapy in Renal Cell Carcinoma

Departments of Oncology and Radiotherapy, and Pathology University of Turku, Turku University Hospital Annales Universitatis Turkuensis Turku, Finland

The five-year survival for renal cell carcinoma (RCC) is  $\sim$ 50%. According to previous studies, for metastatic RCC (mRCC) with interferon- $\alpha$  (IFN- $\alpha$ ) based therapies the five-year survival is 3-16%. Typically, IFN- $\alpha$  treatment duration has been <6 months. Current questions in the use of IFN- $\alpha$  alone or with targeted therapies are the optimal dose and schedule.

The main aims were to investigate 1) the efficacy and tolerance of IFN- $\alpha$  with prolonged and intermittent administration in mRCC, and 2) the prognostic significance of p53, Ki-67 and COX-2 protein expressions in RCC. For 117 mRCC patients, IFN- $\alpha_{2a}$  (Roferon-A®) was planned to be continued at the maximal tolerated dose with a one-week pause each month up to 24 months or progression or intolerable toxicity. Protein expressions were analyzed immunohistochemically from nephrectomized and paraffin-embedded tissues from three different groups: primary metastases (n=29), later metastases (n=37), and no metastases (n=51).

Mean duration of IFN- $\alpha$  therapy was 11 months [0.5-32 months]. The rate of objective responses was 17%, stable disease 42%, and late responses (after 12 months' therapy) 3%. Progression-free survival and median overall survival were 8 and 19.1 months, respectively. Five-year survival was 16%; patients with lung metastases had higher five-year survival. No life-threatening side-effects were observed. Prolonged therapy (>12 months) is recommended for stable and responding patients.

Double positivity for p53 and Ki-67 expression indicates high metastases probability. Positive COX-2 expression indicates slow metastases development. For mRCC, positive p53 and Ki-67 expressions indicate poor prognosis, positive COX-2 expression indicates favorable prognosis. COX-2 positivity in Ki-67 negative tumors strongly indicates improved survival in mRCC.

**Key words**: COX-2, interferon-α, Ki-67, p53, renal cell carcinoma, response, long-term survival

### TIIVISTELMÄ

Minna Kankuri-Tammilehto Molekyylimarkkerit ja pitkäaikainen alfainterferonihoito munuaissyövässä

Onkologia ja sädehoito ja Patologia Turun Yliopisto, Turun Yliopistollinen Keskussairaala Annales Universitatis Turkuensis Turku, Finland

Munuaissyöpäpotilaiden viiden vuoden elossaololuku on noin 50 %. Aikaisempien tutkimuksien mukaan viiden vuoden elossaololuku metastasoituneessa munuaissyövässä on 3-16 %, kun käytettiin alfainterferonia sisältävää hoitoa. Tyypillisesti alfainterferonia on käytetty vähemmäin kuin 6 kuukautta. Avoimia kysymyksiä ovat alfainterferonin optimaalinen hoitoannos ja hoidon kesto yksin tai yhdessä uusien täsmähoitojen kanssa.

Tärkeimmät tavoitteet olivat tutkia 1) jaksotetun pitkäaikaisen alfainterferonihoidon tehoa ja siedettävyyttä metastasoituneessa munuaissyövässä ja 2) p53-, Ki-67- ja COX-2-proteiinituotannon ennusteellista merkitystä munuaissyövässä. Tutkimuksessa 117 metastasoituneelle munuaissyöpää sairastaneelle potilaalle etsittiin yksilöllinen hänen sietämänsä maksimaalinen hoitoannos rekombinanttia alfa<sub>2a</sub>-interferonia (Roferon-A<sup>®</sup>). Hoitoa pyrittiin jatkamaan 24 kuukauden ajan. Kolmen hoitoviikon jälkeen pidettiin yhden viikon tauko. Hoito lopetettiin, jos ilmaantui vakavia haittavaikutuksia tai tauti eteni. Toisessa tutkimuksessa proteiinituotanto analysoitiin immunohistokemiallisesti munuaissyöpäpotilaiden kasvainnäytteistä, joita oli säilytetty parafiinissa. Kasvainnäytteet oli otettu talteen munuaisen poistoleikkauksen yhteydessä. Nämä potilaat jaettiin kolmeen eri ryhmään: metastasointii primaarivaiheessa (n=29), metastasointi myöhemmin (n=37) ja ei metastasointia (n=51).

Keskimääräinen alfainterferonihoidon kesto oli 11 kuukautta (kk) [0,5 – 32 kk]. Objektiivinen hoitovaste todettiin 17 %:lla, tautitilanne pysyi ennallaan 42 %:lla ja myöhäinen vaste (yli 12 kk:tta hoidon aloittamisesta) todettiin 3 %:lla. Aika vasteen saavuttamisesta taudin etenemiseen oli keskimäärin 8 kk ja elinaika 19,1 kk. Viiden vuoden elossaololuku oli 16 %. Jos metastasoituneella munuaissyöpäpotilaalla oli keuhkometastasointi, hän selvisi todennäköisemmin viisi vuotta kuin muut potilaat. Henkeä uhkaavia sivuvaikutuksia ei todettu. Yli 12 kk:n ajan kestävä alfainterferonihoito on hyödyllistä niille potilaille, jotka ovat saaneet objektiivisen hoitovasteen tai tautitilanne on pysynyt ennallaan.

Positiivinen p53- ja Ki-67-ekspressio yhdessä viittaavat suureen metastasoinnin todennäköisyyteen. Positiivinen COX-2-ekspressio viittaa viivästyneeseen metastaasien ilmaantumiseen. Metastasoituneilla potilailla positiiviset p53- ja Ki-67-ekspressiot viittaavat huonoon ennusteeseen, mutta positiivinen COX-2 ekspressio viittaa suotuisaan ennusteeseen. Positiivinen COX-2- ja negatiivinen Ki-67-ekspressio yhdessä viittaavat parantuneeseen ennusteeseen metastasoituneessa munuaissyövässä.

Avainsanat: alfainterferoni, COX-2, elinaika, Ki-67, munuaissyöpä, p53, vaste

# **TABLE OF CONTENTS**

٨R	STRA	CT		5
			VTENTS	
			NS	
			NAL PUBLICATIONS	
1			TION	
2			F THE LITERATURE	
	2.1		CAL BEHAVIOR OF RENAL CELL CARCINOMA (RCC)	
		2.1.1	EPIDEMIOLOGY	
		2.1.2 2.1.3	ETIOLOGY AND RISK FACTORSSIGNS AND SYMPTOMS	
		2.1.3	NATURAL COURSE	
		2.1.4	IMAGING	
	2.2		ING AND PROGNOSTIC FACTORS IN RCC	21
	2.2	2.2.1	WHO AND HEIDELBERG CLASSIFICATIONS FOR TYPING OF RENAL TUMORS	
		2.2.2	TNM CLASSIFICATION SYSTEM FOR PATHOLOGICAL TUMOR STAGING	
		2.2.3	WHO CLASSIFICATION FOR HISTOPATHOLOGICAL TUMOR GRADING	
		2.2.4	PROGNOSTIC MODELS	30
			2.2.4.1 PROGNOSTIC MODELS IN LOCALIZED DISEASE	
			2.2.4.2 PROGNOSTIC MODELS IN METASTASIZED DISEASE	
	2.3		ARKERS RELATED TO MOLECULAR MECHANISMS IN RCC	
		2.3.1	pVHL, VON HIPPEL-LINDAU PROTEIN, MODULATOR OF HYPOXIC RESPONSE	
		2.3.2	CA9, HYPOXIA ASSOCIATED ENZYME	
		2.3.3	p53, BIOMARKER OF CELL CYCLE POINT	
		2.3.4 2.3.5	COX-2, BIOMARKER FOR INFLAMMATION AND NEOPLASIA	
		2.3.6	Her-2, BIOMARKER OF PROTO-ONCOGENE PRODUCT	
	2.4		APIES FOR RCC	
		2.4.1	SURGERY	
			2.4.1.1 NEPHRECTOMY FOR LOCALIZED DISEASE	
			2.4.1.2 NEPHRON-SPARING SURGERY	40
			2.4.1.3 CYTOREDECTIVE NEPHRECTOMY	
			2.4.1.4 METASTASECTOMY	
		2.4.2	RADIOTHERAPY	
		2.4.3	CHEMOTHERAPY	
		2.4.4	INTERFERON-α THERAPY	
			2.4.4.1 RESPONSE TO IFN-α	
			2.4.4.2 ANTI-IFN ANTIBODIES 2.4.4.3 DOSAGE AND SCHEDULE	
			2.4.4.4 QUALITY OF LIFE (QoL)	
			2.4.4.5 NEPHRECTOMY PRIOR TO IFN-α	
			2.4.4.6 IFN-α VERSUS OTHER THERAPIES	47
			2.4.4.7 IFN-α PLUS OTHER THERAPIES	
			2.4.4.8 IFN-α AS SECOND LINE THERAPY	
			2.4.4.9 IFN-α AS ADJUVANT THERAPY	
			2.4.4.10 BIOLOGICAL EFFECTS OF IFN-α	
		2.4.5	INTERLEUKIN-2	
		2.4.6	TARGETED THERAPIES	
		2.4.7	OTHER THERAPIES	

3	AIM	S OF THE PRESENT STUDY	53
4	MA	FERIALS AND METHODS	54
	4.1	EFFICACY AND TOLERABILITY OF PROLONGED ADMINISTRATION	
		OF IFN-α <sub>2a</sub> IN mRCC IN PHASE II CLINICAL STUDIES. (I, II)	56
		4.1.1 PATIENT ELIGIBILITY	56
		4.1.2 TREATMENT SCHEDULE	56
		4.1.3 STAGING AND RESPONSE ASSESSMENT	57
		4.1.4 STATISTICS	57
	4.2	CHANGES IN BLOOD NEUTROPHIL AND MONOCYTE RECEPTOR PROFILE	
		DURING IFN-α <sub>2A</sub> ADMINISTRATION IN mRCC (III)	
		4.2.1 REAGENTS	
		4.2.2 COLLECTION AND PREPARATION OF SAMPLES	
		4.2.3 MEASUREMENTS OF RECEPTOR EXPRESSION	
		4.2.4 IN VITRO EXPERIMENTS	
		4.2.5 STATISTICS	59
	4.3	PROGNOSTIC SIGNIFICANCE OF MOLECULAR MARKERS OF p53, Ki-67	
		AND COX-2 IN RCC ACCORDING TO OCCURRENCE OF METASTASES (IV, V)	
		4.3.1 PATIENTS, STAGING, AND HISTOLOGY	
		4.3.2 IMMUNOHISTOCHEMICAL STAINING AND SCORING OF p53, Ki-67 AND CO	
		4.3.3 STATISTICAL ANALYSIS	
5		ULTS	62
	5.1	EFFICACY AND TOLERANCE OF PROLONGED ADMINISTRATION OF IFN- $\alpha_{2a}$	
		IN mRCC IN PHASE II CLINICAL STUDIES. (I, II)	62
	5.2	CHANGES IN BLOOD NEUTROPHIL AND MONOCYTE RECEPTOR PROFILE	
		DURING IFN-α <sub>2A</sub> ADMINISTRATION IN MRCC (III)	63
	5.3	PROGNOSTIC SIGNIFICANCE OF MOLECULAR MARKERS OF p53, Ki-67	
		AND COX-2 IN RCC ACCORDING TO OCCURRENCE OF METASTASES (IV, V)	
6		CUSSION	
	6.1	IFN-α IN mRCC	70
		6.1.1 RESPONSE TO AND LONG-TERM SURVIVAL WITH PROLONGED USE	
		OF IFN-α THERAPY	
		6.1.2 CLINICOPATHOLOGICAL PROGNOSTIC FACTORS IN MRCC PATIENTS	
		6.1.3 TOXICITY OF IFN-α COMPARED TO OTHER THERAPIES	
		6.1.4 TIMING OF IFN-α IN THE ERA OF NOVEL TARGETED THERAPIES	
	6.2	BIOLOGICAL EFFECTS OF IFN-α	77
	6.3	CLASSIFICATIONS ACCORDING TO MORPHOLOGY AND GENETIC FINDINGS	
		IN RCC	
	6.4	T-STAGE AND GRADE AS PROGNOSTIC FACTORS IN RCC	
	6.5	BIOMARKERS AS PROGNOSTIC FACTORS IN RCC	80
		6.5.1 BIOMARKERS IN RELATION TO T-STAGE, GRADE OR OCCURRENCE	
		OF METASTASES	80
		6.5.2 BIOMARKER ASSOCIATION WITH SURVIVAL	82
		6.5.3 INCIDENCE OF p53, Ki-67, AND COX-2 EXPRESSIONS	
		6.5.4 COMBINING MARKERS	
_		6.5.5 TRENDS IN THE USE OF BIOMARKERS	
7		IMARY AND CONCLUSIONS	
AC	KNO	WLEDGEMENTS	88
RE	FERI	ENCES	90
ΔD	ICIN	AL DUDI ICATIONS	115

# **ABBREVIATIONS**

A 12CD A	12 ais matinaid said	HIE 1	homorio inducible featon 1 alaba
A13CRA	13-cis-retinoid acid	HIF-1α HLRCC	hypoxia inducible factor - 1 alpha hereditary leiomyomatosis and RCC
ACKD ADCC	acquired cystic kidney disease antibody-dependent cell-mediated	HPT-JT	, ,
ADCC	cytotoxicity	HR	hyperparathyroidism-jaw tumor hazard ratio
AJCC	the American Joint Committee on Cancer		
ASR	age- standardized incidence rate	HRPT2	hyperparathyroidism 2 gene
BHD	Birt-Hogg-Dubé syndrome	IFN	interferon
BHD	Birt-Hogg-Dubé gene	IFN-α	leucocyte interferon-alpha
BMI	body mass index	IFN-β	fibroblast interferon-beta
CA9	carbonic anhydrase 9	IFN-γ	immune interferon-gamma
c-erbB2	Her-2	IGF-I	insulin-like growth factor-I
CI	confidence interval	IL	interleukin
CISH	chromogenic in situ hybridzation	IL-2	interleukin-2
c-met	see MET	IVC	inferior vena cava
CNS	central nervous system	Ki-67	proliferation biomarker
COR	cumulative odds ratios	LD-IL-2	low-dose interleukin-2
COX-2	cyclooxygenase 2	lm	later metastases (after nephrectomy),
CR CR	complete response	LOH	metachronous metastases loss of heterozygosity
CR1	complement receptor 1	mAb	** *
CR3	•	MET	monoclonal antibody hepatocyte growth factor receptor
CRP	complement receptor 3		1 , 0
	c-reactive protein	MET	MET proto-oncogene
CT	computerized tomography	MHC	major histocompatibility complex
DNA	deoxyribonucleic acid	MNV	median nuclear volume
DSS	disease-specific survival	mos	months
EAU	European Association of Urology	MPA	medroxyprogesterone acetate
ECOG PS	the Eastern Cooperative Oncology Group	mRCC	metastatic renal cell cancer
ECOG-PS	ECOG performance status	MRI	magnetic resonance imaging
EGFR	epidermal growth factor receptor	mRNA	messenger ribonucleic acid
EPO	erythropoietin	MSKCC	Memorial Sloan Kettering Cancer center
ESMO	European Society for Medical Oncology	mTOR	mammalian target of rapamycin
EU	the European Union	MU	million units
FACT-G	the functional assessment of cancer	NCCN	National Comprehensive Cancer Network
FCR	therapy general scale Finnish cancer registry	NK	natural killer cell
FcγRI	phagocyte receptor expression I	nm	no metastases during 7.5 years' follow-up
FcγRII	phagocyte receptor expression II	NSS	nephron-sparing surgery
•		OR	odds ratios
FcγRIII <i>FH</i>	phagocyte receptor expression III	OS	overall survival
	fumarate hydratase gene	p53	protein p53, biomarker of cell cycle
FHIT	fragile histidine triad gene	D.C.D.	check point
FISH	fluorescent in situ hybridization	PCR	polymerase chain reaction
FKSI	functional assessment of cancer therapy-	PD	progressive disease
FLCN	kidney symptom index folliculin gene	PEG-IFN-α	pegylated interferon-α
FPTC-PRN	familial papillary thyroid carcinoma-	PET	positron emission tomography
1110110.	papillary renal neoplasia	pm	primary metastases (at nephrectomy), synchronous metastases
G250	carbonic anhydrase IX monoclonal	PN	partial nephrectomy
Cv	antibody	PR	partial response
Gy	Gray	PRCC	papillary renal cell carcinoma
HD-IL-2	High dose interleukin-2	pTNM	pathological TNM
Her-2	Human Epidermal growth factor Receptor 2	pVHL	von Hippel-Lindau protein
Her-2	Her-2 proto-oncogene	QoL	Quality of Life
HIF	hypoxia-inducible factor	-	
	>F	RCA	renal cell adenoma

RCC renal cell carcinoma

RhuMAb recombinant humanized monoclonal

antibody

RNA ribonucleic acid

SD

RPLND that retroperitoneal lymph node

dissection stable disease

TCS2 tuberous sclerosis 2 gene tiw three times a week

TNF tumor necrosis factor

TNM T=tumor, N=node, M=metastases

TP53 Tumor protein p53 gene

TSC1 tuberous sclerosis 1 gene

UCLA University of California

UICC Union Internationale Contre le Cancer (The International Union Against Cancer)

US Ultrasonography

VEGF vascular endothelial growth factor VEGFR-TKI vascular endothelial growth factor

receptor tyrosine kinases

VHL von Hippel-Lindau disease

VHL von Hippel-Lindau gene

WHO World Health Organization

YCC Yale Comprehensive Cancer Center

#### LIST OF ORIGINAL PUBLICATIONS

This thesis is based on the following original publications referred to in the text by their Roman numerals (I-VI).

I

**Kankuri M**, Pelliniemi T-T, Pyrhönen S, Nikkanen V, Helenius H, Salminen E (2001): Feasibility of Prolonged Use of Interferon- $\alpha$  in Metastatic Kidney Carcinoma. A Phase II Study. Cancer 92 (4): 761-767.

П

**Kankuri-Tammilehto M**, Söderström K-O, Pelliniemi T-T, Hinkka-Yli-Salomäki S, Pyrhönen S, Salminen E (2009): Long-Term Outcome of Metastatic Renal Cell Carcinoma with Prolonged Use of Interferon-α Administered Intermittently. A Phase II Study. Submitted.

Ш

Salminen E, **Kankuri M**, Nuutila J, Lilius E-M, Pelliniemi T-T (2001): Modulation of IgG and Complement Receptor Expression of Phagocytes in Kidney Cancer Patients during Treatment with Inteferon-α. Anticancer Research 21 (3B): 2049-2055.

IV

**Kankuri M**, Söderström K-O, Pelliniemi T-T, Vahlberg T, Pyrhönen S, Salminen E (2006): The Association of Immunoreactive p53 and Ki-67 with T-stage, Grade, Occurrence of Metastases and Survival in Renal Cell Carcinoma. Anticancer Res 26 (5B): 3825-3833.

V

**Kankuri-Tammilehto M**, Söderström K-O, Pelliniemi T-T, Vahlberg T, Pyrhönen S, Salminen E (2009): Prognostic Evaluation of COX-2 Expression in Renal Cell Carcinoma. Submitted.

VI

**Kankuri-Tammilehto M.** Chapter IV - Association of p53, Ki-67, COX-2, and Her-2 Expressions, as well as T-stage and Histopathological Grade, with Occurrence of Metastases and Survival in Renal Cell Carcinoma. Review. In: Watanabe A, ed. Cancer Metastases Research. New York: Nova Science Publishers, Inc., 2008, pp.109-132.

The original articles are reproduced with permission of the copyright holders.

#### 1 INTRODUCTION

Renal cell carcinomas (RCCs) are renal epithelial neoplasms in renal parenchyma, also called renal adenocarcinomas. The majority of RCCs arise from the cells of renal proximal tubules of nephrons, but 5% of cases arise from the cells of the collecting ducts (Chao *et al.* 2002, Kovacs *et al.* 1997, Störkel *et al.* 1997). RCCs are typically highly vascularized solid tumors (Kovacs *et al.* 1997), and they have the ability to spread via multiple routes, such as directly into the perinephric tissues, or hematogeneously through the renal vein and inferior vena cava and lymphatic tissues. Renal tumors are members of a complex family with unique histology, cytogenetic defects and variable metastatic potential (Linehan *et al.* 2003, Thoenes *et al.* 1986). Of all RCCs, 70-80% are of conventional type, also known as clear cell RCCs. Of these, approximately 75% have a mutation in the von Hippel-Lindau tumor suppressor gene (*VHL*) in the short arm of chromosome 3 (Maxwell *et al.* 1999, Gnarra *et al.* 1994).

Macroscopic hematuria, pain and palpable tumor, together called the classic triad, indicate metastatic disease (Cunningham 1938). Nowadays, 30-60% of RCC tumors are found incidentally in abdominal imaging performed for some other reason than suspected renal tumor, such as the evaluation of non-specific abdominal or musculoskeletal complaints (Jayson and Sanders 1998).

The expected five-year survival rate for all RCC stages is approximately 50%. For those patients with performance status enabling current treatments the rate is slightly higher than 60% (Parkin *et al.* 2003). According to a few previous studies on long-term outcome for metastatic RCC (mRCC), the five-year survival is from 3% to 16% (Atzpodien *et al.* 2002, Motzer *et al.* 2000a, Minasian *et al.* 1993) if metastasectomy is not a possible treatment. For localized RCC, nephrectomy is the only curative treatment (Robson *et al.* 2002), and there is no effective adjuvant therapy. Possible treatments for mRCC, in addition to cytoreductive nephrectomy (Flanigan *et al.* 2001, Mickisch *et al.* 2001), are immunomodulators, such as interferon-α (IFN-α) (Pyrhönen *et al.* 1999), interleukin-2 (IL-2) (Négrier *et al.* 2007, Spanknebel *et al.* 2005), and more recently tyrosine kinase inhibitors, such as sunitinib (Motzer *et al.* 2007), sorafenib (Escudier *et al.* 2007a), and mTOR inhibitor temsirolimus (Hudes *et al.* 2007). Everolimus, another mTOR inhibitor, has an encouraging antitumor activity against mRCC (Motzer 2008a). The efficacy of bevacizumab, an antiangiogenesis monoclonal antibody, has also been shown (Yang *et al.* 2003a). Interferon-alpha (IFN-α) has been considered the standard comparator when investigating novel targeted therapies in mRCC.

Compared to other biological response modifiers, chemotherapy and hormonal therapy, IFN- $\alpha$  is associated with survival benefits (MRCRCC *et al.* 1999, Pyrhönen *et al.* 1999). High dose

interleukin-2 therapy (HD-IL-2) may improve survival in patients with the poorest prognosis (McDermott *et al.* 2005). The current questions concerning the use of immunomodulators, such as IFN-α, are timing (Motzer *et al.* 2007, Stadler *et al.* 2007) and optimal dosage as monotherapy or in combination with targeted therapies (Bracarda *et al.* 2007).

RCC is a heterogeneous disease: the prognosis differs at the same stage and grade (Tsui *et al.* 2000a). Currently, T-stage is the best known prognostic factor for locally confined RCC. The most often represented factors in mRCC are performance status, time to metastases, number of metastatic sites, and prior nephrectomy. All the molecular mechanisms that affect the development, progression and clinical behavior of RCC are not known. A better understanding of these and the factors affecting the response to IFN- $\alpha$  enables us to target treatment more selectively in mRCC. Molecular biomarkers, such as p53, Ki-67 and COX-2, are potential candidates for staging, assessing prognosis, and guiding targeted therapies (Masters 2007).

#### 2 REVIEW OF THE LITERATURE

#### 2.1 CLINICAL BEHAVIOR OF RENAL CELL CARCINOMA (RCC)

#### 2.1.1 EPIDEMIOLOGY

Kidney cancer\* represents 2-3% of all diagnosed malignancies worldwide (Parkin *et al.* 2003). In some Northern and Central European countries the incidence is higher, e.g. in Iceland 4.1%, and in the Czech Republic 5.4% (Parkin *et al.* 2003). In Finland, kidney cancer represents 2-3% of all malignancies, and the age-standardized incidence rate (ASR) is 8.8/100 000 for men and 5.4/100 000 for women (Finnish Cancer Registry (FCR) 2007). Kidney cancer is more common among urban than among rural residents. In the European Union, the estimated annual number of new kidney cancers is 46 000 (Ferlay *et al.* 2001). In Finland, 735 new kidney cancers were diagnosed in 2007, and the number of kidney cancer patients alive on the 1<sup>st</sup> of January 2008 was 6 017. In 2007, altogether 346 patients succumbed to kidney cancer in Finland (FCR 2007).

Since the 1970s, the annual increase in RCC incidence has been 2-4% (Mathew *et al.* 2002). This has been attributed to the use of radiological imaging which is able to find presymptomatic RCC lesions (Jayson and Sanders 1998), as well as the increased prevalence of etiologic risk factors, such as cigarette smoking (Hunt *et al.* 2005) and obesity (Chow *et al.* 2000, Yuan *et al.* 1998). The rise in incidence has been greater in women than in men (Chow *et al.* 2000). The increase has been highest in localized disease, especially in tumors that are less than 4 cm in diameter (Hollingsworth *et al.* 2006). In Scandinavia, a decrease in incidence has been observed since the 1980's in Sweden and Denmark, and since the 1990's in Finland, in contrast to Norway (ANCR 2009). The reason for this phenomenon is unclear, but one explanation may be variations in cigarette smoking (Mathew *et al.* 2002) and tobacco legislation. In Danish men the incidence started to increase again in the 2000's.

Of kidney cancer cases, 84% are diagnosed between 50 and 84 years of age (FCR 2007, Parkin *et al.* 2003), and the median age of diagnosis is around 64 years for Caucasians and 58 years for African-Americans (FCR 2007, Parkin *et al.* 2003). RCC has been observed in as young as sixmonth-old children, possibly as a hereditary disease (Sanchez-Ortiz *et al.* 2004, Renshaw *et al.* 

<sup>\*</sup> In epidemiological statistics, RCC and renal pelvis cancer are usually not reported separately, but combined under the heading of kidney cancer. Eighty per cent of adult kidney tumors are RCCs; transitional-cell carcinomas of the renal pelvis account for approximately 15%, and the remaining (< 5%) kidney tumors include a variety of rare lesions such as embryonic renal neoplasms, renal sarcomas and tumors of primarily mesenchymal origin (Parkin *et al.* 2003).

1999). Young patients are more likely to have unfavorable histopathological features and lymph node metastases than older patients (Sanchez-Ortiz *et al.* 2004, Renshaw *et al.* 1999).

A male-to-female ratio in RCC incidence is from 1.3:1 to 2.5:1 (FCR 2007, Motzer *et al.* 1999). At primary diagnosis, men more often than women have stage III-IV disease (Thrasher and Paulson 1993).

In the USA, mortality caused by kidney cancer rose 1.3% annually from 1975 to 1992 (annual percentage change), but decreased by 0.5% annually from 1992 to 2002 (Edwards *et al.* 2005). In the EU, the fall in kidney cancer mortality between 1994 and 1999 was over 10%, being most marked in those Northern and Central European countries where mortality from kidney cancer had been observed as very high compared to other European countries (Levi *et al.* 2004). It is estimated that kidney cancer is responsible for 102,000 deaths per year worldwide (Parkin *et al.* 2003). In Finland, 345 patients die annually of RCC (FCR 2007)

#### 2.1.2 ETIOLOGY AND RISK FACTORS

The etiology of RCC is still largely undefined. The most consistently established causal risk factors for RCC are cigarette smoking and obesity, in addition to acquired cystic kidney disease and inherited susceptibility (Table 1), but the significance of many other potential risk factors is being investigated.

Cigarette smoking has been found to be a definitive risk factor for RCC (Hunt et al. 2005, McLaughlin et al. 1984). The strong dose-response relationship of cigarette smoking and RCC and the decrease in risk of RCC following cessation of cigarette smoking (15-30% after 10 to 15 years since cessation) (Hunt et al. 2005, McLaughlin et al. 1984) support a causal interpretation of the association. The relative risk is 1.54 for male and 1.22 for female smokers (Hunt et al. 2005). It has been estimated that up to 30% of RCC in men, and up to 20% in women may be directly due to cigarette smoking (McLaughlin et al. 1984).

The carcinogenic agent in cigarette smoke has not been clearly identified (Hunt *et al.* 2005, McLaughlin *et al.* 1984). Most of the constituents in cigarette smoke are metabolized or excreted through the urinary tract. Nitrous compounds, especially N-nitrosodimethylamine, found in cigarette smoke, has been observed to cause epithelial renal tumors in animal models (IARCC 2004), and *VHL* gene mutation has been associated with N-nitrosodimethylamine (Shiao *et al.* 1998). Metabolic gene polymorphisms have been suggested as risk factors for RCC. These polymorphisms are involved in the activation or detoxification of carcinogens in cigarette smoke.

Table 1. Risk factors of RCC.

Risk factors for RCC	Evidence of risk factor	References
Self-inflicted		
Cigarette smoking	Well-established	Hunt et al. 2005, McLaughlin et al. 1984
Obesity	Well-established	Samanic <i>et al.</i> 2006,
Low recreational activity	Potential	Bergström et al. 2001, Chow et al. 2000 Menezes et al. 2003
Illness		
Acquired cystic kidney disease	Well-established	Schwarz <i>et al.</i> 2007, Maisonneuve <i>et al.</i> 1999
Hypertension	Highly potential	Chow <i>et al.</i> 2000, Shapiro <i>et al.</i> 1999, Yuan <i>et al.</i> 1998, McLaughlin <i>et al.</i> 1995
Diabetes mellitus	Potential	Washio et al. 2007, Lindblad et al. 1999
Medication		
Diuretics	Potential	Yuan <i>et al.</i> 1998, McLaughlin <i>et al.</i> 1995
Oral contraceptives	Potential	Lindblad <i>et al.</i> 1995
Reproductive hormone factors		
Prior hysterectomy and	Potential	Lindblad et al. 1995
oophorectomy		
Occupational		
Thrichloroethene	Highly potential	Brüning et al. 2003, Brüning et al. 1997
Asbestos	Potential	Sali and Boffetta 2000
Sedentary work	Potential	Bergström et al. 1999
Arsenic	Potential	Marshall et al. 2007
Cadmium	Potential	Pesch et al. 2000
Nutrition		
Fried meats	Potential	Yuan et al. 1998
Low consumption of orange or green vegetables	Potential	Yuan et al. 1998
Low intake of D-vitamin	Potential	Karami et al. 2008
Low intake of E-vitamin	Potential	Zhang <i>et al.</i> 1997
Low intake of magnesium	Potential	Wolk <i>et al.</i> 1996
Hereditary	Well-established	Table 2

*Obesity*, increased body mass index (BMI), has been established as a risk factor for RCC in both men and women (Bergström *et al.* 2001). It has been estimated that more than 30% of RCC may be due to obesity (Bergström *et al.* 2001). The mechanisms by which obesity influences renal carcinogenesis are unclear, although the risk is suggested to be mediated via, e.g. sex steroid hormones and increased levels of insulin-like growth factor-I (IGF-I) (Bergström *et al.* 2001, Chow *et al.* 2000).

Hypertension seems to be an independent risk factor for RCC (Yuan *et al.* 1998), with the association being stronger for women (Chow *et al.* 2000, Shapiro *et al.* 1999). However, it has been difficult to distinguish the effect of therapy from its indication (McLaughlin *et al.* 1995).

Acquired cystic kidney disease (ACKD) causes an increased risk of RCC (Maisonneuve et al. 1999). ACKD occurs in end-stage renal disease and may develop in 20% of long-term dialysis patients (Schwarz et al. 2007, Maisonneuve et al. 1999), or due to kidney transplantation (Schwarz et al. 2007). The incidence of RCC in patients with end-stage renal disease is approximately 40 to 100 times (Maisonneuve et al. 1999), and in chronic renal failure, nine times higher than in the general population (Takahashi et al. 1993).

Thrichloroethene is a carcinogen that causes renal adenomas and carcinomas in animal models (Mensing et al. 2002), and its toxic metabolites have been associated with the VHL gene mutation (Brüning et al. 1997). Thrichloroethene exposure has been found in the urine of workers at a cardboard manufacturing plant. An association between a long period of high exposure to thrichloroethene and RCC was observed (Brüning et al. 2003).

Hereditary RCC accounts for less than 4% of all RCCs (Eble et al. 2004) (Table 2). Hereditary RCC is often characterized by an early age at onset (approximately 45 years), and frequently the bilaterality and multicentricity of the primary tumor (Gnarra et al. 1994), although, e.g. in HLRCC (Hereditary leiomyomatosis and RCC) and familial non von Hippel-Lindau (VHL) RCC syndromes, primary tumors are typically unilateral and solitary. The aggressiveness of hereditary RCCs varies depending on the syndrome and mutation type. If conventional RCC is of VHL disease origin, tumors of less than 3 cm have a low rate of metastatic potential and minimal invasive surgery is recommended (Zbar et al. 1999). However, if HLRCC is detected, radical nephrectomy is recommended due to the aggressiveness of RCC in this syndrome (Lehtonen et al. 2006). Li-Fraumeni syndrome, a rare autosomal dominant disorder, causes typically breast cancer, sarcoma, and leukemia, but in rare cases also RCC. In 70% of Li-Fraumeni syndrome germline mutations have been found in the TP53 tumor suppressor gene on 17p13.1 (Ruijs et al. 2006). Identification of family members at risk of hereditary RCC is important for organizing follow-up and surveillance programs with regular imaging of the kidneys. The identification of the genes that are responsible for familial renal tumors will result in better understanding of renal tumorigenesis, even in non-hereditary renal tumors.

Table 2. Hereditary syndromes that predispose to RCC.

	von Hippel-Lindau (VHL)	Hereditary PRCC	Hereditary leiomyomatosis and RCC (HLRCC)	Birt-Hogg- Dubé (BHD)	Tuberous	Constitutional chromosome 3 translocation	Familial non- syndromic RCC	Familial papillary thyroid carcinoma- papillary renal neoplasia (FPTC-PRN)	Hyperpara- thyroidism-jaw tumor (HPT-,TT)
RCC subtype	Conventional	Papillary type 1	Papillary type 2, collecting duct	Conventional, papillary, chromophobe	Conventional	Conventional	Conventional	Papillary type 1	Papillary type 1
Risk for RCC	%08-02		20-30%	15-40%	1-3%				
Gene	VHL (3p25)	MET	FH	FLCN (BHD)	TSCI, TSC2	FHIT, unknown	Unknown	Unknown	HRPT2
Chromosome	3p25-26	7q31-34	1q42-q44	e.g. 17p12- q11.2	9q34, 16p13.3	3p14, 3q13.3, 3q21	Unknown	1q21	1q21-q32
Characteristic of gene	Tumor suppressor gene	Proto- oncogene	Tumor suppressor gene	Tumor suppressor gene?	Tumor suppressor genes		Unknown		Tumor suppressor gene
Type of hereditary	Autosomal dominant	Autosomal dominant	Autosomal dominant	Autosomal dominant	Autosomal dominant	Initially balanced translocation between chromosome 3 and chromosome 6 or 8, then somatic lost of chromosome 3 and VHL mutations	Mostly autosomal dominant, also autosomal recessive		
Incidence of syndrome	1:36 000	1:10000000			1:6000	Some known families	Some known families		
Other signs of the syndrome than RCC	Retinal haemangioma, cerebellar or spinal haemangioblastoma, pheochromocytoma, renal or pancreatic cyst, endocrine pancreatic tumor	Breast cancer, pancreatic cancer, biliary tract cancer, lung cancer, lung cancer, malignant melanoma	Cutaneous and uterine leiomyomatosis, leiomyosarcoma, breast cancer, prostate cancer, bladder cancer	Cutaneous hair follicle bening tumor, pulmonary cyst, pneumothorax	Hamartoma, fibroma, renal angiomyolipo ma, rhabdomyoma, hypopigmentat ion, epilepsy, mental retardation	Thyroid cancer, bladder cancer, pancreatic cancer, gastric cancer		Papillary thyroid carcinoma, papillary renal adenoma, renal oncocytoma	Parathyroid adenoma/carcino ma, ossifying jaw tumor, renal cyst, renal hamartoma, adult Wilms
References	Zbar <i>et al.</i> 1999, Zbar <i>et al.</i> 1996	Schmidt et al. 1997	Koski <i>et al.</i> 2009, Lehtonen <i>et al.</i> 2006	Schmidt et al. 2001	Lendvay and Marshall 2003	Bonné <i>et al.</i> 2004, van Kessel <i>et al.</i> 1999	Woodward et al. 2008, Hemminki and Li 2004	Malchoff et al. 2000	Haven <i>et al.</i> 2000

#### 2.1.3 SIGNS AND SYMPTOMS

Due to the location of the kidney deep in the retroperitoneum, RCC may progress unnoticed for a long period: the early stage of RCC is often asymptomatic. Symptoms from the tumor are noticeable when it invades adjacent structures or the kidney's collecting system.

Typical signs of RCC are microscopic or macroscopic hematuria (40-60% of patients), pain (40%) and palpable tumor (30%). The phenomenon of macroscopic hematuria, flank pain and palpable abdominal mass existing at the same time is called *the classic triad*. The classic triad is observed in mRCC (Cunningham 1938), and nowadays it occurs in 10% of patients (Jayson and Sanders 1998). *An inflammatory syndrome* with fever, anemia, erythrocytosis, thrombocytosis, increased serum acute-phase protein levels and hypoalbuminemia (Boxer *et al.* 1978) is frequently associated with RCC. Approximately one-third of RCC patients develop a *paraneoplastic syndrome*, which is caused by a polypeptide hormone production of the RCC or normal cells, or antibody produced in response to the tumor. The signs of paraneoplastic syndrome are, e.g. cachexia, hypertension, humoral hypercalcemia, hyperglycemia, increased erythropoietin (EPO), anemia, c-reactive protein (CRP) elevation and erythrocytosis (Kim *et al.* 2003, Blay *et al.* 1997, Gucalp 1992). Stauffer's syndrome, i.e. paraneoplastic cholestasis, (3-20%) is a non-metastatic hepatic dysfunction causing hepatosplenomegaly, fever, fatigue and weight loss (Blay *et al.* 1997). The etiology of Stauffer's syndrome is unknown. In addition, glomerulonephropathias (30%) and amyloidosis (4%) are observed in RCC patients (Rosenblum 1987).

#### 2.1.4 NATURAL COURSE

Metastatic disease is seen in 20-30% of RCC patients at diagnosis (Janzen et al. 2003, Mc Nichols et al. 1981). Half of the patients diagnosed with local RCC will later have a recurrence of their cancer: two thirds within the first year (Janzen et al. 2003), and the majority within five years (Lam et al. 2005, McNichols et al. 1981). The risk for late recurrence, at over 10 years from nephrectomy, is at least 10% (McNichols et al. 1981). Typical sites of metastases in decreasing order of frequency are lungs, lymph nodes, bone, liver, adrenal gland, opposite kidney, and brain (Saitoh et al. 1982b), but metastases can be found in almost any organ, e.g. in thyroid gland, ureter and pancreas (Saitoh et al. 1982a).

Spontaneous regression has been suggested in approximately 1% of RCC patients, although some studies have reported higher percentages, from 4.4% to 6.6%, for selected patient groups (Gleave *et al.* 1998, Marcus *et al.* 1993). In a Cochrane meta-analysis, complete spontaneous remissions were observed in 3.3% of the assessed population (Coppin *et al.* 2007). Spontaneous regression has been

seen in both nephrectomized and in non-nephrectomized patients. It has most often occurred in RCC with pulmonary metastases, but also with extrapulmonary lesions, such as bone, liver, and central nervous system (CNS) lesions (Gleave *et al.* 1998, Lokich 1997). Additionally, non-progression of RCC at 12 months after nephrectomy has been observed in approximately 7% of patients (Oliver *et al.* 1989). Spontaneous regression has also been verified by biopsy-proven histological and radiological imaging of regressed lesions (Christophersen *et al.* 2006, Nakajima *et al.* 2006).

RCC is able to manipulate and suppress the body's natural immunity. The phenomenon of spontaneous regression reflects the ability of the host immune system to control the RCC tumor by inducing an immune response and enhancing antitumor immunity against the tumor. In previously untreated patients, lymphocytic infiltration in RCC tumors has been found. It is speculated that the rich supply of macrophages, lymphocytes, and immunoglobulin in the lung might suppress the metastases through host immune mechanisms (Freed *et al.* 1977). However, no enhancement of natural killer (NK) cells, lymphokine-activated killer cells or lymphocyte proliferative response has been observed (Abubakr *et al.* 1994), and the exact mechanisms are unclear.

#### 2.1.5 IMAGING

Ultrasonography (US), computerized tomography (CT) and nuclear magnetic resonance imaging (MRI) are in routine clinical use for imaging renal tumors. The ability of ultrasonography to differentiate renal masses from renal cysts was realised in the 1970's. US has become the primary examination in patients with acute flank pain and hematuria, and to differentiate cystic from solid renal mass, but it is not reliable enough to screen multiple cysts in hereditary RCC (Tosaka *et al.* 1990). In the 1980's, CT was shown to accurately image renal masses and cysts (Gillenwater and Howards 1981), and proved to be a cost-effective and a less invasive technique than angiography. Nowadays, CT is the most sensitive imaging method for RCC. MRI became popular during the 1990's, and many advantages of MRI compared to CT have been observed. MRI has a multiplanar imaging capability, particularly in studying the vasculature and lymph node metastases, and it may detect subcentimeter renal lesions (Ergen *et al.* 2004, Hricak *et al.* 1988). Radiation exposure is avoided by using MRI, and it is a safe method for patients with renal insufficiency (Rofsky *et al.* 1991). Bone scintigraphy detects bone metastases about six months earlier than conventional x-rays (Ghanema *et al.* 2005).

Novel potential imaging techniques, such as positron emission tomography (PET) with iodine-124-labeled antibody chimeric G250 (Divgi *et al.* 2008) and immunoscintography with a monoclonal antibody G250 linked to 99m-technetium are being investigated (Oosterwijk and Debruyne 1995).

The frequent use of US, CT and MRI has increased incidental findings in approximately 10-60% of all RCC cases (Homma *et al.* 1995), and has improved the survival of RCC patients since the 1970's (Brenner and Hakulinen 2001). This and the use of immomodulators have increased the five-year survival of RCC patients in twenty years from 52% (1974-76) to 60% (1989-95) (Greenlee *et al.* 2000). Therefore, the development of a protocol for RCC routine screening in the general population has been proposed (Tsui *et al.* 2000b, Tosaka *et al.* 1990).

#### 2.2 STAGING AND PROGNOSTIC FACTORS IN RCC

# 2.2.1 WHO AND HEIDELBERG CLASSIFICATIONS FOR TYPING OF RENAL TUMORS

The classification of renal tumors evolved in the 1980's. In 1981, the World Health Organization (WHO) classified epithelial renal tumors as renal cell adenoma and carcinoma (Mostofi 1981). Thoenes *et al.* classified renal tumors by tumor cell type, growth pattern, and grading of malignancy (Thoenes *et al.* 1986). In Heidelberg, in October 1996, the morphology was combined with genetic findings for a new classification, called the Heidelberg classification of renal tumors, in a workshop organized by the Union Internationale Contre le Cancer (UICC) and the American Joint Committee on Cancer (AJCC) (Kovacs *et al.* 1997, Störkel *et al.* 1997) (Table 3).

Table 3. The Heidelberg classification of epithelial renal tumors (According to Cheville et al. 2003, Amin et al. 2002, Zambrano et al. 1999, Kovacs et al. 1997, and Störkel et al. 1997).

Subtype according to Heidelberg classification	Incidence among all RCCs	Origin of the tumor type	Cell / tissue characteristics	Growth pattern	Five-year DSS for locally confined RCC
RCA, metanephric					
RCA, papillary		Proximal tubules			
RCA, oncocytic		Intercalated cells of collecting ducts	Round and vesicular cells with granular eosinophilic cytoplasm	Solid	
RCC, conventional / clear cell	70-80%	Proximal tubules	Clear cytoplasm; foci with eosinophilic cytoplasm are common	Solid, trabecular, tubular, cystic, rare papillary	69-76%
RCC, papillary*	10-15%	Proximal tubules	Small cells with scanty cytoplasm to large cells with abundant cytoplasm; basophilic, eosinophilic or pale staining  Type1: Basophilic, smaller	Papillary, tubulopapill ary or solid growth pattern	86-87%
RCC, papillary type 1  RCC, papillary type 2			cells with less cytoplasm  Type2: Eosinophilic, larger cells with more abundant cytoplasm		
RCC, chromophobe	3-5%	Intercalated cells of collecting ducts	Pale or eosinophilic granular cytoplasm, that stains blue with Hale's colloidal iron stain	Large solid sheets	87-100%
RCC, collecting duct (includes medullary RCC)	Very rare	Medulla of collecting ducts	Surrounded by demoplastic stroma	Irregular, channels- like structures	
RCC, unclassified	4-5%	Unclassified			24%

RCA=renal cell adenoma, RCC= renal cell carcinoma, DSS=disease-specific survival

Morphological differences exist between the 1998 WHO re-classification and the Heidelberg classification (Mostofi *et al.* 1998, Kovacs *et al.* 1997, Störkel *et al.* 1997). The former included granular cell and spindle cell carcinomas, the latter does not as the granular phenotype is a component of several RCC tumor types (Störkel *et al.* 1997), and the spindle cell type can develop in all types of RCCs as a result of the dedifferentiation process (Störkel *et al.* 1997). In 2004, WHO published the reassessed classification which is now based on both genetic and pathological abnormalities (Eble *et al.* 2004) (Table 4).

<sup>\*</sup>According to Delahunt and Eble (1997), papillary RCC should be divided into two subgroups: papillary RCC 1 and papillary RCC 2.

Table 4. The WHO 2004 histological classification of renal cell tumors (According to Lopez-Beltran et al. 2006).

RCC subtype according to the 2004 WHO classification	Incidence among all RCCs	Cell / tissue characteristics	Crowth notton	Genetic
Clear cell RCC	75%	Clear cytoplasm; cells with eosinophilic cytoplasm occasinaly	Growth pattern Solid, tubular, cystic, rare papillae	-3p, +5q22, -6q, -8p, -9p, -14q
Multilocular clear cell RCC	Rare	Clear cytoplasm, small dark nuclei	Cystic, no solid component	VHL gene mutation
Papillary RCC	10%	Type1 (basophilic) or type 2 (eosinophilic)	Tubulo- papillary, solid	-Y, +3q, +7, +8, +12, +16, +17, +20
Chromophobe RCC	5%	Pale or eosinophilic granular cytoplasm	Solid	-1, -2, -6, -10, -17, - 21
Carcinoma of the collecting ducts of Bellini	1%	Eosinophilic cytoplasm	Irregular channels	-1q, -6p, -8p, -13q, -21q, -3p
Renal medullary carcinoma	Rare	Eosinophilic cytoplasm	Reticular pattern	Unknown
Xp11 translocation carcinomas	Rare	Clear and eosinophilic cytoplasm	Tubulo-papillary	t (X; 1) (p11.2; q21), t (X; 17) (p11.2; q25), Other
Carcinoma associated with neuroblastoma	Rare	Eosinophilic cells with oncocytoid features	Solid	Allelic imbalance at 20q13
Mucinous tubular and spindle cell carcinoma	Rare	Tubules, extracellular mucin and spindle cells	Solid	-1, -4, -6, +7, -8, +11, -13, -14, +16, +17
RCC, unclassified	4-6%	Variable, sarcomatoid	Solid	Unknown

The Heidelberg classification is the basis of future classification systems in which the genetic and molecular properties, reflecting biological behavior rather than descriptive appearance, are the main criteria for classifying renal tumors (Moch 2000, Kovacs *et al.* 1997, Störkel *et al.* 1997). Each pathomorphological epithelial renal tumor entity should express specific chromosomal aberrations that completely reflect the pathomorphological phenotypes (Argani *et al.* 2007, Brunelli *et al.* 2003). Immunohistochemistry can be helpful in the differential diagnosis of renal neoplasms by characterizing the tumor entity according to a special antigen pattern (Magyarlaki *et al.* 2001). The Heidelberg classification identifies, for conventional, papillary, chromophobe and collecting duct RCCs, a strict correlation between the morphological phenotype and the complement of alterations evidenced by cytogenetic analysis of the neoplastic karyotype (Antonelli *et al.* 2003, Bodmer *et al.* 2002) (Figure 1). Progress in our knowledge of genetic alterations leads to new suggestions for RCC entities (Eble 2003). For instance, Gobbo *et al.* (2008) suggested that clear cell papillary RCC should be identified as an entity of its own. Different subgroups have different responses to therapy and outcome. With the progress of research, the Heidelberg classification may lead to more specific treatments in different subgroups of RCC patients.

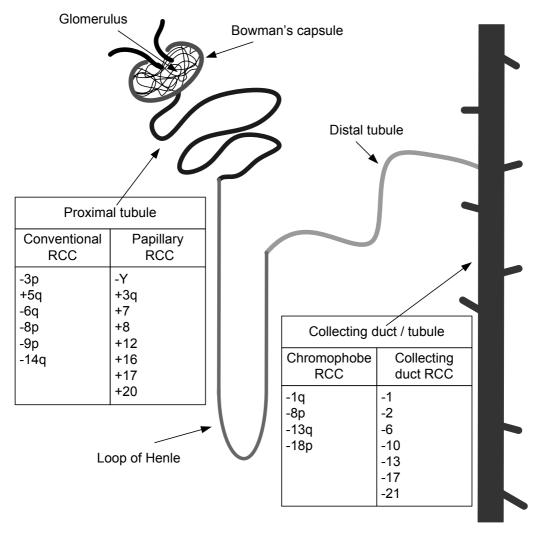


Figure 1. The genetic changes that characterize the different RCC subtypes according to the Heidelberg classification (Modified from Bodmer *et al.* 2002).

The number reflects the chromosome in which its genetic aberration is located.

- means loss of function.
- + means gain of function.
- p is the short arm of the chromosome.
- q is the long arm of the chromosome.

Conventional RCC represents 70-80% of all RCCs. Approximately 75% of sporadic conventional RCCs contain mutations in VHL, the tumor suppressor gene in the short arm of chromosome 3 (Maxwell *et al.* 1999, Gnarra *et al.* 1994), of which 50% show loss of heterozygosity (LOH) (Kovacs *et al.* 1997, Gnarra *et al.* 1994) and 10-20% silencing of the wild-type allele by promoter hypermethylation (Herman *et al.* 1994). Conventional RCC is genetically more homogeneous than the other types of RCC (Alimov *et al.* 2004). These tumors tend to be more symptomatic and have metastases more commonly than papillary and chromophobe RCCs (Amin *et al.* 2002).

Conventional RCC is associated with the best response to and most favorable outcome from IFN- $\alpha$  therapy compared to the other RCC subgroups (Motzer *et al.* 2002a). Cystic conventional RCC is often both low grade and low stage with a generally better prognosis to other conventional RCCs (Han *et al.* 2004).

*Papillary RCC* consists of two subtypes which are biologically and clinically distinct (Yang *et al.* 2005, Delahunt and Eble 1997).

Collecting duct RCC, a rare entity, is highly aggressive and the prognosis in metastatic disease is from some weeks to some months (Antonelli et al. 2003). Patients with collecting duct RCC are also at risk of transitional cell carcinoma / bladder cancer (Antonelli et al. 2003), which may reflect the common embryologic origin of collecting duct and urothelial cells. The positivity for Ulex europaeus lectin staining is characteristic of collecting duct RCCs (Delahunt and Eble 1997). A High incidence of Human Epidermal Growth Factor Receptor 2 (Her-2, c-erbB2) oncogene amplification and strong MET-protein expression has been found in collecting duct RCCs (Choi et al. 2006, Matei et al. 2005).

The *sarcomatoid component* may occur in all histological subtypes of RCCs and it associates with synchronous distant metastases and very short median survival of 6.6 to 8.0 months for stage II-IV RCC (Cheville *et al.* 2003, Cangiano *et al.* 1999)

*Unclassified RCC* includes those tumors that do not fit into other classes according to the Heidelberg classification. Unclassified RCCs are usually highly aggressive tumors (Zisman *et al.* 2002a).

The prognostic power of the Heidelberg classification has been investigated. It does not have independent prognostic ability, and thus it should not be considered as a major prognostic variable comparable to T-stage and histopathological tumor grade (Patard *et al.* 2005).

#### 2.2.2 TNM CLASSIFICATION SYSTEM FOR PATHOLOGICAL TUMOR STAGING

The first documented staging system for RCC, based on physical characteristics and tumor spread was published in 1958 (Flocks and Kadesky 1958). In the 1960's, it was modified by Robson *et al*, with the addition of tumor venous invasion (Table 5) (Robson *et al*. 2002). The poor correlation between the different Robson stages and survival, e.g. Robson stage II and III tumors had equal survival, led to the recommendation to use the TNM (tumor, node, metastases) system as a staging system for the extent of the tumor spread. In the Union International Contre Le Cancer's (UICC) TNM system, T means tumor, N node, and M metastasis. Since 1978, the TNM classification

system has integrated characteristics such as tumor size, vascular involvement, nodal spread and distant metastases (Bassil *et al.* 1985, Harmen 1978). In 1983, the pathological TNM (pTNM) classification system for renal carcinomas divided Robson stage III into subgroups. In 1992, Dinney *et al.* reported that after 10-15 years' follow-up, survival rates did not differ significantly between RCC tumors classified as T1-3b N0 M0 at the time of initial diagnosis.

Table 5. The Robson staging system for RCC in the 60's.

Stage grouping	
Stage I	Tumor confined to kidney
Stage II	Tumor invades perinephric fat but confined to Gerota's fascia or adrenal
Stage IIIa	Tumor invades renal vein or inferior vena cava
Stage IIIb	Tumor involves regional lymph nodes
Stage IIIc	Tumor involves both local vessels and lymph nodes
Stage IVa	Tumor involves local organs (i.e. colon, pancreas)
Stage IVb	Distant metastases

The pTNM classification system was updated by the UICC and the AJCC in 1997 (Table 6, Table 7). The cut-off between T1 and T2 tumors was increased from 2.5 cm to 7 cm, in order to increase the difference in survival from these two tumor types. Also the definitions of tumor thrombus involvement and nodal spread were changed (Guinan *et al.* 1997). Analysis of outcome in nephrectomized patients has shown that the 1997 TNM-system cut-off point between T1 and T2 tumors is too high, and a cut-off point of 4.5 - 5.0 cm has been suggested (Elmore *et al.* 2003, Zisman *et al.* 2001a). In 2002, the pTNM classification system was revised: T1 was divided into T1a and T1b by a cut-off point of 4 cm, according to the suitability for partial nephrectomy, and prognostication (Sobin and Wittekind 2002, Guinan *et al.* 1997). The cut-off point between T1 and T2 was not changed. A uniform staging classification, the TNM staging system, has improved the division of patients into radical or partial nephrectomy candidates (Janzen *et al.* 2003, Javidan *et al.* 1999).

Table 6. UICC stage grouping according to the 2002 TNM classification.

Stage grouping			
Stage I	T1	N0	M0
Stage II	T2	N0	M0
Stage III	T3	N0	M0
	T1, T2, T3	N1	M0
Stage IV	T4	N0, N1	M0
	Any T	N2	M0
	Any T	Any N	M1

Table 7. The UICC TNM classification for staging RCC tumors (ICD-O C64). Comparison of different modifications. (According to UICC TNM classification of malignant tumors, 4<sup>th</sup>, 5<sup>th</sup>, and 6<sup>th</sup> ed.) (Hermanek and Sobin 1987, Sobin and Fleming 1997, Sobin and Wittekind 2002)

	1987 TNM classification	1997 TNM classification	2002 TNM classification				
Prima	ary tumor (T)						
TX	Can not be assessed						
T0	No evidence						
T1	$\leq$ 2.5 cm and confined to the kidney	≤ 7.0 cm in greatest dimens	ion, limited to the kidney				
T1a	Not used		≤ 4 cm				
T1b	Not used		$>$ 4 cm but $\leq$ 7 cm				
T2	> 2.5 cm and confined to the kidney	> 7.0 cm in greatest dimens	ion, limited to the kidney				
Т3	Extends into adrenal gland, perinephric fat, major vessels	Extends into adrenal gland, veins	perinephric tissues, major				
T3a	Extends into adrenal gland, perinephric fat	Extends into adrenal gland, perinephric tissue	Extends into adrenal gland, perinephric tissues <sup>1</sup>				
T3b	Extends into inferior vena cava	Extends into renal vein(s), vena cava	Extends into renal vein(s) <sup>2</sup> , vena cava or its wall below diaphragm				
ТЗс	Extends into vena cava above the diaphragm  Extends into vena cava its wall above diaphragm						
T4	Directly extends beyond Gerota's fascia						
Regio	Regional lymph nodes (N)						
NX	Can not be assessed						
N0	No involvement						
N1	Metastasis into a single lymph node < 2 cm	Metastasis into a single regi	ional lymph node				
N2	Metastasis into a single lymph node 2 - S cm, multiple nodes < 5 cm						
N3	Metastasis into lymph nodes > 5 cm	Not used					
Dista	nt metastases (M)						
MX	Cannot be assessed						
M0	No						
M1	Yes						

<sup>&</sup>lt;sup>1</sup> Includes renal sinus (peripelvic) fat.

Pathological tumor stage (T-stage) has been observed to be the most important factor in predicting the survival of patients who have undergone nephrectomy (Delahunt *et al.* 2002, Robson *et al.* 2002, Tsui *et al.* 2000a). The observed five-year survival is 83% for stage T1, 57% for T2, 42% for T3, and 28% for T4 (Tsui *et al.* 2000a). For patients with stage I disease (tumor confined to the kidney) the five-year survival is approximately 90%, and for those with stage I and chromophobe RCC it is almost 100% (Zisman *et al.* 2001b). The five-year survival rate for stage III disease is approximately 50% (Zisman *et al.* 2001b). There is an 80% difference in survival rates between patients with local disease compared to those with advanced disease and distant metastases (ACS 2004). Additionally, in a retrospective review of 2 473 RCC patients from 1975 - 1985, regardless of T-stage, tumor size was observed to have an inverse association with survival (Guinan *et al.* 

<sup>&</sup>lt;sup>2</sup>Includes segmental (muscle-containing) branches.

1995). However, this kind of association has not been found in all studies (Lopez Hänninen *et al.* 1996).

T-stage can be used in estimating the correct duration and frequency of follow-up of RCC patients after nephrectomy. RCC with a diameter of less than 3.0 cm grows slowly; only 2.5% have metastases during the first three years (Bosniak *et al.* 1995). Therefore, in the treatment of those in whom surgery is contraindicated, careful monitoring (watchful waiting) by computed tomography (CT scan) may be used (Roberts *et al.* 2005, Bosniak *et al.* 1995). Additionally, a high T-stage has been used as an inclusion criterion for adjuvant treatments in trials (Atzpodien *et al.* 2005, Repmann *et al.* 2003). The updated TNM system is a useful tool correlating with survival and disease-free periods, although modifications in the TNM system may cause difficulty in comparing outcome data in different studies (Belldegrun *et al.* 1999, Störkel *et al.* 1989).

Several studies have been published on the prognostic power of the 2002 TNM classification in RCC patients (Ficarra *et al.* 2007, Gilbert *et al.* 2006). The TNM 2002 classification is useful, but some adjustments have been proposed, particularly concerning the tumor size cut-off (Bonsib 2005, Ficarra *et al.* 2005, Wunderlich *et al.* 2004, Zisman *et al.* 2001b), assessment of the invasion of the renal sinus fat tissue (Bonsib 2005), assessment of renal vein or vena cava inferior invasion (Kim *et al.* 2004a), and invasion of the ipsilateral adrenal gland (Kirkali *et al.* 2007a).

Bonsib (2005) observed that the incidence of renal sinus fat tissue invasion is higher when tumors exceed 4 cm (Bonsib 2005), and that T1b-T2 conventional RCC cases are rare if careful evaluation of sinus invasion is performed (Bonsib 2005). According to the study of Ficarra et al. (2004), the 2002 TNM staging system does not seem to be able to differentiate cancer-specific survival between pT1b and pT2 RCC (Ficarra et al. 2004). Ficarra et al. proposed reclassification of pT3-4 RCC. Similar outcomes in patients with stage T3a tumors and patients with tumors confined to the renal capsule (T1-2) have been reported (Gilbert et al. 2006, Roberts et al. 2005). RCC patients with microscopic vascular invasion have been associated with poorer prognosis (Kirkali and van Poppel 2007b, Van Poppel et al. 1997). Approximately 25% of RCCs extend into the renal vein or the inferior vena cava (Hatcher et al. 1991). A thrombus in the inferior vena cava is a prognosticator for cancer recurrence (Kirkali and van Poppel 2007b). A difference in five-year survival has been observed between RCC with a free-floating tumor thrombus in the inferior vena cava (69%) and RCC that has invaded the inferior vena cava wall (25%) (Hatcher et al. 1991). A complete inferior vena cava (IVC) thrombectomy, even in a metastatic setting, may result in improved survival, and it also leads to better quality of life (Kirkali and van Poppel 2007b). Renal vein extension may be a factor related to concomitant positive lymph node status or distant metastases (Ljungberg et al. 1995, Selli et al. 1983, Skinner et al. 1971). However, the therapeutic

value of lymph node dissection remains unproven (Mickish 1999). Renal pelvis invasion seems to associate with higher stage tumors with their attendant prognosis (McNichols *et al.* 1981). Collecting system invasion has been associated with decreased survival in low stage tumors (Palapattu *et al.* 2003).

#### 2.2.3 WHO CLASSIFICATION FOR HISTOPATHOLOGICAL TUMOR GRADING

The first grading system was reported in 1932 (Hand and Broders 1932). Later, many grading systems were created by, e.g. WHO (Table 8) with revision in 2004 (Eble *et al.* 2004), Fuhrman *et al.* in 1982 (Table 9), and Thoenes *et al.* in 1986. None of the tumor garding systems has gained universal acceptance (Kanamaru *et al.* 2001, Medeiros *et al.* 1997).

In grading systems, the major criteria are nuclear and nucleolar appearances, while in some systems, tumor architecture and cell type is also included (Mostofi *et al.* 1998, Goldstein 1997, Fuhrman *et al.* 1982, Syrjänen and Hjelt 1978, Skinner *et al.* 1971). The WHO grading system is based on the size and prominence of nucleoli (Eble *et al.* 2004, Mostofi *et al.* 1998) (Table 8), while the Fuhrman grading system is based on nuclear size, shape, and presence or absence of nucleoli (Fuhrman *et al.* 1982). The WHO grading system contains three grades, whereas the Fuhrman contains four.

Table 8. The WHO classification 1998 / 2004 for histopathological tumor grading.

Grade	WHO
1	Anaplasi and nuclei are smaller than in non-neoplastic tubule
2	Anaplasi and nuclei are same size as in non-neoplastic tubule
3	Anaplasi and nuclei are larger than in non-neoplastic tubule

Additionally, the percentage of sarcomatoid lesions is analyzed.

Table 9. The Fuhrman classification for histopathological tumor grading.

Grade	Fuhrman
1	Nuclei are small, round and uniform (10um) with inconspicuous or absent nucleoli.
2	Nuclei are slightly irregular (15 um) with small nucleoli
3	Nuclei are very irregular (20um) with large and prominent nucleoli
4	Nuclei are large and pleomorphic often polylobed and bizarre (>20um)

Several studies have failed to demonstrate any statistically significant differences in the survival of patients with different grades, when all three or four grades are analyzed separately (Uchida *et al.* 2002, Kanamaru *et al.* 2001, Rioux-Leclercq *et al.* 2000, Usubutyn *et al.* 1998, Selli *et al.* 1983). In one study, the median five-year disease-specific survival (DSS) was 94% for G1, 86% for G2, 59% for G3, and 31% for G4 (Ficarra *et al.* 2001). In a Scandinavian study, the five-year DSS rate was 87% for G1, 71% for G2, 46% for G3, and 15% for G4, when the study included consecutive patients from 1971 to 2000 (Gudbjartsson *et al.* 2005).

Currently, different grading systems are utilized at different institutions. Tumor-grading systems have been criticized because of their subjectivity in tumor evaluations (Lanigan et al. 1994). Therefore, comparison of different patient cases with respect to histopathological grade is difficult. More quantitative measures which describe the size or the shape of the nuclei have been requested by pathologists. Nuclear roundness (Delahunt et al. 1994) and a measure of the median nuclear volume (MNV) (Soda et al. 1999) have been found to be independent prognostic factors in a few studies. In 1997, an international consensus conference on RCC by UICC and AJCC outlined recommendations for the grading of RCC (Goldstein 1997): the grading system should be based on standardized and reproducible criteria that reflect the heterogeneity of nuclear and nucleolar features within a tumor, and each grade should result in significant differences in patient outcome. Recently again, a joint group of urologists and pathologists has published a proposal that the criteria for nuclear grading should be different for the subtypes of RCC according to the Heidelberg classification (Paner et al. 2006). The chromophobe type typically consists of irregular nuclei and prominent nucleoli, and therefore, a new classification model for chromophobe RCC has been proposed (Paner et al. 2006). Additionally, reducing the grades in the Fuhrman system has been proposed, for better outcome stratification (Rioux-Leclercq et al. 2007, Lohse et al. 2002, Bretheau et al. 1995).

#### 2.2.4 PROGNOSTIC MODELS

RCC is a heterogeneous disease: the prognosis differs even within the same stage and grade. Prognostic factors aim to specify the diagnosis, staging and prognosis, and they may help to determine the follow-up. Additionally, prognostic factors are used in clinical trial design, in interpretation, and they may guide targeted cancer therapies. Prognostic models, anagrams and nomograms, have been developed to find those nephrectomized RCC patients who potentially have a long-term recurrence-free interval and survival, as well as those mRCC patients who have long-term survival.

#### 2.2.4.1 PROGNOSTIC MODELS IN LOCALIZED DISEASE

Tumor size and grade are the most important prognostic factors for the survival of RCC patients with a locally confined disease. Table 10 shows the prognostic algorithms and nomograms for survival in RCC.

The MSKCC nomogram was developed for use in predicting disease recurrence after nephrectomy in localized RCC patients, and it consisted of TNM stage, tumor size, histology, and symptoms at presentation (Kattan *et al.* 2001). The MSKCC nomogram was reassessed in 2005. It focused on

conventional RCC and also included Fuhrman grade, histological necrosis, and microvascular invasion (Sorbellini *et al.* 2005).

At the Mayo Clinic, the *SSIGN-nomogram* was developed after Frank *et al.* (2002) found the following independent prognostic factors: TNM staging system, tumor size, nuclear grade and coagulative tumor necrosis. In the SSIGN nomogram, scores are presented as follows: +1 for pT2, +2 for pT3, +2 for pN1 or pN2, +4 for pM1, +2 for tumor size  $\geq 5$  cm, +1 for grade 3, +3 for grade 4, and +2 for tumor necrosis. Five-year cancer-specific survival rates were 99.4% for scores 0 to 1, and 7.4% for score 10. SSIGN has been validated and found to have a high degree of prognostic accuracy (Ficarra *et al.* 2007).

According to Lungberg *et al.* (1999), no follow-up in patients with diploid pT1-2 tumors or with aneuploid pT1 tumors of < 5 cm is needed.

Table 10. Prognostic algorithms and nomograms for survival in RCC.

Reference	Year	No. of Patients	Tumor Subtype	Prognostic Factors	Prognostic Information
Ljungberg <i>et al.</i> (Umeå)	1999	187	All	T-stage, DNA ploidy in T1-T2 tumors	Recurrence, Survival
Tsui <i>et al.</i> (2000a). (UCLA)	2000	643	All	TNM stage, nuclear grade, performance status	Survival
Kattan et al. (MSKCC)	2001	601	All	TNM stage, tumor size, histology, symptoms	Recurrence
Zisman et al. (2002b) (UCLA)	2002	292	All	Nuclear grade, performance status	Survival
Frank <i>et al.</i> (SSIGN, at Mayo Clinic)	2002	1801	Conventional, also metastatic	TNM stage, tumor size, nuclear grade, histologic necrosis	Recurrence
Patard et al. (Rennes University Hospital)	2004	4202	All	TNM stage, nuclear grade, performance status	Survival
Sorbellini <i>et al.</i> (MSKCC)	2005	701	Conventional	TNM stage, tumor size, nuclear grade, histologic necrosis, microvascular invasion, symptoms	Recurrence

MSKCC = Memorial Sloan Kettering Cancer Center

UCLA = University of California

#### 2,2,4,2 PROGNOSTIC MODELS IN METASTASIZED DISEASE

Therapies for mRCC cause a wide variety of adverse effects, which reduce the quality of life. Determining the prognostic factors for survival in mRCC patients is valuable in directing therapy for those patients who would benefit from it.

In mRCC, many independent prognosticators have been reported. The most often presented factors are performance status, time to metastases, number of metastatic sites and prior nephrectomy. Several models have been developed for predicting the likelihood of response to therapy and to predict survival. Table 11 and Table 12 show the prognostic algorithms and nomograms in mRCC for response to treatment and long-term survival.

Table 11. Prognostic algorithms and nomograms for survival in mRCC between 1980 and 1999.

Reference	Year	No. of Patients	Therapy Administered	Tumor Subtype	Prognostic Factors	Prognostic Information
Maldazys and deKernion (UCLA)	1986	181	Chemotherapy	All	Performance status, disease- free interval, nephrectomy status, metastatic site	Survival
Elson <i>et al.</i> (ECOG)	1988	610	Unspecified	All	Performance status, time from diagnosis to treatment, nephrectomy status, no. of metastatic sites	Survival
de Forges <i>et al.</i> (Institut Gustave Roussy)	1988	134	Unspecified	All	Disease-free interval, presence of liver metastasis, no. and size of lung metastases, ESR / weight loss	Survival
Neves <i>et al.</i> (Mayo Clinic)	1988	158	Unspecified	All	Grade of the primary lesion, weight loss, no. of metastatic sites, nephrectomy status	Survival
Palmer <i>et al</i> . (Cetus)	1992	327	IL-2	All	Performance status, time from diagnosis to treatment, no. of metastatic sites	Survival
Fosså <i>et al.</i> (Multi-institutional study)	1994	295	IFN-α, chemotherapy	All	Performance status, ESR, ≤10% weight loss	Survival
Mani et al. (YCC)	1995	84	IFN-α, IFN-β, IFN-γ, IL-2	All	Performance status, sarcomatoid histology, bone metastases	Survival
Lopez Hänninen <i>et al.</i> (Medizinische Hochschule Hannover)	1996	215	IFN-α + IL-2 ± 5-FU ± 13CRA	All	ESR, LDH, neutrophil count, hemoglobin, extrapulmonary metastases, bone metastases	Survival
Motzer et al. (MSKCC)	1999	670	Cytokines, chemotherapy	All	Performance status, nephrectomy status, LDH, hemoglobin, corrected calcium	Survival

13CRA = 13-cis-retinoid acid

ECOG = the Eastern Cooperative Oncology Group

ESR=erythrocyte sedimentation rate

LDH=lactate dehydrogenase

MSKCC = Memorial Sloan Kettering Cancer Center

UCLA = University of California,

YCC = Yale Comprehensive Cancer Center

Table 12. Prognostic algorithms and nomograms for survival in mRCC between 2000 and 2008

Reference	Year	No. of Patients	Therapy Administered	Tumor Subtype	Prognostic Factors	Prognostic Information
Motzer et al. (MSKCC) (2002b)	2002	463	IFN-α	All	Performance status, time from diagnosis to start of therapy, LDH, hemoglobin, corrected calcium	Survival
Zisman et al. (UCLA)	2002	262	IL-2 or IFN-α (197 pts), other (65 pts)	All	T-stage, nodal involvement, nuclear grade, no. of symptoms, immunotherapy	Survival
Négrier <i>et al</i> . (Group Francais d'Immunotherapie)	2002	782	IFN- $\alpha \pm IL$ -2	All	Performance status, no. of metastatic sites, disease-free interval, signs of inflammation, hemoglobin	Survival, rapid progression
Atzpodien (Medizinische Hochschule Hannover)	2003	425	IFN- $\alpha$ + IL-2 ± 5-FU ± 13CRA	All	Neutrophil count, LDH, CRP, time from diagnosis to start of therapy, no. of metastatic sites, bone metastases	Survival
Motzer et al. (MSKCC)	2004	251	New agents	All, if cytokine refractory disease	Performance status, hemoglobin, corrected calcium	Survival for those who enter clinical trials of new agents
Choueiri <i>et al.</i> (Cleveland Clinic Foundation)	2007	358	IFN- $\alpha$ ± IL-2 ± chemotherapy	All	Performance status, hemoglobin, no. of metastatic sites, involved kidney of primary tumor	Long-term survival
Cho et al (Yonsei University)	2008	197	Immunotherapy	All	Performance status, N stage, no. of metastatic sites, sarcomatoid differentiation, liver metastasis	Survival
Motzer et al. (MSKCC) (2008a)	2008	375	Sunitinib	Conventi onal RCC	Performance status, time from diagnosis to start of therapy, nephrectomy status, no. of metastatic sites, presence of liver or lung metastases, LDH, corrected calcium, hemoglobin, alkaline phosphatase, thrombosytosis	Probability of 12-month progression- free survival

LDH=lactate dehydrogenase

MSKCC = Memorial Sloan Kettering Cancer Center

UCLA = University of California,

One of the first publications on prognostic factors in mRCC was published in 1986. At *UCLA*, Maldazys and deKernion (1986) found that performance status, disease-free interval and presence of pulmonary metastases should be included as independent factors for predicting outcome of mRCC patients. Elson *et al.* (1988) found that age and sex were not independent prognostic factors. A scoring system was developed to classify patients with metastatic disease into five categories based on ECOG performance status (ECOG-PS), time from diagnosis to metastasis, prior chemotherapy, weight loss, and number of metastatic sites.

UCLA developed the mathematical equations for estimating survival after radical nephrectomy for education and counseling purposes (Patard *et al.* 2004, Zisman *et al.* 2002b). In mRCC patients, T-stage, nodal involvement, nuclear grade, number of symptoms, and previous immunotherapy were observed to be independent prognostic factors for survival, while in non-mRCC patients, only nuclear grade and performance status were independent prognostic factors (Zisman *et al.* 2002b).

In *a multi-institutional study*, prognostic factors and survival were analyzed in mRCC patients treated with either chemotherapy or IFN- $\alpha$  (Fosså *et al.* 1994). In those RCC patients with favorable risk, the three-year survival rate was 48% for IFN- $\alpha$ , and 15% for chemotherapy, but in those mRCC patients with poor or intermediate risk, no significant difference in survival was observed.

In the study of the *Group Francais d'Immunotherapie*, the following prognostic factors were observed: ECOG performance status, number of metastatic sites, disease-free interval, biological signs of inflammation, and hemoglobin level (Négrier *et al.* 2002). Négrier *et al.* (2002) also found four independent factors for rapid disease progression. These were presence of hepatic metastases, less than one-year disease-free interval from primary RCC tumor, more than one metastatic site, and elevated neuthrophil counts. If three of these factors were found, the probability of rapid progression within three months was more than 80%, regardless of the treatment.

MSKCC models for mRCC are among the most widely used models. In the first analysis, 670 mRCC patients were treated with chemotherapy or immunotherapy (Motzer et al. 1999). Independent prognostic factors for survival were: Karnofsky performance status less than 80%, absence of prior nephrectomy, baseline levels of serum lactate dehydrogenase more than 1.5 times the upper normal limit, hemoglobin below the lower limit of normality, and corrected serum calcium above 10 mg/dL (Motzer et al. 1999). The survival in the poor risk group was less than six months with both chemotherapy and IFN-α therapy (Motzer et al. 1999), and the difference in survival between chemotherapy and IFN-α therapy was greater in those patients who belonged to the groups of intermediate or favorable risk (Motzer et al. 1999). Motzer et al. (1999) also found that survival was greater for those RCC patients who were treated in the 1990's compared to those treated in earlier years.

After the establishment of the role of cytoreductive nephrectomy in mRCC patients prior to IFN- $\alpha$  therapy (Flanigan *et al.* 2001, Mickisch *et al.* 2001), the MSKCC model was reassessed (Motzer *et al.* 2002b). The new prognostic factor was an interval from diagnosis to treatment of less than one year. In the study of 463 patients treated with IFN- $\alpha$  as first-line therapy, median survival time was 13 months, and time to progression 4.7 months. Patients were categorized into three different risk

groups according to the number of risk factors predictive of short survival: zero scores for the group with favorable risk, one or two scores for the group with intermediate risk, and three or more scores for the group with poor risk. The patients were classified into three groups with three different predictive outcomes. Median survival time was 30 months for the group with favorable risk, 14 months for the group with intermediate risk, and five months for the group with poor risk. This model was validated independently at the Cleveland Clinic Foundation (Mekhail *et al.* 2005); it has been used in phase III trial design and in evaluation of the new, targeted agents (Escudier *et al.* 2007b, Hudes *et al.* 2007, Motzer *et al.* 2007).

Motzer *et al.* (2004) from MSKCC created a new model for those mRCC patients who have received prior cytokine therapy and are candidates for clinical trials of novel targeted agents as second line therapy. They found that poor performance status, low level of hemoglobin, and high level of corrected calcium represented poor prognosis. This model can be used in the clinical trial design of phase III trials as stratification factors and in interpreting the outcome of phase II trials.

A new MSKCC model for predicting the probability of 12-month progression-free survival in patients who receive sunitinib was published in 2008 (Motzer *et al.* 2008a).

Choueiri *et al.* (2007) at *Cleveland Clinic Foundation* developed a simple scoring system with three different risk groups for determining the long-term survival. They found in their study that independent prognostic factors for short-term survival were ECOG performance status higher than zero, number of involved metastatic sites more than two, baseline levels of hemoglobin below the lower limit of normal, and primary RCC in the left kidney in previously untreated mRCC patients who are treated with IFN- $\alpha$  or IL-2 with or without chemotherapy. In the study, 63% of all the patients were short-term survivors. The other two groups represent long-term survivors.

#### 2.3 BIOMARKERS RELATED TO MOLECULAR MECHANISMS IN RCC

Carcinogenesis is a multistep process in which mutations accumulate with no proper activation of repair mechanisms. According to current knowledge, mistakes in deoxyribonucleic acid (DNA) repair are an important reason for carcinogenesis. Additionally, if cancer occurs, some defect in the immune system has taken place.

The following factors lead to carcinogenesis: 1) self-sufficient production of growth signals activated by oncogenes, 2) insensitivity to negative growth-inhibitory signals which leads to inactivation of tumor suppressor genes, 3) ability to avoid apoptosis, 4) limitless proliferative capacity, 5) induction of angiogenesis, and 6) ability to invade new organs and metastasize

(Hanahan and Weinberg 2000). Oncogenes promote growth as an accelerator, and tumor suppressor genes inhibit growth as a brake in the cell cycle. The activation of oncogenes and the inactivation of tumor suppressor genes are important phenomena in cancer development.

#### 2.3.1 pVHL, VON HIPPEL-LINDAU PROTEIN, MODULATOR OF HYPOXIC RESPONSE

pVHL, a tumor suppressor gene product, is expressed especially in the kidney's proximal renal tubule (Corless *et al.* 1997, Iliopoulos *et al.* 1995). Approximately 61-75% of sporadic conventional RCCs contain mutations in *VHL*, in the short arm of chromosome 3 (3p25-26) (van Houwelingen *et al.* 2005, Gnarra *et al.* 1994, Maxwell *et al.* 1999), of which 50% show loss of heterozygosity (LOH) (Kovacs *et al.* 1997, Gnarra *et al.* 1994) and 10-20% silencing of the wild-type allele by promoter hypermethylation (Herman *et al.* 1994). VHL is associated with carcinogenesis.

The function of pVHL is ubiquitylation of hypoxia-inducible factor (HIF); therefore, it modulates the hypoxic response; VHL protein can bind to hypoxia inducible factor-1 alpha (HIF- $1\alpha$ ) and target this factor for destruction in the presence of oxygen. HIF in turn controls the expression of several proteins, including carbonic anhydrase 9 (CA9) and proteins involved in angiogenesis, i.e. vascular endothelial growth factor (VEGF) and EPO, via oxygen-dependent ubiquitination (van Houwelingen *et al.* 2005, George and Kaelin 2003). Normally, VHL down regulates vascular endothelial growth factor (VEGF) by different pathways. In VHL-defective cancer cells, increased concentrations of VEGF and EPO are observed.

#### 2.3.2 CA9, HYPOXIA ASSOCIATED ENZYME

CA9, a member of the carbonic anhydrase family, is suggested to play a role in the regulation of cell proliferation in response to hypoxic conditions. Low CA9 expression associates with the absence of VHL mutation and aggressive tumor characteristics in conventional RCC (Pantuck *et al.* 2007). CA9 may indicate those patients who benefit from IL-2, as low CA9 expression associates with lower survival compared to high CA9 expression in mRCC patients who receive IL-2 (Atkins *et al.* 2005, Bui *et al.* 2003). It has also been suggested that CA9 may indicate those patients who benefit from CA9-targeted therapies. It is also being investigated whether CA9 may indicate those patients who are potential candidates for adjuvant therapy.

#### 2.3.3 p53, BIOMARKER OF CELL CYCLE POINT

p53, a tumor suppressor gene product, is a promoter of cell growth arrest and apoptosis (Choisy-Rossi and Yonish-Rouach 1998). Activated p53 elicits several cellular responses, including apoptosis and cell cycle arrest (Reich and Levine 1984), and responds to DNA damage at the

restriction checkpoint of the G1 phase of the cell cycle (May and May 1999). In normal cells, p53 is usually undetectable (Finlay *et al.* 1988). Mutant p53 accumulates in cell nuclei and can be immunostained (Reich and Levine 1984), whereas wild-type p53, because of its short half-life, is usually undetectable by routine immunohistochemistry (Reich and Levine 1984). p53 may be upregulated in part by VHL, accounting for some of the tumor suppressive functions of VHL in RCC (Galban *et al.* 2003).

Published results on the association of p53 with survival have been controversial, some studies suggesting positive p53 associating with poor survival (Shvarts *et al.* 2005, Zigeuner *et al.* 2004, Uchida *et al.* 2002, Haitel *et al.* 2000, Moch *et al.* 1997, Uhlman *et al.* 1994), while others have observed no association (Itoi *et al.* 2004, Olumi *et al.* 2001, Rioux-Leclercq *et al.* 2000, Hofmockel *et al.* 1996, Bot *et al.* 1994, Lipponen *et al.* 1994). In the study of Phuoc *et al.* (2007), p53 was significantly associated with survival in univariate analysis, but the association was not independent. In some studies, the association of p53 and survival has been investigated in a group of RCC patients with both locally confined and primary metastatic RCC; thus, patient selection varies in different studies (Olumi *et al.* 2001).

# 2.3.4 Ki-67. PROLIFERATION BIOMARKER

Ki-67, a proliferation biomarker, is expressed throughout the active phases of the cell cycle, and serves as a good marker for proliferative activity in cell nuclei (Gerdes *et al.* 1984). Ki-67 accumulates during the cell cycle from G1 to mitosis, and is at its lowest level after mitosis (du Manoir *et al.* 1991). The percentage of nuclei staining by immunohistochemistry reflects Ki-67 expression (Olumi *et al.* 2001). Ki-67 has been reported to independently predict survival following nephrectomy in many studies (Dudderidge *et al.* 2005, Bui *et al.* 2004, Itoi *et al.* 2004, Rioux-Leclercq *et al.* 2000, Aaltomaa *et al.* 1997). Ki-67 has been observed to increase in sarcomatoid change (Kanamaru *et al.* 1999), indicating different protein expression profiles in different entities according to the Heidelberg classification.

# 2.3.5 COX-2, BIOMARKER FOR INFLAMMATION AND NEOPLASIA

Cyclo-oxygenase-2 (COX-2), an isoform of the COX<sup>3</sup> enzyme, is an inducible form of an enzyme involved in the first steps of prostaglandins and thromboxane synthesis. COX-2 converts arachidonic acid first into prostaglandin G2, and afterwards by peroxidase activity into prostaglandin H<sub>2</sub>, a precursor of the prostaglandins (Taketo 1998). COX-2 is suggested to play a physiological role in fetal nephrogenesis (Khan *et al.* 2001). COX-2 increases in inflammation and neoplasia (Miyata *et al.* 2003, Hara *et al.* 2002, Maitra *et al.* 2002, Nose *et al.* 2002, *et al.* Taketo 1998), and is undetectable in most normal tissues (Mungan *et al.* 2006, Yoshimura *et al.* 2004).

The conversion of procarcinogens to proximate carcinogens is catalyzed by the peroxidase activity of COX-2 (Elinq *et al.* 1990).

COX-2 is highly induced by stimulus of oncogenes, cytokines, growth factors, and tumor promoters (Smith *et al.* 2000, Herschman 1996, Subbaramaiah *et al.* 1996). Associations between COX-2 over-expression and antiapoptotic ability, tumor invasiveness, tumor growth, angiogenesis, and immunosuppression, as well as multidrug resistance in cancer have been reported (Cao and Prescott 2002, Masferrer *et al.* 2000, Subbramaiah *et al.* 1996, Tsujii and DuBois 1995).

Cytoplasmic/membranous COX-2 staining by immunohistochemistry reflects COX-2 protein expression (Cho *et al.* 2005). The study results on associations of COX-2 with tumor stage, grade, and survival have been contradictory. Yoshimura *et al.* (2004) demonstrated that COX-2 was expressed at its highest in G1, as well as in pT1 RCC tumors, compared to other RCC tumors in grade and T-stage, while in Hashimoto *et al*'s study (2004), more COX-2 was found at the higher tumor grade, as well as stage. A significant association has been observed between COX-2 and Ki-67 expression (Miyata *et al.* 2003).

# 2.3.6 Her-2, BIOMARKER OF PROTO-ONCOGENE PRODUCT

Her-2, a proto-oncogene product, is a member of the ErbB family of receptor tyrosine kinases. Her-2 functions in secretory epithelial tissues, and regulates intracellular signaling cascades (Arteaga *et al.* 2001, Olayioye *et al.* 2000). Her-2 is over-expressed in approximately 20-30% of human adenocarcinomas (Latif *et al.* 2002, Lipponen *et al.* 1994, Slamon *et al.* 1989), and the over-expression is associated with metastatic phenotype and poorer prognosis, e.g. in breast and ovarian cancer (Slamon *et al.* 1989).

Gene amplification of *Her-2* can be investigated by cytogenetic analyses, such as fluorescent *in situ* hybridization (FISH), chromogenic *in situ* hybridization (CISH), and polymerase chain reaction (PCR). In breast cancer, FISH and CISH positivity are accurate predictors of response to trastuzumab (anti-Her2 therapy) (Isola *et al.* 2004, Lebeau *et al.* 2001). Receptor-mediated targeted tumor therapy with Herceptin® (RhuMAb HER-2), a recombinant humanized monoclonal anti-Her-2 antibody, has improved the survival of breast carcinoma patients both in adjuvant therapy and in therapy for metastatic disease (Smith *et al.* 2007, Montemurro *et al.* 2003).

Membranous staining of HER-2 in immunohistochemistry reflects HER-2 protein expression (Zhang *et al.* 1997). Her-2 receptor-specific tumor toxin, in an animal model, effectively reduced pulmonary tumors of advanced RCC (Maurer-Gebhard *et al.* 1998). Parallel associations of Her-2 expression between tumor stage and grade in RCC patients have been observed in many studies

(Zhang *et al.* 1997, Stumm *et al.* 1996), although in the study of Seliger *et al.* (2000) no such association was found. In the study of Hofmockel *et al.* (1997), higher tumor grades were seen when Her-2 expression was low, and higher T-stage associated with high Her-2. In the study of Phuoc *et al.* (2007), Her-2 protein expression did not correlate with Ki-67 protein expression.

In most *HER-2* gene amplification studies, *Her-2* gene amplification was observed neither by FISH analysis (Latif *et al.* 2002), messenger ribonucleic acid (mRNA) analysis (Stumm *et al.* 1996), nor PCR analysis (Selli *et al.* 1997, Zhang *et al.* 1997). Selli *et al.* (1997) found *HER-2* gene amplification in collecting duct RCC cases (45%). Therefore, *HER-2* gene amplification may be more pronounced in collecting duct RCC, than in other more common RCC types (Matei *et al.* 2005, Zhang *et al.* 1997). The association of *HER-2* gene amplification and HER-2 protein expression with the prognosis of RCC patients has been estimated in few studies and the results have been contradictory (Phuoc *et al.* 2007, Lipponen *et al.* 1994). Further studies are needed to determine whether HER-2 protein expression or *HER-2* gene amplification may be used as prognostic factors in RCC patients.

# 2.4 THERAPIES FOR RCC

# 2.4.1 SURGERY

# 2.4.1.1 NEPHRECTOMY FOR LOCALIZED DISEASE

Nephrectomy is the only curative treatment for locally confined RCC (Robson *et al.* 2002). For RCC patients with isolated metastases, nephrectomy in conjunction with metastasectomy can be considered aiming at curative treatment, especially if the disease-free interval has been long (Kavolius *et al.* 1998).

Giuseppe Zambeccarrius performed a successful nephrectomy in a dog in 1678. Gustav Simon of Heidelburg performed the first successful intentional nephrectomy in a woman in 1869. Postoperative sepsis and massive bleedings after nephrectomy were common in those days. Partial nephrectomy was first performed for renal tumor excision in 1887 by Czerny.

In 1969, Robson and colleagues reported a technique for radical nephrectomy, which increased survival rates in RCC (Robson *et al.* 2002). It still remains the gold standard for the treatment of RCC. Radical nephrectomy includes removal of the kidney with intact Gerotas fascia and the ipsilateral adrenal gland with early vascular ligation.

Many patients may be over treated by removal of the adrenal gland, but this allows for correct staging, and long-term follow-up has indicated that its removal is not harmful (Hellström et al.

1997). The incidence of adrenal involvement in histologically confirmed RCC is 3-4%, and the risk of adrenal metastases has been associated with left-sided, upper pole, advanced T-stage, poor differentiation, and multifocality (Li *et al.* 1996, Sagalowsky *et al.* 1994).

The therapeutic value of lymph node dissection remains unproven (Mejean *et al.* 2003, Mickisch *et al.* 1999). There is evidence that retroperitoneal lymph node dissection (RPLND) may prolong survival in selected patients (Canfield *et al.* 2006). An increase in five-year survival from 65% to 80% was reported after lymphadenectomy with stage II RCC, and from 47% to 60% with stage III RCC (Golimbu *et al.* 1986). In a retrospective analysis, radical RPLND was beneficial in less than 4% of 1 035 patients without distant metastases (Schafhauser *et al.* 1999).

The rate of lymph node involvement ranges from 13% to 30% (Blom *et al.* 1999, Giuliani *et al.* 1990). Extensive lymph node dissection may improve staging (Blom *et al.* 1999). Locoregional lymph node involvement has been associated with unfavorable prognosis with a five-year survival of 5-30% (Bassil *et al.* 1985, Nurmi 1984). Independent lymph node involvement without distant metastases has been found to be rare, found in fewer than in 1% of the patients (Johnsen and Hellsten 1997).

### 2.4.1.2 NEPHRON-SPARING SURGERY

Radical nephrectomy is not suitable for patients with bilateral tumors or a small unilateral tumor in a solitary functioning kidney. Partial nephrectomy (PN), also called nephron-sparing surgery (NSS) (Novick 1995, Patard 1983), has been developed for such patients. PN is also indicated when a disease might threaten the future functioning of the contralateral kidney. Such a disease may be chronic pyelonephritis, renal artery stenosis, ureteral reflux, nephrosclerosis, or diabetes (Licht and Novick 1993). During the past decade, the trend has been to promote nephron-sparing surgery at the expense of radical nephrectomy (Gill *et al.* 2002). Recently, PN has also been performed in patients with a normally functioning contralateral kidney (Novick 1995).

After PN, in patients with a clearly localized RCC tumor of 4 cm or less in diameter, particularly for a low TNM stage, such as  $T_1N_0M_0$ , the survival rates have been observed to be comparable to those for radical nephrectomy (Lee *et al.* 2000, Belldegrun *et al.* 1999). No differences in survival rates have been observed between centrally or peripherally located tumors following PN (Hafez *et al.* 1998). The criterion for PN is that the tumor has not invaded vascular structures located in the center of the kidney. Of RCC cases, 10-25% are multifocal, and thus not suitable for PN (Whang *et al.* 1995).

After PN, the observed five-year cancer-specific survival is 88-100% (Fergany *et al.* 2000, Hafez *et al.* 1999), and the 10-year cancer-specific survival 73% (Fergany *et al.* 2000). A long-term (10-years) preservation of renal function has been achieved in 93% of patients (Fergany *et al.* 2000, Lau *et al.* 2000). Postoperative local tumor recurrence after PN has occurred in 3-6% of patients (Derweesh and Novick 2003, Hafez *et al.* 1999), but for the subgroup of T<sub>3b</sub> RCC, a local recurrence rate of 10.6% has been observed (Hafez *et al.* 1997). It is suggested that local recurrence after PN is a manifestation of undetected microscopic multifocal RCC in the remaining kidney (Derweesh and Novick 2003). The risk of local recurrence after radical nephrectomy is not known, but it is very low (Derweesh and Novick 2003).

The maximum size of RCC suitable for PN has not been defined. Cancer-free survival is better in patients with tumors smaller than 4 cm than in patients with tumors between 4 and 7 cm (Hafez *et al.* 1999). Recently, also tumors of this size have been resected by partial nephrectomy with results that are comparable with radical nephrectomy (Leibovich *et al.* 2004). Data suggest that PN provides a long-term renal functional advantage over radical nephrectomy in those with an abnormal opposite kidney (Derweesh and Novick 2003, Lau *et al.* 2000). Quality of life is improved following PN (Clark *et al.* 2001), regardless of a complication rate of 9%, with the most bothersome being prolonged urinary fistula and delayed hemorrhage, which rarely require reoperation (Stephenson *et al.* 2004). PN may also be indicated in patients with a normal contralateral kidney, providing that the RCC tumor is single, small, and localized (Derweesh and Novick 2003). Patient selection has been a significant factor for outcome in this group (Derweesh and Novick 2003).

# 2.4.1.3 CYTOREDECTIVE NEPHRECTOMY

Cytoreductive nephrectomy has been performed since the 1960's to reduce symptoms from the primary tumor. It can be used to control flank pain, hematuria, anemia, hypercalcemia, and severe paraneoplastic syndrome. Embolization with postinfarction nephrectomy has also been used (Kurth *et al.* 1987). Angioinfarction instead of cytoredyctive nephrectomy may control local symptoms. Kim and Louie (1992) reported that in those patients with partial response to IL-2 treatment, the subsequent surgical resection of residual tumor may be beneficial.

Nephrectomy alone prolongs survival only in a minority of mRCC patients; it is more feasible in those patients with favorable prognostic factors such as low ESR and high hemoglobin (Onishi *et al.* 1989). Prior cytoreductive nephrectomy is justified in mRCC when later immunotherapy is planned (Flanigan *et al.* 2001, Mickisch *et al.* 2001). Radical nephrectomy followed by IFN- $\alpha$  monotherapy improves survival by 50% in patients with good performance status (Flanigan *et al.* 2001, Mickisch *et al.* 2001). However, in these studies, the combined morbidity rate was 18% due

to nephrectomy; 3% had a fatal complication. The patient selection for surgery is crucial due to high mortality rates (up to 17%) (Bennett *et al.* 1995). mRCC patients who have renal vein/inferior vena caval thrombus benefit from radical nephrectomy and thrombectomy, which improves disease-free survival (Parekh *et al.* 2005). Cytoreductive nephrectomy has also been associated with spontaneous regression in mRCC patients (Marcus *et al.* 1993, Gleave *et al.* 1998, Oliver *et al.* 1989).

### 2.4.1.4 METASTASECTOMY

Metastasectomy may prolong survival in patients with favorable prognostic factors (Samellas 1963). Surgery is potentially curative. Five-year survival rates from 20% to 70% have been achieved, when both primary tumor and solitary metastasis (mostly from lungs) have been resected (Kavolius *et al.* 1998, McNichols *et al.* 1981, Skinner *et al.* 1971). Resections of solitary second and third recurrences have been observed to increase five-year survival rates to 46% and 44%, respectively (Kavolius *et al.* 1998). Brain metastases are well-circumscribed with a surrounding pseudocapsule and they are often removable with low morbidity and mortality. The palliative benefit from the resection of brain metastases seems to be greater than from radiotherapy alone, and it also improves the performance status (Takashi *et al.* 1995). Long disease-free survival after adrenalectomy (with later steroid replacement) because of solitary adrenal gland metastasis has been achieved (Ertl and Darras 1999).

# 2.4.2 RADIOTHERAPY

Preoperative radiotherapy in local RCC has not been proved beneficial in prospective randomized studies. The applied radiation doses to the involved kidney and regional lymph nodes have been 30 Gy to 40 Gy with daily fractions of 2 Gy (Juusela *et al.* 1977). In this study, patients who received preoperative radiation therapy doses of 33 Gy with 15 fractions had a poorer five-year survival rate compared to those with only nephrectomy. However, preoperative radiotherapy has been observed to transform an inoperable tumor to an operable one in some RCC patients (van der Werf-Messing *et al.* 1973).

Many prospective randomized studies have found no benefit from postoperative adjuvant radiotherapy to the kidney bed and regional ipsilateral and contralateral lymph nodes (Kjaer *et al.* 1987, Finney 1973). In the study of Kjaer *et al.* (1987), the dose was 50 Gy in 20 fractions. Significant toxicity was observed in the stomach, duodenum, and liver.

Currently, according to retrospective studies, radiotherapy may be indicated in inoperable advanced local tumor stages and residual tumors, increasing the local control rate in RCC with lymph node

metastases, perinephric fat or adrenal invasion, or surgically positive margins (Kao *et al.* 1994, Rabinovitch *et al.* 1994, Stein *et al.* 1992). Although radiotherapy may decrease the local recurrence rate, no improvement in overall survival has been observed. Modern CT-assisted treatment planning decreases the toxicity of radiotherapy (Stein *et al.* 1992).

Palliative radiotherapy is used for ameliorating the symptoms due to metastases in, e.g. the brain, lung, or bone (Halperin and Harisiadis 1983). A typical palliative dose is 30 Gy in 10 fractions.

# 2.4.3 CHEMOTHERAPY

RCC has been proved to be unresponsive to chemotherapy with no improvement in survival. Vinblastine has been observed to have some activity in RCC with a response rate of less than 10%, except in one study where the response to vinblastine was 16% (Fosså *et al.* 1992a, Kuebler *et al.* 1984). Gemcitabine with capecitabine or 5-fluorouracil has a similar response rate of around 10% (Tannir *et al.* 2008, George *et al.* 2002), except in the study of Rini *et al.* (2000) where the rate was 17%. Gemcitabine with doxorubicin has antitumor activity in collecting duct, sarcomatoid, or with rapidly progressing RCC (Nanus *et al.* 2004, Milowsky *et al.* 2002). The addition of 5-fluorouracil to immunotherapy does not seem to increase the response rate (Négrier *et al.* 2000), but does increase the amount of adverse effects (Olencki *et al.* 2001). Capecitabine with thalidomide, docetaxel, or immunotherapy have produced response rates of less than 10% (Harshman *et al.* 2008, Marur *et al.* 2008, Wenzel *et al.* 2003). Treatment with pemetrexed also results in response rates of less than 10% (Thödtmann *et al.* 2003).

# 2.4.4 INTERFERON-α THERAPY

Immunotherapy was shown to be active in RCC by Tykkä *et al.* in 1978. Partially purified human leukocyte IFN- $\alpha$  (Cantell-type IFN) (Cantell and Hirvonen 1977) and lymphoplastoid IFN- $\alpha$  (Wellferon, Burroughs Wellcome Company) were first found to have activity in mRCC (Kirkwood *et al.* 1985, Neidhart *et al.* 1984, deKernion *et al.* 1983, Quesada *et al.* 1983). Subsequently, the cloning of the gene for IFN- $\alpha$  and recombinant genetic engineering allowed the production of rIFN- $\alpha$  (Pestka 1983). Two preparations of IFN- $\alpha$  are commercially available: rhIFN- $\alpha$ 2a (Roferon®, Hoffman LaRoche) and rhIFN- $\alpha$ 2b (Intron®, Schering-Plough Laboratories). It is generally accepted that there is no difference in the efficacy of these two preparations, although no direct comparison has been made. In addition, rhIFN- $\alpha$ 2c (Boehringer Ingelheim) has been licensed in some countries. The US Food and Drug Administration (FDA) approved rIFN- $\alpha$  for clinical use in hairy cell leukemia in 1986 (Pestka 1997), and afterwards for many other malignancies. IFN- $\alpha$  is active in multiple tumors, e.g. in cutaneous T-cell lymphomas, superficial bladder cancer, malignant neuroendocrine tumors. Kaposi sarcoma, and malignant melanoma. For

RCC, IFN- $\alpha$  therapy is preferentially used in Europe, while IL-2, the other cytokine therapy, is predominantly used in North America.

### 2.4.4.1 RESPONSE TO IFN- $\alpha$

The response rate to IFN- $\alpha$  is approximately 12%, ranging most often from 10% to 20% (Table 13). The majority (approximately 70%) of objective responses are partial responses (PR). The median duration of response is six to eight months, and rarely exceeds two years (Fosså 1988). In the Cochrane meta-analysis, based on four studies, on average, a 2.8 month improvement in median survival was achieved with IFN- $\alpha$  therapy compared to controls, although, the confidence limits could not be estimated (Coppin *et al.* 2007). Using overall survival as the appropriate measure of benefit, IFN- $\alpha$  has been associated with an increase in median survival of 3.8 months, a reduction in one-year mortality of 44%, and a reduction in two-year mortality of 26%, compared to non-immunotherapy controls (Coppin *et al.* 2007, MRCRCC 1999, Pyrhönen *et al.* 1999). Three percent of patients have shown prolongation of survival due to IFN- $\alpha$  (Minasian *et al.* 1993). IFN- $\alpha$  is effective in conventional type mRCC but its efficacy in other mRCC types is uncertain (Motzer *et al.* 2002a).

Table 13 shows the results of IFN- $\alpha$  treatment analysed in terms of survival in randomized studies. IFN- $\alpha$  is effective and increases survival in RCC patients with good prognosis. Pyrhönen *et al.* (1999) suggested in their study (IFN- $\alpha$  versus IFN- $\alpha$  plus vinblastine) that the survival benefit of IFN- $\alpha$  may be greater in those patients with adverse prognostic factors, such as poor performance status, age over 60 years, and male gender (Pyrhönen *et al.* 1999). In the study by Négrier *et al.* (2007), IFN- $\alpha$  was not beneficial in patients with intermediate prognostic factors (Négrier *et al.* 2007). IFN- $\alpha$  produces long-lasting remission in some patients (Motzer *et al.* 2000a, Minasian *et al.* 1993).

The prognostic value of secondary leucopenia as response to IFN-α therapy has been found contradictory in different studies (Buzaid *et al.* 1987, Muss *et al.* 1987, Umeda and Niijima 1986, Kirkwood *et al.* 1985, Quesada *et al.* 1985b).

Pegylated IFN- $\alpha$  (PEG-IFN- $\alpha$ ) has a longer half-life compared to standard IFN- $\alpha$ , which enables weekly doses. PEG-IFN- $\alpha$  is effective on a weekly schedule at a dose of 4.5 µg/kg subcutaneously (Feldman *et al.* 2008, Motzer *et al.* 2001a) with median overall survival of 31 months and comparable safety with standard IFN- $\alpha$  (Feldman *et al.* 2008, Motzer *et al.* 2001a).

Table 13. Benefit from IFN-α analysed in randomized studies.

No.	T)	Dose (million units MU),	CR + PR	Median overall survival	Survival
of pts	Therapy	schema and route	(%)	(months)	benefit
30	IFN-α2a	10-50 MU/m <sup>2</sup> im tiw until progression	6	7	No
30	MPA	1000mg im tiw for 5 wks, thereafter 1000mg im q1wk until progression	3	7	
41	IFN-α2b + vinblastine	IFN-α 8 MU sc tiw, vinblastine 0.1 mg/kg iv q3wk until progression or no change after a period of 3 months	20.5	16	No
35	MPA	500 mg im q1wk until progression or no change after a period of 3 months	0	10	
79	IFN-α2a + vinblastine	IFN-α max 18 MU tiw sc, until 12 months or 3 months after CR, but in PR and SD beyond 12 months + vinblastine 0.1 mg/kg iv q3wk	16.5	16.9	Yes
81	vinblastine	vinblastine 0.1 mg/kg iv q3wk until 12 months or 3 months after CR, but in PR and SD beyond 12 months	2.5	9.5	
167	IFN-α-2b	10 MU sc tiw until 3 months, 23 patients continued after 12 weeks	16*	8.5	Yes
168	MPA	300 mg po daily, 3 months	2*	6	
122	IFN-α	9 MU sc tiw, max 6 months	4.4*	15.2	No
125	IL-2	9 MU sc, max 6 months	4.1*	15.3	
122	IFN-α+ IL-2	IFN-α 6 MU tiw + IL-2 9 MU sc, max 6 months	10.9*	16.8	
123	MPA	200mg po daily, 6 months	2.5*	14.9	
hs.	122	122 IFN-α + IL-2	122 IFN- $\alpha$ + IFN- $\alpha$ 6 MU tiw + IL-2 9 IL-2 MU sc, max 6 months 123 MPA 200mg po daily, 6 months $sc = subcutane$	122 IFN- $\alpha$ + IFN- $\alpha$ 6 MU tiw + IL-2 9 10.9* IL-2 MU sc, max 6 months	122 IFN-α + IFN-α 6 MU tiw + IL-2 9 10.9* 16.8 IL-2 MU sc, max 6 months  123 MPA 200mg po daily, 6 months 2.5* 14.9 $c = c = c = c$

qXwk = every X weekstiw = three times a week

im = intramuscularly

iv = intravenously

po = orally

MPA = medroxyprogesterone acetate

Performance status is the most consistently reported prognostic factor in mRCC. Remission is more likely in those patients with only lung metastases. However, this may be in part a result of bias relating to the different radiological methods for evaluating lesions. Brief lesions are more likely to be seen by X-ray in the lung than by scintigraphy or CT in bone, liver or other inner organ.

#### 2.4.4.2 ANTI-IFN ANTIBODIES

Antibodies have frequently been observed in patients receiving recombinant IFN-α2a, but less frequently in patients receiving IFN-α2b. On the contrary, antibodies have not been observed in patients receiving lymphoplastoid IFN. This preparation of interferon (IFN) is genetically identical to its natural form. The most frequently detected antibodies are IFN-binding and neutralizing antibodies (Prümmer 1993, Fosså *et al.* 1992b, Figlin *et al.* 1988, Buzaid *et al.* 1987, Quesada *et al.* 1985a). Prednisone may be capable of blunting the formation of anti-IFN antibodies (Ernstoff *et al.* 1990). However, prednisone has not been associated with decreased survival (Fosså *et al.* 1992b). The data suggest that anti-IFN antibodies rarely cause resistance to IFN-α.

### 2.4.4.3 DOSAGE AND SCHEDULE

The optimal dose and treatment schedule of IFN- $\alpha$  therapy has not yet been defined (Coppin *et al.* 2007). The dose of IFN- $\alpha$  has varied in different studies from 1 million units (MU) to 36 MU. Low doses of less than 5 MU have been observed to be ineffective (Quesada *et al.* 1985b, Kirkwood *et al.* 1985, Muss *et al.* 1987). A dose of Cantell-type IFN of 1 MU was compared to a dose of 10 MU, with IFN- $\alpha$  being administered intramuscularly daily. None of the patients responded to 1 MU, but 19% responded to 10 MU (Kirkwood *et al.* 1985).

Toxicity of IFN- $\alpha$  is dose-dependent. High doses of IFN- $\alpha$  result in excessive toxicity requiring dose reduction and maybe discontinuation (Krown 1987, Trump *et al.* 1987). Fosså *et al.* (1988 and 1986) observed severe adverse effects with doses of IFN- $\alpha$  ranging from 18 to 36 MU three times a week (tiw). For example, 7% of patients developed mental confusion and 4% visual disturbances due to retinal exudation. In more than half of the patients, IFN- $\alpha$  had to be reduced, delayed or discontinued due to toxic effects (Fosså *et al.* 1988, Fosså *et al.* 1986). In the study by Muss *et al.* (1987), the amount of grade 3 and 4 toxicity was greater in the group of patients who received an IFN- $\alpha$  dose from 30 to 50 MU/m² intravenously for five consecutive days every three weeks, compared to those patients with 2MU/m² subcutaneously tiw. The response rates were 10% for the former group, and 7% for the latter group. IFN- $\alpha$  administered subcutaneously or intramuscularly provides more prolonged systemic exposure than when administered intravenously. In a study by Pyrhönen *et al.* (1999), the dose of IFN- $\alpha$  was escalated from the starting dose of 3 MU tiw to 18 MU tiw. Thereafter, for patients unable to tolerate 18 MU, the dose was reduced to 9 MU tiw.

# 2.4.4.4 QUALITY OF LIFE (QoL)

IFN- $\alpha$  therapy is safe and no IFN- $\alpha$ -related deaths have been reported (Coppin *et al.* 2007), but IFN- $\alpha$  causes many adverse effects affecting the quality of life. Almost all patients experience symptoms of a flu-like syndrome, such as fever, fatigue, taste change, loss of appetite, and myalgia. As the dose of IFN- $\alpha$  increases, depression, anorexia, malaise, leucopenia, and elevation of liver function tests become common. Fatigue and depression result in reduction of physical activity in mRCC patients receiving IFN- $\alpha$ . In the study of Steineck *et al.* (1990), very high elevation of liver function tests associated with intolerable tiredness. Severe neuropsychiatric effects during IFN- $\alpha$ 

therapy, such as confused state of mind, are rare (Renault *et al.* 1987). Depression has successfully been treated with antidepressant agents. The association between toxicity and remission has not been defined.

In the study of Négrier *et al.* (2007), formal quality of life was assessed using the EORTC QLQ-C30 instrument. At three months, 16% of patients with IFN- $\alpha$  therapy had impaired quality of life, while the same was true for 11% of patients with medroxyprogesterone acetate (MPA) therapy. Reducing the dose ameliorates the symptoms. Dose reductions or discontinuations of IFN- $\alpha$  therapy are common events in various studies.

### 2.4.4.5 NEPHRECTOMY PRIOR TO IFN-α

Initial cytoreductive nephrectomy prior to planned IFN- $\alpha$  treatment improves survival and delays time to progression, despite an unimproved response rate in mRCC (Flanigan *et al.* 2001, Mickisch *et al.* 2001). Nephrectomy followed by IFN- $\alpha$  monotherapy improved one-year survival by 47%; the improvement in median survival was nearly five months (Flanigan *et al.* 2001, Mickisch *et al.* 2001). The eligibility of patients was restricted; these studies included only patients with good performance status (ECOG 0-1). The dose of IFN- $\alpha$  was 5 MU and most often IFN- $\alpha$  was started within four week after nephrectomy. The response rate was only 6% in both studies combined. The mechanism of improved survival is currently unknown. The benefit of using nephrectomy in mRCC patients prior to other forms of systemic therapy remains unproven.

# 2.4.4.6 IFN-α VERSUS OTHER THERAPIES

When comparing IFN- $\alpha$  to chemotherapy, hormonal therapy, or biological response modifiers (Motzer *et al.* 2001b), such as thalidomide (Motzer *et al.* 2002c), the effectiveness of IFN- $\alpha$  has been found to be superior or similar. However, of the modern targeted therapies, sunitinib, a VEGF receptor tyrosine kinase inhibitor, has been shown to produce a superior objective response rate and progression-free survival in the sunitinib group as compared to IFN- $\alpha$  (Motzer *et al.* 2007).

# 2.4.4.7 IFN-α PLUS OTHER THERAPIES

There have been efforts to enhance the efficacy of IFN-α by additional agents, but so far there is no evidence of enhanced survival except with bevacizumab (Rini *et a. l*2004) for increasing efficacy. In randomized studies, combining low dose IL-2 (LD-IL-2) with IFN-α increases the response rate compared to either therapy alone; the combined therapy response rate being 18.6% versus approximately 7% (Négrier *et al.* 2007, Boccardo *et al.* 1998, Négrier *et al.* 1998, Vuoristo *et al.* 1994, Vogelzang *et al.* 1993). However, similar response rates have been achieved by IFN-α alone

in other studies. Additionally, due to higher toxicity, the combination of LD-IL-2 and IFN- $\alpha$  led to a decreased quality of life in 33% of patients, compared to 16% for either therapy alone (Négrier *et al.* 2007).

The additional agents have included hormones such as toremifene and MPA (Oh *et al.* 2002, Porzsolt *et al.* 1988), chemotherapies such as vinblastine (Fosså *et al.* 1992c, Kellokumpu-Lehtinen and Nordman 1990) and 5-fluorouracil (Lopez Hänninen *et al.* 1996), other biological therapies, such as IFN-β (Mani *et al.*1995), IFN-γ (Mani *et al.*1995), and 13CRA (Aass *et al.* 2005, Motzer *et al.* 2000b). Additionally, aspirin (Creagan *et al.* 1991), prednisone (Fosså *et al.*1992b), and cimetidine with coumarin (Sagaster *et al.* 1995) for alleviating adverse effects of therapy, have been used.

# 2.4.4.8 IFN-α AS SECOND LINE THERAPY

Second-line immunotherapy with IFN- $\alpha$  or IL-2 (IFN- $\alpha$  after failure of IL-2, or IL-2 after failure of IFN- $\alpha$ ) is not effective (Escudier *et al.* 1999).

# 2.4.4.9 IFN-α AS ADJUVANT THERAPY

IFN- $\alpha$  as adjuvant therapy following nephrectomy has been investigated in patients at high risk of relapse (Messing *et al.* 2003, Pizzocaro *et al.* 2001, Prümmer 1993). An early pilot study was promising as a significant increase in NK cell activity was detected after initiation of IFN- $\alpha$  (Takahashi *et al.* 1994). However, current evidence does not support the use of IFN- $\alpha$  as adjuvant therapy in RCC (Messing *et al.* 2003, Pizzocaro *et al.* 2001), although Pizzocaro *et al.* (2001) found a protective effect in the subgroup of pN2/pN3 patients.

# 2.4.4.10 BIOLOGICAL EFFECTS OF IFN-α

There are two types of human IFNs: IFN- $\alpha$ , the leucocyte IFN, and IFN- $\beta$ , the fibroblast IFN. These are called type I IFNs, while IFN- $\gamma$ , the immune IFN, is called type II IFN (Zav'Yalov and Zav'Yalova 1997). IFN- $\alpha$  was discovered by Isaacs and Lindemann in 1957 while searching for a substance that blocked viral infection of cells.

IFN- $\alpha$  is a glycosylated polypeptide of 16-25 kDa. Fourteen genes, considered tumor-suppressor genes, on chromosome 9p21 encode 12 distinct protein forms of IFN- $\alpha$  (Pestka 1997). IFN- $\alpha$  is produced in response to enveloped viruses, virus-infected cells, bacteria, tumor cells, and double-stranded RNA (Ruszczak and Schwartz 1997). IFNs were named for their ability to mediate viral

interference, where one virus interferes with the replication of a second (Pfeffer and Donner 1990). Lymphocytes and macrophages produce IFN- $\alpha$ .

IFN- $\alpha$  is pleiotropic cytokine with immunomodulatory, antiangiogenic, proapoptotic effects, and antiviral and antiproliferative effects. IFN- $\alpha$  has direct antiproliferative effects on renal-tumor cells (Nanus *et al.* 1990, Kuebler *et al.* 1987). IFN- $\alpha$  has different cytostatic and immunomodulatory effects on mammalian cells (Pestka 1983). The exact antitumor mechanisms against RCC have not been defined.

The inhibitory effect of IFNs is not specific to the phase of the cell cycle (Kasuya *et al.* 2001). It is known that IFN- $\alpha$  enhances class I major histocompatibility complex (MHC) expression of cells. Moreover, IFN- $\alpha$  has the capacity to affect cellular differentiation.

IFN- $\alpha$  has multiple immunomodulatory effects, such as activating monocytes and macrophages, and induction of IL-2 receptors (Holan *et al.* 1991). IL-2 is produced by T and NK cells and plays a pivotal role in the regulation of lymphocyte activation and proliferation (Trinchieri *et al.* 1996). IFN- $\alpha$  acts as a positive and negative regulator of NK cells, and modifies the susceptibility of target cells to lysis (Ernstoff *et al.* 1983). IFN- $\alpha$  increases the amount of T-lymphocytes. IL-2 is presumed to activate NK cells, T-lymphocytes and lymphocyte-activated killer cells and tumor-infiltrating lymphocyte cells (Trinchieri *et al.* 1984). Phagocyte receptors are crucial for the cytotoxic function of neutrophils and macrophages (Wallace *et al.* 1994).

# 2.4.5 INTERLEUKIN-2

The development of HD-IL-2 therapy was based on animal model data (Rosenberg *et al.* 1985). HD-IL-2 appears to benefit some mRCC patients by producing durable objective responses (CRs or PRs) (Fyfe 1995). When comparing HD-IL-2 therapy to combined LD-IL2 and IFN-α, or to LD-IL-2 or IFN-α therapy alone, the response rate increases by adding another cytokine to IFN-α, but is highest with HD-IL-2 therapy. The reported response rates have typically been more than 20% with HD-IL-2, from 10% to 20% with LD-IL-2 plus IFN-α, or less than 10% with LD-IL-2 alone (McDermott *et al.* 2005, Négrier *et al.* 1998), but lower responses with HD-IL-2 have also been reported (Wagner *et al.* 1999). Remissions have not translated into a similar hierarchy for overall survival. HD-IL-2 may improve survival in the subgroup of patients with the poorest prognosis, such as those with primary tumor still in place or with either liver or bone metastases (McDermott *et al.* 2005). Of patients treated with HD-IL-2, 10-20% are estimated to live for 5 to 10 years (Fisher *et al.* 2000). However, patients for HD-IL-2 therapy should have (very) good performance status and organ function, such as cardiac function, since the treatment is toxic. Therefore, HD-IL-2

is the therapy of choice for only few patients only. Additionally, for other types of RCC than the conventional type, HD-IL-2 should not be considered, because the efficacy of cytokine therapy in other types of RCC is uncertain (Rosenberg *et al.* 1993). For mRCC, IL-2 therapy is predominantly used in the USA, while IFN-α in Europe (Decatris *et al.* 2002). HD-IL-2 therapy is not a common therapy because of its toxicity with multi-organ system effects, such as hypotension, tachyarrhythmias, capillary leak syndrome, and renal or hepatic failure (McDermott *et al.* 2005, Yang *et al.* 2003b). In the United States, HD-IL-2 therapy is usually administered by bolus, while in Europe it is administrated continuously. LD-IL-2 has been used subcutaneously (Négrier *et al.* 2007).

The additional agents connected with LD-IL-2 therapy have been lymphocyte-activated killer cells (Rosenberg *et al.* 1993), tumor-infiltrating lymphocytes (Figlin *et al.* 1999), IFN-β (Witte *et al.* 1995), histamine (Donskov *et al.* 2005) and melatonin (Lissoni *et al.* 2000), but there is no evidence has that these affect survival.

# 2.4.6 TARGETED THERAPIES

The molecular pathways with multiple targets that are of special interest in RCC are angiogenesis and intracellular signal transduction pathways.

Sunitinib, inhibitor of VEGF receptor tyrosine kinase, has been investigated in conventional RCC in a randomized study (Motzer *et al.* 2007). The response rate to sunitinib was superior compared to IFN-α, 47% and 12%, respectively; the difference was statistically significant. Median progression-free survival with sunitinib was superior to IFN-α; 11.0 months and 5.0 months, respectively; the difference was also statistically significant. Median progression-free survival was superior for those patients with good or intermediate prognosis (Motzer *et al.* 2007). Sunitinib may also be effective in papillary and chromophobe RCC (Choueiri *et al.* 2008). Administration of sunitinib in mRCC patients without nephrectomy may be feasible, and lead to a reduction in tumor size that can facilitate subsequent surgical resection (Bex *et al.* 2009, Thomas *et al.* 2009). The most often seen adverse effects with sunitinib are diarrhea, fatigue, nausea, anemia, and leucopenia. QoL was significantly improved by sunitinib when the functional assessment of the cancer therapy general scale (FACT-G) and the functional assessment of the cancer therapy-kidney symptom index (FKSI) self-assessment tools were used (Motzer *et al.* 2007). Sunitinib has shown to be potentially cost-effective as a second-line treatment for cytokine-refractory mRCC compared with the best supportive care, including palliative biochemotherapy (Purmonen *et al.* 2008).

Sorafenib, inhibitor of VEGF receptor tyrosine kinase, which can also suppress immune responses by inhibiting dendritic cell function (Hipp *et al.* 2007), was superior to placebo at a dose of 400 mg twice daily as a second-line therapy in patients with prior cytokine therapy (Escudier *et al.* 2007a). Progression-free survival for the patients given sorafenib was 5.5 months, while for placebo, it was 2.8 months. The risk of brain metastases was reduced by sorafenib; these were observed in 3% of patients with sorafenib, and in 12% of patients with placebo (Stein 2006). The adverse effects caused by sorafenib included cardiac ischemia, hypertension, hand-foot syndrome, and diarrhea. Sorafenib was found by QoL measures to ameliorate symptoms. Bone pain was found to be more common in those patients who were given placebo (Bukowski *et al.* 2007). The study was discontinued early when progression-free survival was clearly better in the sorafenib group. Sorafenib may also be effective in papillary and chromophobe RCC (Guevremont *et al.* 2009).

Temsirolimus, also called rapamycin, is metabolized to sirolimus, and is an inhibitor of an intracellular kinase called mTOR (mammalian target of rapamycin), disrupting cell cycle progression and angiogenesis. Temsirolimus is a macrolide antibiotic and immunosuppressive drug, derived from the Streptomyces species. A phase III study compared temsirolimus at a dose of 25 mg i.v. weekly to IFN-α at a dose of 3 MU tiw (Hudes et al. 2007) in patients with intermediate or poor prognosis. Progression-free survival for patients with temsirolimus was 3.8 months, while for IFN- $\alpha$  it was 1.9 months. Overall survival for patients on temsirolimus was 10.9 months, while for IFN- $\alpha$  it was 7.3 months; the difference was statistically significant. Even in patients with nonconventional RCC, overall survival was statistically significantly improved with temsirolimus compared to IFN-a (Dutcher et al. 2009). Grade 3 and 4 toxicity was more often seen in the patients on IFN-α compared to those on temsirolimus. The typical adverse effects caused by temsirolimus are anemia, thrombocytopenia, hypertriglyceridemia, hypercolesterolemia, hyperglycemia, rash, acne, and increased creatine (Bellmunt et al. 2008). There is an increased risk of stomatitis and weight loss if temsirolimus and IFN-α are administered together (Bellmunt et al. 2008).

Everolimus, an orally administered inhibitor of mTOR, has an encouraging antitumor activity against mRCC. Progression-free survival of more than or equal to six months for approximately 70% of patients, and a median overall survival of 22.1 months have been observed (Amato et al. 2009). In the patients with prior progression on sunitinib or sorafenib, median progression-free survival of four months was found in the group on everolimus compared to 1.9 months in the placebo group (Motzer et al. 2008b). Stomatitis, rash and fatigue are the typical side effects caused by everolimus (Motzer et al. 2008b). Pneumonitis has been observed in 8% of patients, and the severity of pneumonitis may be as high as grade 3 (Motzer et al. 2008b).

The addition of *bevacizumab*, an antiangiogenesis monoclonal antibody, to IFN- $\alpha$  in conventional RCC has been examined in a placebo-controlled study (Escudier *et al.* 2007b). IFN- $\alpha$  plus bevacizumab 10 mg/kg every two weeks gave a major remission rate of 31%, compared to 13% with IFN- $\alpha$  plus placebo. Median progression-free survival was 10.2 months for IFN- $\alpha$  plus bevacizumab and 5.4 for IFN- $\alpha$  plus placebo; the difference was statistically significant. The overall survival analysis showed a trend in favor of IFN- $\alpha$  plus bevacizumab. Therefore, the monitoring committee recommended that those patients with IFN- $\alpha$  plus placebo who had not progressed should change to IFN- $\alpha$  plus bevacizumab. Proteinuria and hypertension are the most often found adverse effects of this therapy, attributable mainly to bevacizumab.

# 2.4.7 OTHER THERAPIES

Nonmyeloablative allogeneic hematopoetic stem-cell transplantation can induce regression in mRCC patients who have not responded to prior IFN- $\alpha$  or IL-2 (Takahashi *et al.* 2008, Childs *et al.* 2000). Response rates with hormonal therapy, such as progestin, tamoxifen or toremifen have been approximately 10% (Gershanovich *et al.* 1996, Igel'nik *et al.* 1991, Jacqmin *et al.* 1988).

Vaccines and gene therapies are potential future therapies, particularly when the disease burden is low. On the basis of their potent antigen-presenting potential, dendritic-cell-based therapies have been investigated (Guse *et al.* 2009, Hawkins *et al.* 2009, Gitlitz *et al.* 2003). Also T-lymphocyte-based gene therapies have been developed to generate cytotoxic T lymphocytes (Mulders *et al.* 1998). WX-G250 is a chimeric monoclonal antibody that binds to carbonic anhydrase 9, which is expressed in approximately 95% of conventional RCCs (Bleumer *et al.* 2006). The assumed working mechanism of WX-G250 is by antibody-dependent cell-mediated cytotoxicity (ADCC) (Bleumer *et al.* 2006).

# 3 AIMS OF THE PRESENT STUDY

I To investigate the efficacy and tolerance of prolonged and intermittent administration of IFN- $\alpha_{2a}$  in mRCC in a phase II clinical study.

II To investigate the changes in the blood neurophils and monocyte receptor profile during IFN- $\alpha_{2a}$  administration in mRCC.

III To explore the prognostic significance of molecular markers of p53, Ki-67 and COX-2 in RCC according to the occurrence of metastases and survival.

# 4 MATERIALS AND METHODS

The patient characteristics in the different studies are presented in Table 14. No mRCC patient had biochemotherapy before IFN- $\alpha$  treatment, and after progression to IFN- $\alpha$ , no one was treated with bevacizumab or tyrosine kinase inhibitors. Histopathological samples of the studies were reevaluated: the tumors were categorized according to the Heidelberg classification (Kovacs *et al.* 1997), and re-graded according to the WHO classification (Mostofi *et al.* 1998) by an experienced pathologist (Karl-Ove Söderström). For T-staging categorization, the 2002 updated UICC pTNM classification system of renal carcinomas was used (Sobin and Wittekind 2002). Table 15 shows the follow-up of the patients in the studies.

Table 14. Characteristics in different studies.

Characteristics	No. (%)	No. (%)	No. (%)	No. (%)	No. (%)
Publication	I	II	III	IV	V
Total no. of RCC pts	75 (100)	117 (100)	18 (100)	117 (100)	102 (100)
Sex					
Female	29 (39)	47 (40)	6	54 (46)	44 (43)
Male	46 (61)	70 (60)	12	63 (54)	58 (57)
Age					
years, median [range]	63 [43-77]	63 [38-78]	64 [52-77]	61 [37-82]	61 [37-79]
Time to metastases in M0-pts					
months, median [range]	1 [0-156]	0 [0-157]	22 [0-82]		
WHO performance status					
< 2	57 (76)	80 (68)			89 (87)
= 2	18 (24)	37 (32)			13 (13)
T-stage					
T1		20 (20) 1		42 (37)	36 (36)
T2		19 (19) 1		29 (26)	26 (26)
T3		50 (50) 1		35 (31)	31 (31)
T4		12 (12) <sup>1</sup>		7 (6)	7 (7)
Tumor grade					
G1		38 (37) 2		34 (29)	27 (26)
G2		39 (38) <sup>2</sup>		57 (49)	51 (50)
G3		25 (25) <sup>2</sup>		26 (22)	24 (24)
No. of metastases					
M0	0 (0)	0 (0)		51 (44)	45 (44)
M1	75 (100)	117 (100)		66 (56)	57 (56)
Site of metastases					
lung	52 (69)		3, other 5		
skin or soft tissue	28 (37)				
bone	26 (35)				
lymph node	19 (25)				
liver	11 (15)				
pleural	10 (13)				
Heidelberg classification type					
conventional				101 (86)	86 (84)
papillary				4 (3)	4 (4)
chromophobe				6 (5)	6 (6)
unclassified				6 (6)	6 (6)
Prior therapy					
Nephrectomy	70 (93)	107 (91)	17	117 (100)	102 (100)
Radiotherapy to kidney	1 (1)	2 (2)			
Response to IFN-α					
complete response	5 (6.6) <sup>3</sup>	9 (7.7)4			
partial response	8 (10.7) <sup>3</sup>	11 (9.4)			
stable disease	32 (42.7) <sup>3</sup>	49 (41.9) 4			
progressive disease	27 (36.0) <sup>3</sup>	48 (41.0) <sup>4</sup>			

<sup>&</sup>lt;sup>1</sup>Sixteen patients were not evaluable for T-stage.
<sup>2</sup>Fifteen patients were not evaluable for tumor grade.
<sup>3</sup>Three patients were non-evaluable as IFN-α was discontinued before 4 weeks
<sup>4</sup>An intention-to-treat analysis.

Table 15. Follow-up of RCC patients after nephrectomy and mRCC patients after the beginning of IFN-α. Staging RCC was based on the American Joint Committee on Cancer staging (Miller *et al.* 1981).

Staging investigation:	Follow-up interval in RCC	Follow-up interval in mRCC
Follow-up evaluation: symptoms of the disease signs of the disease performance status neurological status body weight	every 6 months	every 1-4 weeks, thereafter bimonthly
response to IFN-α	N/A	
Laboratory evaluation: blood counts serum concentrations of calcium liver enzymes creatinine	every 6 months	every 1-4 weeks, thereafter bimonthly
Radiological evaluation:		
chest X-ray and abdominal US /	every 6 months if signs of disease manifestation	alternative evaluation method, every 3-4 months
bone scintigraphy	if bone pain	if bone pain

# 4.1 EFFICACY AND TOLERABILITY OF PROLONGED ADMINISTRATION OF IFN- $\alpha_{2a}$ IN mRCC IN PHASE II CLINICAL STUDIES. (I, II)

# 4.1.1 PATIENT ELIGIBILITY

Between December 1994 and December 2002, 117 patients with mRCC gave informed consent to participate in the recombinant IFN- $\alpha$ 2a (Roferon®) study. The inclusion criteria were: age younger than 80, performance status of 0-2 (WHO), serum creatinine concentration of less than 200 umol/l, and serum concentrations of liver enzymes less than twice the upper reference limit. Patients with brain metastases, other malignancies, and severe concomitant disease, or with a life expectancy of less than three months were excluded.

# 4.1.2 TREATMENT SCHEDULE

Patients were treated with subcutaneous IFN- $\alpha$  three times a week. The dose was increased weekly from 4.5 MU to the maximal tolerable dose of 9, 12, 13.5 or 18 MU. The maximum dose was to be reached during the first four weeks. The maintenance dose was chosen on the basis of the patient's tolerance of adverse effects. The treatment cycle consisted of three weeks' treatment with IFN- $\alpha$  followed by a one-week pause. IFN- $\alpha$  was planned to be continued until progression of over 25% in tumor measurements (PD), substantial adverse effects, or for up to two years. Adverse effects were recorded according to ECOG toxicity criteria (Oken *et al.* 1982).

IFN- $\alpha$  was self-administered on an outpatient basis. In addition to the injection of IFN- $\alpha$ , patients were recommended to use naproxen prophylactics to ameliorate flu-like symptoms. The treatment protocol was designed by Professor E. Salminen. The patients were treated by IFN- $\alpha$  until December 2002, and followed at the Department of Oncology and Radiotherapy of Turku University Hospital, until December 2008.

# 4.1.3 STAGING AND RESPONSE ASSESSMENT

The evaluation of the response followed the criteria of the World Health Organization (WHO) (Table 16). For the patient to qualify as a responder, two successive measurements at least four weeks apart confirming response (CR or PR) were required. A PR in bone metastases was defined as clinically stable lesions with a decrease in uptake in skeletal scintigram and normalizing of alkaline phosphatase. The response duration was measured from the first observed response.

Table 16. Evaluation of the response following the criteria of the World Health Organization (Miller *et al.* 1981).

Type of response	Abbreviation	Description of the response
complete response	CR	Disappearance of all known disease, determined by two
		observations not less than four weeks apart
partial response	PR	A 50% or more decrease in total tumor load of the lesions that have been measured to determine the effect of therapy by two observations not less than four weeks apart. Bi-dimensional: single lesion, greater than or equal to 50% decrease in tumor area (multiplication of longest diameter by the greatest perpendicular diameter)
stable disease	SD	A 50% decrease in total tumor size cannot be established nor has a 25% increase in the size of one or more measurable lesions been demonstrated.
progressive disease	PD	A 25% or more increase in the size of one or more measurable lesions or the appearance of new lesions.

# 4.1.4 STATISTICS

The association of prognostic factors with survival time or time to progression was analyzed using survival analysis applying Kaplan-Meier estimation, log-rank test and Cox's regression analysis. The associations between separate prognostic factors were analyzed with cross-tabulation and Pearson's chi-square test. p-values less than 0.05 were interpreted as statistically significant. Computing was performed with SAS System version 9.2 (2002, SAS Institute Inc., Cary, NC, USA).

# 4.2 CHANGES IN BLOOD NEUTROPHIL AND MONOCYTE RECEPTOR PROFILE DURING IFN- $\alpha_{2A}$ ADMINISTRATION IN mRCC (III)

Eighteen mRCC patients, aged 52 to 77 years, receiving IFN- $\alpha$  were included in the study. Their phagocyte receptor expressions (Fc $\gamma$ RI, Fc $\gamma$ RII and Fc $\gamma$ RIII) and complement receptors (CR1 and

CR3) were studied in neutrophils and monocytes. No previous radio-, chemo-, or hormonotherapy had been given. The initial dose of IFN- $\alpha$  was 4.5 MU s.c. tiw, followed by escalation up to 9.0-13.5 MU tiw. This dose was continued intermittently with a three-week-on one-week-off schedule. As controls, samples from 39 healthy subjects were studied.

Samples from three patients were collected after the maintenance level of IFN- $\alpha$  was reached, during the treatment weeks prior to and after the rest week, and during the rest week, in order to study receptor expression during the treatment cycle, and evaluate the significance of the sampling time.

# 4.2.1 REAGENTS

Hanks balanced salt solution without Ca<sup>2+</sup> and Mg<sup>2+</sup> ions (CMF-HBSS, pH 7.4) was prepared and supplemented with 0.1% gelatine. FITC-conjugated anti FcγRI (CD64), anti-FcγRII (CD16), anti-CR1 (CD35) and mouse IgGl isotype control antibody and PE conjugated anti-FcγRII (CD23), anti-CR3 (CDIIb), mouse IgGl isotype antibody and mouse IgG2a isotype antibody were purchased from Immunotech (Marsseille, France).

# 4.2.2 COLLECTION AND PREPARATION OF SAMPLES

EDTA-anticoagulated (1.5mg EDTA/ml, blood) blood samples were collected from the 18 study subjects prior to and during the first two months of treatment. After the maintenance level was reached, samples were collected from three patients during the treatment cycle. Blood erythrocytes were lysed with 0.83% ammonium chloride (1.5 ml blood, 8.5 ml ammonium chloride) at +20 °C for 15 minutes. After centrifugation (400 x g for 10 minutes at +4 °C), the leukocytes were resuspended in 500 μl ice-cold CMF-HBSS. The leukocyte count was determined with a Coulter counter model S blood cell analyzer (Coulter Electronics Inc., Hialeah, FL, USA).

# 4.2.3 MEASUREMENTS OF RECEPTOR EXPRESSION

Leukocytes (3x10<sup>5</sup>) were incubated with monoclonal antibodies (mAb) in 12x75 mm polystyrene vials for 30 minutes at +4°C. The incubation volume was 90 µl. The control sample was incubated with isotype-matched mAbs directed to an irrelevant antigen. After incubation, the cells were washed with cold CMF-HBSS. Leukocytes were re-suspended in 500 µl cold CMF-HBSS and analysed in a Coulter EPICS (Coulter, Miami, Florida, USA) flow cytometer. The fluorescence of 5000 cells was measured using logarithmic amplification. A relative measure of receptor

expression was obtained by determining the mean log fluorescence intensity. The percentage of positive cells was generally 98-100 unless otherwise indicated.

# 4.2.4 IN VITRO EXPERIMENTS

Heparin anticoagulated blood samples, containing 0, 2.25, 11, or 55 ng/ml interferon- $\alpha$  were incubated at 37°C for 60 minutes. Whole blood lacing interferon- $\alpha$  served as control. After incubation, the red blood cells were lysed with 0.83% ammonium chloride, and the receptor expression was measured.

### 4.2.5 STATISTICS

Analysis of variance for repeated measurements was used for comparing differences between groups at different time points (SAS 6.12 version, proc mixed). Pair-wise comparison was tested by the Tukey HSD test with confidence limits between means. Confidence intervals for group means were calculated using the Scheffe method. The two-sample t-test was used for comparison of controls to treated patients. Values obtained in different treatment phases were analysed with one-way ANOVA. The level of significance was p<0.05.

# 4.3 PROGNOSTIC SIGNIFICANCE OF MOLECULAR MARKERS OF p53, Ki-67 AND COX-2 IN RCC ACCORDING TO OCCURRENCE OF METASTASES (IV, V)

# 4.3.1 PATIENTS, STAGING, AND HISTOLOGY

The study included samples from 117 patients with local or metastatic RCC treated in Turku University Hospital. Consecutive samples were collected from patients with local RCC treated between 1986 and 1996, with metastatic RCC between 1995 and 2001. As the patients with local disease had been free of metastases for at least 7.5 years, their samples were from an earlier period than the samples from patients with metastatic disease. The patients of the study were divided into three categories according to the occurrence of metastases. The patients with M0 staging (pTNM classification system) at primary diagnosis were divided into two subcategories: the first group, no metastases (nm), i.e. those patients whose RCC had no metastases within the follow-up of 7.5 years, and the second group, late metastases (lm), i.e. those patients whose RCC developed later metastases (= metachronous metastases) after the primary diagnosis. The third group, primary metastases (pm), was formed by those RCC patients who were M1 patients at primary diagnosis (pTNM classification system), (= synchronous metastases).

# 4.3.2 IMMUNOHISTOCHEMICAL STAINING AND SCORING OF p53, Ki-67 AND COX-2

From archival paraffin-embedded blocks, containing well-preserved cancer tissue, 5-µm-thick sections were cut, deparaffinized with xylene, and rehydrated through a graded series of alcohol. For antigen retrieval, the samples were boiled for 10 minutes in a microwave oven in 10mM sodium citrate buffer (pH 6.0). An automated processor (TechMate 500, DAKO) was used for immunohistochemical staining. Steps were performed in the immunostainer using the avidin-biotin-peroxidase staining methods.

Table 17. Evaluation of immunoreactivity of p53, Ki-67, and COX-2.

Monoclonal antibody	Antigen to detect	Antibody dilution	Counting of immunoreactivity	Classification of staining in the sample
DO-7 (Dako, Denmark)	p53	1:300	Percentage of carcinoma cells exhibiting p53 nuclei staining	As continuous data from undetectable levels (0%) to homogeneous (100%).
MIB-1 (Dako, Denmark)	Ki-67	1:100	Percentage of carcinoma cells exhibiting Ki-67 nuclei staining	As continuous data from undetectable levels (0%) to homogeneous (100%).
COX-2 (Dako, Denmark)	COX-2	1:100	a) Percentage of carcinoma cells exhibiting COX-2 nuclei staining b) Degree of intensity (absent/weak, pale, strong)	As three classes: 0 (no), absent/weak intensity in less than 10% of cancer cells; 1 (low), pale intensity in 10% or over of cancer cells; and 2 (high), strong intensity in majority of cancer cells.

The samples were incubated with commercial monoclonal antibody at optimal dilution, for 27 minutes as shown in Table 17, after which they were visualized by avidin-biotin-peroxidase staining. The immunoreactivity of p53, Ki-67, and COX-2 was counted for each tumor slide. Also, peritumoral inflammation as immunologic effect was analyzed: the amount of peritumoral lymphocytes was counted as 0 = no; 1 = mild; 2 = moderate or 3 = severe (Tuna *et al.* 2004).

The 10% cut-off value was selected to achieve statistically reliable results, as well as in accordance with a previous study on the subject (Olumi *et al.* 2001). Staining without the primary antibody served as negative control. No significant background staining was detectable. The reliability of staining was measured by standard positive controls used as weekly standard controls in the routine pathological laboratory.

Immunohistochemistry of p53, Ki-67, and COX-2 was scored as a consensus of two investigators (Minna Kankuri-Tammilehto, Karl-Ove Söderström). The pathologist (Karl-Ove Söderström) also analyzed the reaction of peritumoral inflammation.

# 4.3.3 STATISTICAL ANALYSIS

Univariate associations between the variables were evaluated using contingency tables and  $\chi^2$  or Fisher's exact test. When metastatic group, tumor size, and grade were the dependent variables, the univariate and multivariate associations of dependent variables and the prognostic factors, p53 and Ki-67, were analyzed using logistic regression analysis (Hosmer *et al.* 2000). As the ordinal-type dependent variables consisted of more than two categories, the cumulative logistic models (proportional odds model) were used instead of the traditional binary logistic regression analysis. The results of logistic regression were quantified by calculating odds ratios (OR) and cumulative odds ratios (COR) with 95% confidence intervals (95% CI). Kaplan-Meier survival curves for p53 and Ki-67 were calculated for patients with primary metastases (pm) or later metastatic disease (lm), and the curves were compared using the Log-Rank test. Additionally, for the patients with later metastatic disease (lm), prognostic value of the clinicopathological variables and biomarkers, p53 and Ki-67, for metastases-free and overall survival were analyzed using the Cox proportional hazards model. In all tests, p-values less than 0.05 were considered statistically significant. Statistical calculations were performed using SAS System for Windows, release 8.02/2001 (SAS Institute, Cary, NC).

The associations between patient characteristics and COX-2 expression were evaluated using cross-tabulation,  $\chi^2$  or Fisher's exact test, and one-way analysis of variance (age). The Cox proportional hazards model was used to analyze the association of the clinicopathological variables, the biomarkers, p53 and Ki-67, and COX-2 expression, with metastases-free survival and overall survival. Variables significantly associated with survival in univariate Cox models were included in the multivariate Cox model. The results were quantified by calculating hazard ratios with 95% confidence intervals. Survival curves were calculated using the Kaplan-Meier method. The differences between the curves were tested with the log-rank test.

The association of COX-2, p53 and Ki-67 with the metastatic group was analyzed using univariate and multivariate multinomial logistic regression. The results were quantified by calculating odds ratios with 95% confidence intervals. In all tests, p-values less than 0.05 were considered statistically significant. Statistical calculations were performed using SAS System for Windows, release 8.02/2001 (SAS Institute, Cary, NC).

# 5 RESULTS

# 5.1 EFFICACY AND TOLERANCE OF PROLONGED ADMINISTRATION OF IFN- $\alpha_{2a}$ IN mRCC IN PHASE II CLINICAL STUDIES. (I, II)

At the time of analysis the median progression-free survival was eight months, and the median overall survival 19.1 months. The five-year survival rate was 16%. There were statistically significant differences in survival by response to treatment (log-rank test, p<0.001, Figure 2). In four (20%) of 20 responders the IFN- $\alpha$  dose was under 9 MU 3 tiw. The dose for the remaining 16 (80%) responders was between 9 and 18 MU 3 tiw. The mean duration of treatment with interferon- $\alpha$  was 11 months (range 2 weeks to 32 months).

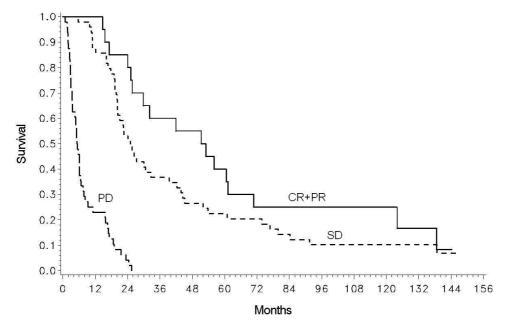


Figure 2. Kaplan–Meier survival curves for responding patients, patients with stable and progressive disease (log-rank P< 0.001). CR: complete response; PR: partial response; SD: stable disease; PD: progressive disease.

The patients were classified according to the Cleveland Clinic Foundation scoring system into three risk groups (Choueiri *et al.* 2007). The median overall survival and five-year survival proportions, as well as the number of objective responses (CR + PR) in each risk group are shown in Table 18. Figure 3 shows the Kaplan-Meier survival curves for the three risk groups.

Table 18 Survival for risk grou	ins according to Cleveland	Clinic Foundation scoring system.

Risk group	Number of poor prognostic factors	No. of patients	No. of objective responses (CR+PR)	Proportion of five-year survivors (%) <sup>1</sup>	Median overall survival (mos) [range]
Low	<2	60 (52%)	13 (22%)	23	24.3 [19.1 to 32.7]
Intermediate	=2	33 (29%)	6 (18%)	15	14.7 [5.6 to 22.3]
High	>2	22 (19%)	1 (5%)	0	6.2 [3.0 to 9.4]

<sup>1</sup>p-value <0.001

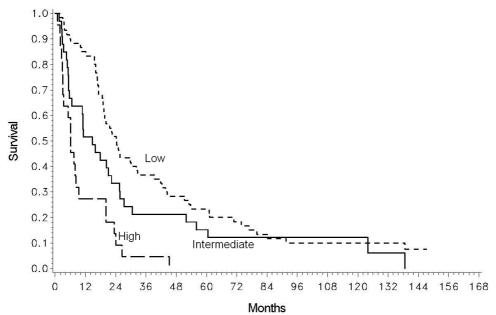


Figure 3. Kaplan–Meier survival curves for risk groups according to the Cleveland Clinic Foundation scoring system (log-rank p<0.001).

# 5.2 CHANGES IN BLOOD NEUTROPHIL AND MONOCYTE RECEPTOR PROFILE DURING IFN- $\alpha_{2A}$ ADMINISTRATION IN MRCC (III)

The phagocyte receptor expression in kidney cancer patients (n=18) compared with normal controls (n=39) is presented in Table 19 (neutrophils) and Table 20 (monocytes). The mean fluorescence intensity of neutrophils labeled with anti-CR3 mAbs was significantly higher in renal cancer patients than in controls (p=0.002). Furthermore, the proportion of Fc $\gamma$ RI positive neutrophils was significantly raised in patients with renal cancer when compared to controls (p=0.0003). No alterations were observed in CR1, Fc $\gamma$ RII or Fc $\gamma$ RIII levels between healthy controls and cancer patients.

In the monocytes of the cancer patients, mean fluorosence intensities of all phagocyte receptors, except Fc $\gamma$ RIII, were raised when compared to controls (Table 20). Furthermore, the proportion of Fc $\gamma$ RIII positive monocytes was significantly increased (Table 20). No significant changes were observed in the proportion of monocytes expressing other phagocyte receptors.

Table 19. Mean fluorescence intensity of Fcγ- and complement receptor expression in neutrophils.

	Controls		Pati	Patients		
Neutrophils	Mean	Std	Mean	std	p-value	
CR1	8.04	3.35	7.04	2.49	0.2656	
CR3	8.34	4.67	14.87	7.34	0.0020	
FcγRI %	28.87	20.46	52.37	23.06	0.0003	
FcγRI	1.60	0.37	1.61	0.30	0.9468	
FcγRII	6.80	3.08	7.98	3.29	0.1966	
FcyRIII	117.26	32.70	113.53	35.30	0.6983	

Healthy control subjects, n=39

Patients with mRCC prior to IFN-α therapy, n=18

Table 20. Mean fluorescence intensity of Fcy- and complement receptor expression in monocytes.

	Controls		Patio	_	
Monocytes	Mean	std	Mean	std	p-value
CR1	8.48	2.86	10.34	3.49	0.0379
CR3	8.39	4.49	21.12	11.30	0.0002
FcγRI	8.24	1.81	10.06	3.56	0.0534
FcγRII	7.17	3.23	11.22	4.27	0.0002
FcγRIII	4.21	2.52	4.10	1.74	0.8635
FcγRIII %	42.06	16.90	54.43	18.80	0.0166

Healthy control subjects, n=39

Patients with mRCC prior to IFN-α therapy, n=18

To investigate the effects of IFN- $\alpha$  on receptor expression, samples during treatment and recovery phases were collected for analysis. In neutrophils, the expression of CR1 receptor decreased significantly during the treatment week (p=0.0114, baseline vs. treatment), whereas the others remained fairly constant. In monocytes, the receptor level of Fc $\gamma$ RII increased significantly during treatment (p=0.0027, baseline vs. treatment), whereas in CR1, Fc $\gamma$ RII and Fc $\gamma$ RIII expression, a transient and statistically non-significant change was observed. The results are presented in Table 21 for neutrophils and in Table 22 for monocytes.

Table 21. Mean fluorescence intensity of Fcy- and complement receptor expression in neutrophils.

	Before treatment					Reco		
Neutrophils	Mean	std	Mean	std	Mean	std	p-value	
CR1	7.04	2.49	4.79	3.47	5.75	2.16	0.0263*	
CR3	14.87	7.34	15.12	6.77	11.09	3.58	0.2154	
FcγRI %	52.37	23.06	52.55	28.7	52.15	28.7	0.8315	
FcγRI	1.61	0.30	1.66	0.27	1.60	0.41	0.8324	
FcγRII	7.98	3.29	7.23	3.05	6.55	2.69	0.1750	
FcγRIII	113.53	35.30	114.7	27.20	102.8	37.6	0.6524	

<sup>\*=</sup> CR1, baseline vs. treatment p=0.0114

Patients with mRCC prior to IFN-α therapy and recovery, n=18

Table 22. Mean fluorescence intensity of Fcy- and complement receptor expression in monocytes.

	Before treatment		Treat	ment	Reco		
Monocytes	Mean	std	Mean	std	Mean	std	p-value
CR1	10.34	3.49	8.29	4.76	9.76	4.27	0.2031
CR3	21.12	11.30	27.11	15.00	15.13	8.38	0.0959
FcγRI	10.06	3.56	13.98	5.07	10.85	3.19	0.0102*
FcγRII	11.22	4.27	10.71	2.96	8.99	2.74	0.2332
FcγRIII	4.10	1.74	4.16	1.57	4.31	1.29	0.9044
FcγRIII %	54.43	18.80	54.40	19.60	50.51	13.7	0.3430

<sup>\* =</sup> FcγRI, baseline vs. treatment p=0.0027

Patients with mRCC prior to IFN-α therapy and recovery, n=18

# 5.3 PROGNOSTIC SIGNIFICANCE OF MOLECULAR MARKERS OF p53, Ki-67 AND COX-2 IN RCC ACCORDING TO OCCURRENCE OF METASTASES (IV, V)

Both p53 and Ki-67 were markers for overall survival in the pm/lm patients (Figure 4 and Figure 5, respectively). The median overall survival was 24 months in p53-positive, and 59 months in p53-negative patients (Log-rank test, p=0.030), and 24 months in Ki-67-positive, and 63 months in Ki-67 negative patients (Log-rank test, p=0.031).

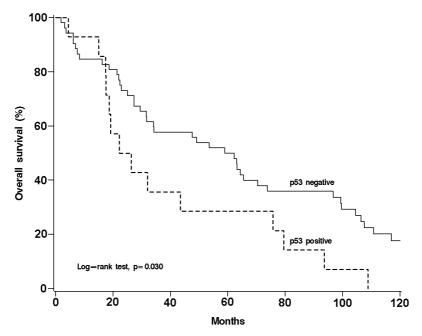


Figure 4. Kaplan-Meier survival curve for p53 in mRCC (n=66).

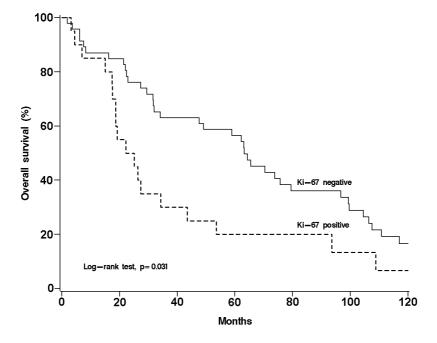


Figure 5. Kaplan-Meier survival curve for Ki-67 in mRCC (n=66).

In mRCC, COX-2 negativity/Ki-67 positivity was associated with shorter overall survival from nephrectomy when compared to COX-2 positivity/Ki-67 negativity (median overall survival time 19 vs. 97 months) (HR=3.5, 95% CI 1.5-8.1, p=0.004) (Figure 6, Log-rank test). Additionally, double positivity (HR=0.4, 95% CI 0.1-1.1, p=0.083) or double negativity (HR=0.5, 95% CI 0.3-1.0, p=0.061) of COX-2/Ki-67 was associated with longer overall survival when compared to COX-2 negativity/Ki-67 positivity (Figure 6, Log-rank test).

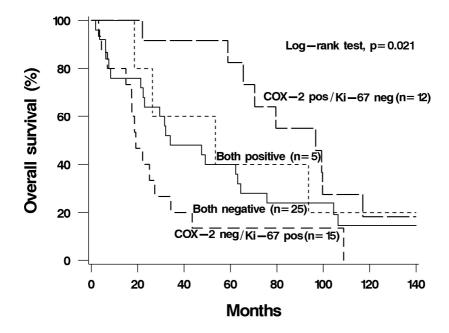


Figure 6. The prognostic value of covariation of COX-2/Ki-67 for overall survival from nephrectomy in RCC patients with metastases (either primary presentation or later) (n=57, Kaplan-Meier method): the median overall survival time was 97 months with COX-2 positivity/Ki-67 negativity, and 19 months with COX-2 negativity/Ki-67 positivity (p=0.004).

The prognostic value of variables for overall survival in the pm/lm group of patients is presented in Table 23 (Cox regression analysis). COX-2 negativity was associated with shorter overall survival, when compared to COX-2 positivity (median overall survival time 28 vs. 94 months)(p=0.027). The higher the T-stage (T3,T4), the shorter was the overall survival (p=0.012); patients with high T-stage (T3,T4) had double the risk of death compared to patients with low T-stage (T1,T2). Patients with grade 3 tumors had a 2.5 times higher risk of death when compared to patients with grade 1 tumors (p=0.039). p53 and Ki-67 negativity showed a trend toward longer overall survival (p=0.063 and p=0.068, respectively). In multivariate analysis, only T-stage was an independent variable for overall survival.

The median metastases-free survival was shorter with COX-2 negative tumors when compared to those with COX-2 positive tumors (15 vs. 46 months)(HR=2.5, 95% CI 1.1 - 5.3, p=0.020, Logrank test) (Figure 7).

Table 23. Prognostic value of variables for overall survival in RCC patients with metastatic disease (n=57).

Variable	<u>Univariate</u> <u>analysis</u> p-value	Hazard ratio	95% confidence interval	Multivariate analysis <sup>4</sup> p-value	Hazard ratio	95% confidence interval
T-stage (3+4 vs. 1+2) <sup>1</sup>	0.012	2.3	1.2 to 4.4	0.048	2.0	1.0 to 3.9
Tumor grade <sup>2</sup>	$0.109^2$					
2 vs. 1	0.277	1.5	0.7 to 3.3	0.582	1.3	0.6 to 2.9
3 vs. 1	0.039	2.5	1.0 to 6.0	0.264	1.7	0.7 to 4.1
COX-2 (negative vs. positive)	0.027	2.1	1.1 to 3.9	0.105	1.8	0.9 to 3.6
p53 (positive vs. negative)	0.063	1.8	1.0 to 3.4			
Ki-67 (positive vs. negative)	0.068	1.7	1.0 to 3.1			
Sex (female vs. male)	0.119	1.6	0.9 to 2.8			
Age at nephrectomy (< 65 yrs. vs. others)	0.417	1.3	0.7 to 2.3			
Heidelberg <sup>3</sup> (conventional vs. other types)	0.296	1.4	0.7 to 2.8			

Analyzed using Cox regression analysis. ns=not significant. Analyzed from nephrectomy.

<sup>&</sup>lt;sup>4</sup>The variables that were positive in univariate analysis were analyzed in multivariate analysis.

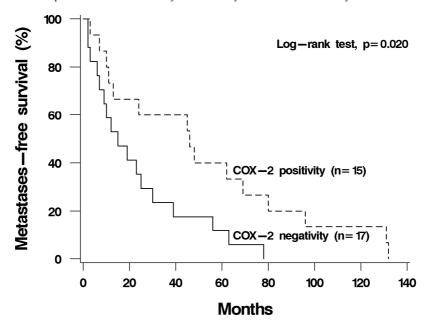


Figure 7. The prognostic value of COX-2 for metastases-free survival from nephrectomy in RCC patients who later developed metastatic disease (n=32, Kaplan-Meier method): the median metastases-free survival time was 46 months with COX-2 positivity, and 15 months with COX-2 negativity.

One patient was not evaluable for T-stage.

<sup>&</sup>lt;sup>2</sup>Overall p-value for difference between tumor grades.

<sup>&</sup>lt;sup>3</sup>According to Heidelberg classification

The prognostic value of variables for metastases-free survival in the lm group of patients is presented in Table 24 (Cox regression analysis). COX-2 was the only variable to have prognostic value for metastases-free survival.

Table 24. Prognostic value of variables for metastases-free survival in RCC (Im patients, n=32).

Variable	<u>Univariate</u> analysis p-value	Hazard ratio	95% confidence interval
T-stage (3+4 vs. 1+2) <sup>1</sup>	0.089	1.9	0.9 to 4.1
Tumor grade <sup>2</sup>	0.218		
2 vs. 1	0.886	0.9	0.4 to 2.1
3 vs. 1	0.101	2.6	0.8 to 8.3
COX-2 (negative vs. positive)	0.024	2.5	1.1 to 5.3
p53 (positive vs. negative)	0.684	1.2	0.5 to 3.2
Ki-67 (positive vs. negative)	0.196	1.8	0.7 to 4.2
Sex (male vs. female)	0.823	1.1	0.5 to 2.2
Age at nephrectomy (65 or older vs. < 65 yrs)	0.668	1.2	0.6 to 2.5
Heidelberg <sup>3</sup> (Other types vs. conventional)	0.989	1.0	0.3 to 2.9

Analyzed using Cox regression analysis. Analyzed from nephrectomy. One patient was not evaluable for T-stage.

<sup>&</sup>lt;sup>2</sup>Overall p-value for difference between tumor grades.

<sup>&</sup>lt;sup>3</sup>According to Heidelberg classification

# 6 DISCUSSION

# 6.1 IFN-α IN mRCC

# 6.1.1 RESPONSE TO AND LONG-TERM SURVIVAL WITH PROLONGED USE OF IFN-α THERAPY

Randomized trials have indicated that IFN- $\alpha$  increases the survival of RCC patients (MRCRCC 1999, Pyrhönen *et al.* 1999). Therefore, before the era of novel targeted therapies, IFN- $\alpha$  was considered as the standard comparator for first-line therapy of mRCC. The current questions concerning the use of biological response modifiers, such as IFN- $\alpha$ , are the timing (Motzer *et al.* 2007, Stadler *et al.* 2007) and the optimal dosage of IFN- $\alpha$ , alone or in combination with targeted therapies (Bracarda *et al.* 2007).

One of the aims of this study was to investigate whether prolonged IFN- $\alpha$  therapy with intermittent administration is feasible and effective. In most previous studies, the IFN- $\alpha$  treatment duration has been 6 to 24 weeks (Négrier *et al.* 2007, Atzpodien *et al.* 2002, MRCRCC 1999), and most responses have been observed within 8 - 24 weeks (Négrier *et al.* 2007, MRCRCC 1999). Since host immune mechanisms are apparently important in regulating tumor growth and in patients responding to IFN- $\alpha$  (Motzer *et al.* 1996, Marcus *et al.* 1993, Rosenberg *et al.* 1993, Oliver *et al.* 1989), a prolonged schedule of IFN- $\alpha$  administration was used in the present study.

The patients in the current study achieved median overall survival of 19.1 months, which was longer compared to the weighted average median survival of 11.4 months in a meta-analysis of 644 mRCC patients treated with IFN- $\alpha$  (Coppin *et al.* 2007). In the study of MRCRCC (1999), when comparing IFN- $\alpha$  for 12 weeks versus medroxyprogesterone acetate, the median survival time was 8.5 months in the IFN- $\alpha$  group and 6 months in the other group. In the study of Négrier *et al.* (2007), the median overall survival was 15.2 months for the IFN- $\alpha$  group, and 16.8 months for the IFN- $\alpha$  plus IL-2 group, with treatment duration of 24 weeks. Pyrhönen *et al.* (1999) observed median overall survival of 16.9 months for IFN- $\alpha$  plus vinblastine following a median treatment duration of 6.0 months. The results of the current study show that the overall survival was even longer despite the treatment cycle pause, compared to other reports on continuous IFN- $\alpha$  therapy with or without additional agents.

Previously reported five-year long-term survival of patients with IFN- $\alpha$  based therapy has been approximately between 9% and 16% (Atzpodien *et al.* 2002, Marincola *et al.* 1995, Fosså *et al.* 1992c). In all these studies, additional agents have been used, e.g. IL-2 (Marincola *et al.* 1995), 5-

fluorouracil (Atzpodien *et al.* 2002), vinblastine (Fosså *et al.* 1992c), and 13-cis-retinoid acid (Atzpodien *et al.* 2002). Only few reports of long-term outcome with IFN- $\alpha$  therapy alone in mRCC have been published. In the study of Minasian *et al.* (1993), a five-year survival rate of only 3% was reported with IFN- $\alpha$  monotherapy. In that study, the planned treatment duration was three months in one trial and until progression of disease in the other trials; the median treatment duration was not reported. The current long-term outcome study (an intention-to-treat analysis) shows that 16% of mRCC patients with prolonged IFN- $\alpha$  therapy achieved five-year survival.

Stratifying the patients into different prognostic groups, e.g. according to the Cleveland Clinic Foundation scoring system (Choueiri et al. 2007), enables better comparison of the results in different studies. The results of the current study demonstrate that approximately one out of four patients in the low risk group achieved five-year survival, and even in the intermediate risk group, approximately one in seven patients survives for at least five years. With prolonged IFN- $\alpha$ , the median overall survival in the low risk group was 24.3 months. Even in the intermediate risk group it was better, 14.7 months, than the weighted average median survival of 11.4 months in the previously mentioned meta-analysis in mRCC patients (Coppin et al. 2007). The benefit of IFN-α in mRCC patients with intermediate risk has been discussed recently. In the study of Négrier et al (2007), the survival benefit was not observed in patients with intermediate prognostic factors, but Pyrhönen et al. (1999) observed in their study (IFN-α plus vinblastine versus vinblastine) that the survival benefit of IFN-α may be greater in those patients with adverse prognostic factors, such as poor performance status, age over 60 years, and male gender. In the present study, the considerably large number of patients with intermediate risk and, additionally, the rather good median overall survival, may indicate that the intermediate risk group patients would also benefit from prolonged IFN- $\alpha$  therapy.

The objective response rate of 17% in the current study is comparable to rates in other studies with IFN- $\alpha$  alone (Négrier *et al.* 2007, MRCRCC 1999), or IFN- $\alpha$  with IL-2 (Négrier *et al.* 2007), and IFN- $\alpha$  with vinblastine (Pyrhönen *et al.* 1999). In the present study, patients with stabilized disease (42%) evidently benefitted from the prolonged treatment. The survival of patients with stabilized disease was close to the survival of responding patients. Those patients who achieve objective responses (CR + PR) reach ten times better median overall survival compared to those with progressive disease. The complete responders had metastases mainly in the lungs (7 out of 9 complete responders). In addition, one of the complete responders had bone metastases, which disappeared as observed by bone scintigraphy, and a decrease in alkaline phosphatase to normal levels. One responder had liver metastases, which disappeared as observed by CT. It has been suggested that responses to IFN- $\alpha$  rarely last more than two years (Wirth 1993). The data from this

study support the fact that durable responses to IFN- $\alpha$  are possible, and that a response duration for CR patients of a median of four years, and even more than 11 years, is possible to achieve. In the current study, four patients (3%) achieved partial response after 12 months of treatment, one of them at 17 months after the onset of therapy. One of these patients later achieved a complete response at 22 months after the onset of therapy. In one patient, the metastases seemed at first to progress slightly at 12 months and then decreased in size. This observation of late responses is a new clinical finding. This is important since response to IFN- $\alpha$  is a significant prognosticator for overall survival in mRCC patients. The observation of late responses in the present study indicates that prolonged IFN- $\alpha$  may be beneficial, as the overall survival and five-year survival data indicate.

Median progression-free survival was eight months with the present IFN- $\alpha$  treatment. After the progression during IFN- $\alpha$  therapy, no patients were treated with bevacizumab or tyrosine kinase inhibitors. Two of the patients received HD-IL-2 therapy after the progression without success. In addition, two patients were treated with capecitabine without response. Therefore, the achieved five-year survival rate is specifically due to IFN- $\alpha$  therapy.

The incidence of primary or late brain metastases due to RCC has been reported to be as high as 11% in a large autopsy series (Saitoh 1981). In the present study, no brain metastases were present at the beginning of the treatment since it was an exclusion criterion. The incidence of brain metastases during IFN- $\alpha$  or after discontinuation of IFN- $\alpha$  was one in seven patients: one third developing brain metastases during the IFN- $\alpha$  therapy, and two thirds after discontinuation of the IFN-α therapy. The RCC patients with pulmonary metastases were the most likely subjects to develop brain metastases. This is in accordance with previous reports, where the lungs are the most common metastatic location associated with brain metastases (Mori et al. 1998). We suggest that this is due to the fact that patients with lung metastases have the longest survival and, consequently are the most susceptible to brain metastases. The median overall survival after detection of brain metastases was only 2.7 months, which is comparable to findings in other studies (Wronski et al. 1997, Decker et al. 1984). The median detection time of brain metastases was 21 months after the start of IFN-\alpha therapy. Men were more likely to suffer from brain metastases than women. Brain metastases are a late manifestation of the disease and are usually associated with progression also in extracranial sites. More effective treatments than are currently available are needed for those mRCC patients who develop brain metastases; both to treat actual brain metastases, as well as the extracranial disease.

The frequency of spontaneous regression in mRCC has been a controversial issue (Gleave *et al.* 1998, Marcus *et al.* 1993). Spontaneous regression has been suggested for approximately 1% of

RCC patients, although a higher percentage of up to 6.6% has been reported for selected patient groups (Gleave *et al.* 1998). In the present study, two patients (2%) had spontaneous regression prior to IFN- $\alpha$  therapy, one of whom, with lung metastases, achieved complete response to IFN- $\alpha$ , the other, with pleural and para-aortic lymph node metastases, had a stable disease. Spontaneous regression has most often occurred in RCC patients with pulmonary metastases (Gleave *et al.* 1998), as was also the case in this study.

## 6.1.2 CLINICOPATHOLOGICAL PROGNOSTIC FACTORS IN MRCC PATIENTS

Several prognostic models including different clinicopathological prognostic factors for survival have been created for mRCC. The heterogeneity of RCC within the same T-stage and grade has resulted in a need for prognostic models for prognostication and treatment modality selection. The Cleveland Clinic Foundation scoring system (Choueiri *et al.* 2007) was used in this study as the prognostic factors used in the model are easily applied and were available from the patient records. The widely used MSKCC 2002 prognostic model (Motzer *et al.* 2002b) could not have been used as LDH and corrected calcium values were not routinely recorded for the patients, as the study included patients from 1994 onwards. Nor could the Négrier *et al.* 2002 model have been used as the CRP value was not routinely recorded for the patients.

According to previous reports, prognostic models are good tools for prognostication and patient selection for risk groups. In clinical work, prognostic models could be more widely used to guide the laboratory schema. According to this study, the Cleveland Clinic Foundation scoring system differentiates well three different prognostic groups. However, different prognostic models are needed for different treatment modalities, for example, the MSKCC 2008 prognostic model (Motzer *et al.* 2008a) has been created for those RCC patients with sunitinib therapy.

In metastatic RCC, the following independent clinical prognostic factors for poor survival have been reported, e.g. poor performance status, high number of metastatic sites, and Hb level lower than normal baseline (Négrier *et al* 2002, Motzer *et al*. 1999). Similarly, in the present study, all these, as well as other metastatic site than the lung, and the presence of bone or liver metastases, were significant predictors of poor survival in Cox regression analysis. In multivariate analysis, response to IFN-α was associated with prolonged progression-free survival. De Forges *et al*. (1988) included the presence of liver metastasis in their prognostic model, but in the newer prognostic models, metastatic site has rarely been included (Motzer *et al*. 2004, Motzer *et al*. 2008a). Performance status is the most consistently used prognostic factor in the models. According to the current study, performance status and response to treatment are the most important clinical predictors in mRCC. Of course, response to treatment cannot be included in prognostic models at

the initiation of IFN- $\alpha$ , but it can be used when following and evaluating the patient's response to predict their prognosis. The presence of lung metastases was a significant prognostic factor for five-year survival in multivariate analysis. It has been previously reported that remission is more likely in those patients with only lung metastases, approximately 30%. This may be in part due to the methods of measuring pulmonary lesions. Small lesions are more likely to be seen by X-ray in the lung than by scintigraphy or CT in bone, liver or other viscera. Thus, it is possible that lung metastases are observed earlier than other metastases. In future, prognostic models should be developed by adding novel prognostic factors such as biomarkers.

## 6.1.3 TOXICITY OF IFN-α COMPARED TO OTHER THERAPIES

The intermittent administration of IFN- $\alpha$  was chosen since continuous therapy as reported, and also in our experience, has been associated with significant toxicity. The treatment with IFN- $\alpha$  was planned to be continued for 24 months or as long as progression or severe adverse effects were encountered. In some responding patients with good performance status, IFN- $\alpha$  therapy was continued for even longer.

Toxicity of IFN- $\alpha$  is dose-dependent and high doses of IFN- $\alpha$  result in excessive toxicity requiring dose reduction or discontinuation (Krown 1987, Trump *et al.* 1987). In this study, as in the studies of Minasian *et al.* (1993) and Steineck *et al.* (1990), the highest tolerable dose was defined for each patient by escalating the dose in the beginning of treatment. Previous studies have indicated that doses of 5 to 18 MU of IFN- $\alpha$  three times a week seem to be effective and tolerated (Krown 1987, Muss *et al.* 1987, Kirkwood *et al.* 1985). This is supported by the present result; only two patients had a dose of lower than 4.5 MU, one in a responding patient.

Altogether 8% of patients discontinued the treatment because of fatigue, elevation of liver enzymes, or cardiac arrhythmias. The degree of discontinuation is low compared to other studies with IFN- $\alpha$  (Fosså 1988, Fosså *et al.* 1986). In the study of Muss *et al.* (1987), the frequency of grade 3 and 4 toxicity was greater in the group of patients who received an IFN- $\alpha$  dose from 30 to 50 MU/m² intravenously for five consecutive days every three weeks compared to those patients with 2 MU/m² subcutaneously tiw. In our study, grade 3 toxicity was rare. In the long-term analysis of 117 mRCC patients, no life-threatening side-effects were observed. However, for patients with cardiac problems, such as cardiac arrhythmias or insufficiency, IFN- $\alpha$  therapy must be carefully considered as IFN- $\alpha$  may cause sudden cardiac death (Olencki *et al.* 2001). IFN- $\alpha$  is also neurotoxic, causing depression, and in rare cases, confusion, but in this study mostly of grade 1 toxicity. Fatigue, fever, poor appetite, nausea, abnormal liver enzymes, and muscular or joint pain were found in more than 20% of patients in the current study. These data show that prolonged and

intermittently administered IFN- $\alpha$  is well tolerated; the one-week pause every four weeks allows most patients to continue prolonged treatment with the highest tolerable dose.

The toxicity profile of IFN- $\alpha$  is different to that of sunitinib, which often causes diarrhea, fatigue, and nausea, and may cause anemia, leucopenia, hypertension, migraine, palmar-plantar erythrodysesthesia, hypothyreosis, and decreased blood glucose level (Gore *et al.* 2009). In more rare cases, sunitinib causes cardiac failure, venous thromboembolic events, and pulmonary events, such as dyspnea. Both IFN- $\alpha$  and sunitinib may in very rare cases cause encephalopathy syndrome (Cumurciuc *et al.* 2008, Mitsuyama *et al.* 1992). Neither the subcutaneously administered IFN- $\alpha$  nor the orally administered sunitinib require ward or policlinical resources as do the intravenous therapies. The different toxicity profiles of IFN- $\alpha$  and sunitinib enable the choosing of medication, paying attention to the patients' concomitant diseases and medication.

## 6.1.4 TIMING OF IFN-α IN THE ERA OF NOVEL TARGETED THERAPIES

In the era of novel targeted therapies, the choices for mRCC therapy have increased. Sunitinib, sorafenib, temsirolimus, everolimus, and bevacizumab have been approved for clinical use in the EU between 2006 and 2009. Sunitinib improves disease-free survival with acceptable toxicity. The addition of bevacizumab to IFN- $\alpha$  in conventional RCC increases the median progression-free survival. With IFN- $\alpha$  plus bevacizumab, fatigue, stomatitis and hematological toxicity have been less common compared to sunitinib therapy. Temsirolimus has recently been observed to improve overall survival of mRCC patients, even those with non-conventional RCC (Dutcher *et al.* 2009). As a second-line therapy in cytokine refractory mRCC, bevacizumab increases time to progression for patients in the intermediate risk group with good performance status (Yang *et al.* 2003a), while sorafenib increases progression-free survival. After the progression on other targeted therapies, everolimus prolongs progression-free survival (Motzer *et al.* 2008b). ESMO, NCCN, and EAU have created guidelines for the treatment of RCC, INF- $\alpha$  has been defined as an optional therapy (Escudier *et al.* 2009, Motzer *et al.* 2009, Ljundberg *et al.* 2007).

The current status of biological response modifiers to treat mRCC is not established. Sunitinib or bevacizumab plus IFN- $\alpha$  are currently considered the drugs of choice for good or intermediate risk groups (Escudier *et al.* 2007b, Motzer *et al.* 2007). For the poor risk group, temsirolimus or sunitinib is recommended (Motzer *et al.* 2007, Hudes *et al.* 2007). The progression-free survival of the patients with IFN- $\alpha$  in the present study was better compared to the progression-free survival with IFN- $\alpha$  or IFN- $\alpha$  with placebo in the studies of sunitinib and bevacizumab with IFN- $\alpha$  (Motzer *et al.* 2007, Escudier *et al.* 2007b). IFN- $\alpha$  based treatment duration in those studies was less than or equal to 5.1 months, whereas, in the present study, the treatment duration was longer (mean 11

months), a duration evidently made possible evidently by the one-week pause every four weeks. The long treatment duration may be the reason for better progression-free survival in the present study. The median overall survival and long-term survival analyses of sunitinib and bevacizumab with IFN- $\alpha$  are still ongoing.

Many ongoing trials are trying to answer the question of timing of IFN- $\alpha$ . According to the data of the present study, prolonged IFN- $\alpha$  therapy may also be considered as an additional choice for first-line therapy, especially in those patients whose concomitant disease or medication does not allow the use of sunitinib or bevacizumab. Prolonged IFN- $\alpha$  therapy may not be beneficial in patients in the poor risk group but it is beneficial in those in the intermediate risk group. Previously, conventional RCC has been proven to have the best response to and most favorable outcome from IFN- $\alpha$  compared to the other RCC subgroups (Motzer *et al.* 2002a). Differences in survival between IFN- $\alpha$  and the novel therapies cannot be reliably compared, as long-term survival data on novel therapies are not yet available. However, comparing prolonged IFN- $\alpha$  therapy to sunitinib, or bevacizumab with IFN- $\alpha$  in a randomized trial would give more exact information about whether there are differences in survival.

Response rates in mRCC vary greatly in different studies. In different sunitinib trials, response rates have varied from 9% to 44% (Gore *et al.* 2007, Motzer *et al.* 2007) With temsirolimus therapy, objective responses were infrequent as was the case with IFN- $\alpha$  therapy (Dutcher *et al.* 2009). Prior observations have shown that the difference in remission rates with different cytokine therapies has not been found to be a reliable surrogate for survival in mRCC (Coppin *et al.* 2007). The two following examples describe this phenomenon. Initial nephrectomy prior to planned IFN- $\alpha$  for mRCC patients improves survival and delays time to progression despite an unimproved response rate (Flanigan *et al.* 2001, Mickisch *et al.* 2001). Also, combining LD-IL-2 with IFN- $\alpha$  increases the response rate compared to one of the therapies alone, but the improvement in response rate does not translate into better survival (Négrier *et al.* 2007, Négrier *et al.* 1998). For this reason, survival data are more precise than response rates for comparison of results in different studies.

High-dose-IL-2 (HD-IL-2) may increase the complete response rate and improve survival in conventional RCC patients with the poorest prognosis (Coppin *et al.* 2007, Spanknebel *et al.* 2005); those with primary tumor still in place or with either liver or bone metastases (McDermott *et al.* 2005). HD-IL-2 has not been compared to LD-IL-2 or IFN-α in a randomized trial, but it has been observed that HD-IL-2 may cause durable responses in 7-8% of patients (McDermott *et al.* 2005, Yang *et al.* 2003b). HD-IL-2 therapy may increase cardiac toxicity, when administered after

VEGFR-TKI therapy (vascular endothelial growth factor receptor tyrosine kinases). HD-IL-2 is the therapy of choice for only a few patients because of its high toxicity; patients have to have (very) good performance status and organ function, such as cardiac function (Spanknebel *et al.* 2005). The immunohistochemical analysis of CA9 expression can find those patients who will benefit from IL-2 therapy (Atkins *et al.* 2005, Bui *et al.* 2003): higher CA9 expression predicts longer survival compared to low CA9 expression.

Currently, patients should be treated in trials, if possible, to obtain more knowledge about the timing of different therapies. Also, more survival data with different therapies are needed from non-conventional types of RCC. PEG-IFN- $\alpha$  is more convenient to administer and has potential for increased efficacy and less toxicity compared to IFN- $\alpha$  (Sunela *et al.* 2009, Feldman *et al.* 2008); this should also be further assessed in clinical trials. Assessing the patients into different prognostic groups is nowadays possible with many models; e.g. for IFN- $\alpha$  with MSKCC (Motzer *et al.* 2002b), the Cleveland Clinic Foundation (Choueiri *et al.* 2007), or Group Francais d'Immunotherapie (Négrier *et al.* 2002), and for sunitinib with MSKCC (Motzer *et al.* 2008a). In future, vaccine delivery systems for generation of immune responses against RCC with IFN- $\alpha$  (Hawkins *et al.* 2009, Viaud *et al.* 2009) or alone (Gitlitz *et al.* 2003), and gene therapy (Guse *et al.* 2009) as targeted routine therapies may be used clinically for mRCC patients. Additionally, vaccine therapy is currently the only promising systemic therapy in the adjuvant setting for RCC patients.

## 6.2 BIOLOGICAL EFFECTS OF IFN-α

IFN- $\alpha$  is a pleiotropic cytokine and it has immunomodulatory, antiangiogenic, proapoptotic, antiviral and antiproliferative effects. IFN- $\alpha$  has different cytostatic and immunomodulatory effects (Pestka 1983). IFN- $\alpha$  is known to activate monocytes and NK cells. The exact antitumor mechanisms against RCC have not been defined. For targeted therapy, knowledge of specific types of the biochemical derangements created by IFN- $\alpha$  is needed. In this study, the impact of IFN- $\alpha$  on phagocyte receptors for IgG and complement in monocytes and neutrophils was investigated. The impact of IFN- $\alpha$  on phagocyte receptors has not been previously reported.

According to the present study, in mRCC patients in neutrophils the expression of CR3 receptors and the proportion of Fc $\gamma$ RI positive neutrophils was significantly raised. As CR3 is previously known to serve as an adhesion molecule, which binds neutrophils to other cells, e.g. cancer cells, the observed activation may reflect a response of neutrophils to cancer cells. Fc $\gamma$ RI has previously been shown to be important in cell-mediated cytoxocity. In monocytes, the expression of all phagocyte receptors, except Fc $\gamma$ RIII, were raised when compared to controls. In neutrophils, IFN- $\alpha$ 

treatment lowered the elevated receptor expressions. During early treatment (<2 months) a significant decrease in the expression of CR1 receptors in neutrophils was observed. In monocytes, a significant activation of the expression of Fc $\gamma$ RI receptors was observed. The data of the current study show that changes in receptor expression reflect the inflammatory activation of phagocytes in mRCC. IFN- $\alpha$ , both in vivo and in vitro, modulates the expression of phagocytic receptors.

The observed receptor expression in mRCC patients in the present study differs from previous observations in patients with infectious disease, as reported by Leino *et al.* (1997). The observed receptor expression during IFN- $\alpha$  therapy differs from previous observations after the induction of IFN- $\gamma$  reported by Buckle and Hogg (1989). Previous clinical studies with mRCC have indicated a lack of efficacy of IFN- $\gamma$ , whereas IFN- $\alpha$  improves overall survival in mRCC with a response rate of 14% to 30% (MRCRCC *et al.* 1999, Pyrhönen *et al.* 1999, Gleave *et al.* 1998). In this study, the differences in the receptor expression reflect the impact of IFN- $\alpha$ . The differences are not similar to what has been observed due to infectious disease or IFN- $\gamma$ .

This sensitive, although arbitrary, method of investigation is useful in characterizing and differentiating specific clinical conditions. Although it is not sensitive enough for cancer diagnosis, it may add information on specific treatment effects and immunomodulation. This is in agreement with our clinical observation of early radiological progression in some patients responding later when treatment with IFN- $\alpha$  was continued. By using whole blood leukocytes, expression provoked by purification procedures can be avoided (Leino *et al.* 1997).

# 6.3 CLASSIFICATIONS ACCORDING TO MORPHOLOGY AND GENETIC FINDINGS IN RCC

The Heidelberg classification, which was published in 1997, subclassifying RCCs, was the first to combine morphology and genetic findings, and is considered a pioneer work in the field. Therefore, it was the natural choice in the studies of p53, Ki-67 and COX-2 at the time. Later, in 2004, WHO published the reassessed classification which is also based on both genetic and pathological abnormalities. The Heidelberg classification is still widely used worldwide. Both the Heidelberg and the WHO 2004 classifications contain conventional, papillary, chromophobe and collecting duct RCCs, which are the most often observed subtypes in RCC. In the Heidelberg classification, other RCC types are classified as unclassified type of RCC. In the WHO 2004 classification, many rare subtypes are added. These rare subtypes expand our knowledge of the biology of RCCs. In statistical regression analysis, the Heidelberg classification is a useable and clear classification, because the very rare subtypes, excluding collecting duct RCC, are classified as unclassified RCC. It is known that conventional, papillary, chromophobe and collecting duct RCCs differ from each

others in the five-year DSS in localized RCC, and response to treatment modalities in mRCC. However, in studies, neither the Heidelberg nor the WHO 2004 classification are found as independent prognostic factors for survival in localized or metastatic RCC. In future, classifications according to both morphology and genetic findings will be developed as new knowledge of the RCC biology is acquired.

## 6.4 T-STAGE AND GRADE AS PROGNOSTIC FACTORS IN RCC

Tumor stage and grade have previously been identified as the most important prognostic factors in RCC. The current study indicates that T-stage is a prognostic factor for metastases-free and overall survival in RCC patients. In patients who later developed metastatic disease, high T-stage caused twice the risk of metastatic disease and three times the risk of death compared with low T-stage. This indicates that as the tumor size increases, the more aggressive its growth becomes and the more probable is tumor cell dissemination, as can be expected. These results parallel a previous observation (Kirkali *et al.* 2001), and confirm recent analyses on the predictive power of T-stage in the 1997 and 2002 pTNM classification (Sobin and Wittekind 2002, Tsui *et al.* 2000a, Javidan *et al.* 1999). A uniform staging classification, the TNM staging system, has increased the cooperation between oncologists and pathologists concerning the outcome of RCC patients (Javidan *et al.* 1999).

Previously, it has been suggested that T-stage is not an important prognostic factor in the survival of patients who have neither lymph node nor distant metastases (Giuliani *et al.* 1990). However, the therapeutic value of lymph node dissection remains unproven (Mickish 1999). For this reason, extensive lymph node dissection was not carried out, and no systematic data on metastatic lymph nodes in nephrectomized patients were available in the present study. Current results suggest that T-stage alone is a valuable prognostic factor for survival, even when the status of lymph nodes is unknown.

In the present study, T-stage was found to be an important factor in predicting the survival of patients who underwent nephrectomy. Therefore, T-stage can be used in estimating the correct duration and frequency of surveillance of RCC patients after nephrectomy. Additionally, high T-stage has been used as an inclusion criterion for adjuvant treatments in trials (Atzopodien *et al.* 2005, Repmann *et al.* 2003).

Moreover, T-stage seems to be an independent prognostic factor in mRCC patients. In this study, the association between T-stage and overall survival was also found in those with primary metastases at the time of nephrectomy. T-stage is not typically used in prognostic models in

mRCC, a UCLA model (Zisman *et al.* 2002b) being an exception. T-stage seems to be a good tool in prognostic evaluation in mRCC patients and could be included in prognostic models.

Nuclear grade is typically used in prognostic models in locally confined RCC. In the present study, in the group of patients with later metastases, tumor grade was not associated with overall survival. Several other studies have also failed to demonstrate any difference in the survival of patients with different grades (Uchida *et al.* 2002, Rioux-Leclercq *et al.* 2000, Usubutum *et al.* 1998). This is partly because, as yet, no consensus has been reached on a universal tumor grading system (Kanamaru *et al.* 2001). However, the present results did point out differences in metastases-free survival between the highest and the lowest grades, although when all three grades were included the differences were no longer statistically significant. Similar results were also found in other studies on RCC. Overall, histopathological grade seem to be imprecise for prognostic evaluation in RCC patients (Uchida *et al.* 2002, Rioux-Leclercq *et al.* 2000, Lanigan *et al.* 1994).

## 6.5 BIOMARKERS AS PROGNOSTIC FACTORS IN RCC

Molecular tumor markers are expected to revolutionize the staging of RCC in the future (Srigley *et al.* 1997), as nowadays stratifying the patients into risk groups is largely done on the basis of clinopathological factors, e.g. clinical stage of the disease. Advances in the understanding of the pathogenesis, behavior, and molecular biology of RCC may help to better predict tumor prognosis, and thus improve survival of RCC carcinoma patients when a more tailored therapy can be given to each individual patient. Biomarkers, such as p53, Ki-67 and COX-2, are candidates for defining prognostic subgroups (Delahunt *et al.* 2002), and for guiding targeted therapies (Choisy-Rossi and Yonish-Rouach 1998, May and May 1999), as shown in the current study, where p53, Ki-67 and COX-2 had prognostic value in predicting survival.

In the current study, the p53 and Ki-67 analyses were performed not only in the group of all RCC types, but also in the conventional type RCC subgroup, to achieve a more homogenous group. However, the results were similar in both analyses, maybe due to the high proportion of conventional type RCC of all RCC types (more than 80%). Therefore, in other studies, the analyses were performed in all types of RCC. The size and distribution of the patient material in the study were typical of other RCC studies.

## 6.5.1 BIOMARKERS IN RELATION TO T-STAGE, GRADE OR OCCURRENCE OF METASTASES

The association between p53 and Ki-67 protein expressions in the present study is in accordance with findings in other studies (Olumi *et al.* 2001, Rioux-Leclercq *et al.* 2000), indicating that p53

accumulation and increased cell proliferative activity are parallel phenomena in RCC. The present finding of no association between COX-2 and p53 is in accordance with a previous observation (Cho *et al.* 2005), but the finding of no association between COX-2 and Ki-67 differs from a previous observation (Miyata *et al.* 2003).

p53 seems to associate weakly with tumor grade, as the association was seen only in univariate analysis. Nor was an association between p53 and grade observed in a previous microarray study (Zigeuner *et al.* 2004). In both studies, the nuclear grade was determined according to the WHO guidelines. The present results and others (Dudderidge *et al.* 2005, Rioux-Leclercq *et al.* 2000) show an association between Ki-67 and high T-stage and metastases development, indicating that Ki-67 is a marker for aggressive disease in RCC with an increased risk of early metastases development. In the present study, no association between COX-2 and tumor grade or T-stage was found. Published associations between COX-2 and T-stage or tumor grade in RCC have been contradictory. Yoshimura *et al.* (2004) demonstrated that COX-2 expression was highest in G1, as well as in pT1 RCC tumors, compared to other grades and stages, while in Hashimoto *et al.*'s study (2004), the results were the opposite, with increased COX-2 expression in the higher tumor grade and stage.

The current study indicates that the proportion of COX-2 positive tumors is highest in RCC with the ability to develop later metastases, when compared to both RCC without metastatic potential and RCC with primary metastases. To our knowledge, this finding is new. Previously, Miyata *et al.* (2003) observed that positive COX-2 expression associated with primary metastases in univariate analysis (when M0-patients were compared to M1-patients). Cho *et al.* (2005) found no association between positive COX-2 expression and metastases (when M0-patients were compared to M1-patients, or appearance of metastatic disease was compared to non-metastatic disease). In those studies, the method of analysis differs from that of the present study, where patients were divided into three categories according to the appearance of metastases. According to the present study, metastases-free survival is longer in patients with COX-2 positive tumors. The median metastases-free survival was 46 months in RCC with COX-2 positivity compared to 15 months in RCC with COX-2 negativity. The present results indicate that COX-2 positivity associates with the delay of metastatic formation in RCC patients who do not have disseminated disease at presentation. The results of the current study indicate that COX-2 negativity associates with an aggressive phenotype in mRCC disease.

## 6.5.2 BIOMARKER ASSOCIATION WITH SURVIVAL

MRCC is an extremely heterogeneous disease, with patients having an overall survival from a few months to several years, and to date, no biomarker is capable of predicting the survival of mRCC patients. Earlier published results on the associations of p53 and Ki-67 protein expression with survival have been controversial. Some studies have suggested that positive p53 protein expression associates with poor survival (Shvarts *et al.* 2005, Uchida *et al.* 2002), while others have observed no association between p53 and survival (Haitel *et al.* 2000, Rioux-Leclercq *et al.* 2000). In a tissue array study on metastasized patients, overexpression of p53 was associated with impaired DSS in renal carcinoma (Kim *et al.* 2004b). Dudderidge *et al.* (2005) found Ki-67 to be an independent prognostic factor for disease-free survival in RCC, but opposite results have also been published (Donskov *et al.* 2004, Yildiz *et al.* 2004). The present study supports the finding that there is no association between p53 or Ki-67 alone and survival in RCC patients. The difference between the previous and the present study was in the classification of metastases: Kim and coworkers classified both distant and local lymph node metastases as metastatic disease, whereas in the present study, only tumors with distant metastases were classified as metastatic.

The present study indicates that p53 and Ki-67 are not able to predict which patients will develop metastatic disease after nephrectomy, but interestingly, they predict poor survival in mRCC patients. Therefore, p53 and Ki-67 can help in determining metastatic patients with a poor prognosis and, e.g. those who might benefit from aggressive treatment, such as high-dose interleukin-2 (Spanknebel *et al.* 2005) or temsirolimus (Hudes *et al.* 2007).

Few studies have reported the results of an association between COX-2 expression and survival in RCC patients. Previously, Miyata *et al.* (2003) found that the five-year survival of patients with COX-2 positive tumors from nephrectomy was 66%, and of COX-2 negative patients 91% (Miyata *et al.* 2003). In Miyata's study, the patients were 86% M0 and 14% M1 at nephrectomy. Previously, to our knowledge, no results of COX-2 and overall survival in mRCC patients have been published. The current study indicates that COX-2 positivity predicts improved overall survival in patients with mRCC treated with IFN- $\alpha$ . This is in line with the previous study of Rini *et al.* (2006), in which COX-2 positivity associated with longer time to progression in the patients treated with celecoxib plus interferon- $\alpha$ . According to the present study, there is no association between COX-2 staining and response to IFN- $\alpha$  alone, while Rini *et al.* (2006), in a small-scale, study reported that all the RCC patients with objective responses to celecoxib plus interferon- $\alpha$  expressed COX-2 staining. Additionally, the present study indicates that COX-2 does not associate with the Heidelberg classification, which is in line with a previous result (Yoshimura *et al.* 2004).

## 6.5.3 INCIDENCE OF p53, Ki-67, AND COX-2 EXPRESSIONS

The incidence of p53- and Ki-67-positive expression in RCC tumors in the present study was low but similar to that in other RCC studies (Kirkali et al. 2001, Haitel et al. 2000, Rioux-Leclercq et al. 2000). It is known that in addition to melanoma, RCC belongs to tumors with a low incidence of p53 mutations compared to, e.g. prostate and bladder cancer (Haitel et al. 2000, Kirkali et al. 2001, Rioux-Leclercq et al. 2000). The low p53 mutation in different cancers (Olivier et al. 2002) and the low immunohistochemical staining of RCC tissue blocks for the p53 protein in this and other studies (Haitel et al. 2000, Rioux-Leclercq et al. 2000) suggest that mutations in p53 result in an accumulation of the p53 protein. In the study of Oda et al. (1995), p53 expression was found only in those components with p53 mutations, mainly in the sarcomatoid components. The 10% cut-off value of p53 and Ki-67 was selected to achieve statistically reliable results, and in accordance with previous studies on the subject (Olumi et al. 2001). Previously published reports indicate that the proportion of COX-2 positive cells varies in human RCCs (Cho et al. 2005, Miyata et al. 2003). In the present study, weak intensity of COX-2 staining was considered as COX-2 negative, which resulted in a lower number of positive COX-2 cells than in some other RCC studies (Tuna et al. 2004, Cho et al. 2005). For comparison, in the study of Miyata et al. (2003), the criterion for positive COX-2 expression was 5%, whereas in the present study it was considered to be 10%. Also different antibodies have been used in other studies (Rini et al. 2006, Cho et al. 2005, Hashimoto et al. 2004). This fact and the criteria for immunohistochemical classification may contribute to the difference in the results. Validation of immunohistochemical methods is needed before the methods could be widely adopted for in clinical use.

## 6.5.4 COMBINING MARKERS

In the present study, p53, Ki-67 and COX-2 were associated with metastatic appearance and survival. In multivariate analysis, COX-2 and Ki-67 were independent variables, indicating that they are both stronger biomarkers than p53 for the development of metastases in RCC. However, combining markers may specify prognostic subgroups better than observing a single marker. As shown in a study by Haitel *et al.* (2000), p53 was not an independent predictor for survival, but p53 and mdm2, a negative regulator of p53, showed a strong association with poor survival. In the present study, in RCC patients, double positivity for p53 and Ki-67 expression seems to indicate a higher probability of metastases than either marker alone. Additionally, combining COX-2 and Ki-67 increases their ability to predict survival in mRCC. In this study, median overall survival time of RCC with COX-2 negativity/Ki-67 positivity was 19 months, which was almost five times shorter than of RCC with COX-2 positivity/Ki-67 negativity. Median overall survival time of RCC with COX-2 negativity alone was 28 months, which was three times shorter than that of RCC with COX-2 positivity.

Prognostic markers can be used in patient counseling, to select treatment modalities, and to determine eligibility for clinical trials. Different prognostic models have been created to specify the prognosis of RCC patients (Motzer *et al.* 2004, Motzer *et al.* 1999); they typically include conventional prognostic markers. However, combining biomarkers and conventional clinical markers seems to predict DSS more accurately than grade or TNM stage alone, both in locally confined and metastatic RCC (Kim *et al.* 2004b).

## 6.5.5 TRENDS IN THE USE OF BIOMARKERS

Prospective clinical trials on the clinical use of p53, Ki-67, and COX-2 protein expression in predicting overall survival could answer the question of whether the expression of these biomarkers can be reliably used in mRCC. The present data show that these biomarkers cannot predict response to IFN- $\alpha$ , whether these biomarkers can predict response to novel targeted therapies should be investigated in trials.

The new era of genetic cancer studies shows great promise in terms of patient evaluation for new targeted therapies or immunotherapy. By means of the tissue microarray technique, thousands of tumors can be investigated simultaneously to determine the protein expression profile. However, creating a consensus in the tissue microarray construction protocol is challenging, as RCC is a relatively large-size tumor of a highly heterogenous nature (Signoretti *et al.* 2008). At current, whole tissue sections are considered the gold standard, but the more cores per tumor are sampled the fewer errors are introduced by limited sampling. Using gene chips to profile kidney tumors defines the genes that determine patient survival and response to therapy, thus enabling precise prognosis determination and individual treatment planning (Tan *et al.* 2008). Additionally, tissue microarrays enable the analysis of protein expression profiles in specimens to determine their potential clinical significance and role in RCC biology.

## 7 SUMMARY AND CONCLUSIONS

The present study led to the following conclusions.

I

Performance status and response to IFN- $\alpha$  are independent prognostic factors for overall survival in mRCC. Those patients who achieve objective responses (CR + PR) to prolonged IFN- $\alpha$  achieve twice as long median overall survival compared to those with stabilized disease, and ten times longer compared to those with progressive disease. Late objective responses can be seen even at 17 months after the initiation of therapy. For CR patients, a median response duration of four years is achievable. Approximately one in six patients is able to reach five-year survival, and patients with lung metastases are the most likely to achieve long-term survival. Also, a significant subgroup of patients in the intermediate risk group, seems to benefit from prolonged IFN- $\alpha$  therapy. T-stage is an independent prognostic factor for overall survival in those patients with primary metastases at the time of nephrectomy. Additionally, more than one metastatic site, bone or liver metastases, and Hb level lower than normal at baseline, are significant prognostic factors in mRCC. One in seven patients seems to develop brain metastases after the start of IFN- $\alpha$  as a late manifestation of the disease.

The highest tolerable doses of up to 18 MU of IFN- $\alpha$  three times a week seem to be effective. Responses are typically seen between the doses of from 9 to 18 MU. The rate of discontinuation of prolonged IFN- $\alpha$  therapy because of fatigue, elevation of liver enzymes or cardiac arrhythmias is low, approximately 8%, and the treatment does not seem to have a life-threatening effect. Prolonged IFN- $\alpha$  is feasible; the one-week pause every four weeks allows most patients to continue prolonged treatment at the highest tolerated dose.

In the era of novel targeted therapies, one choice in the treatment of mRCC patients is still IFN- $\alpha$  with a highest tolerated dose of 3 to 18 MU three times a week in a treatment cycle of three weeks, followed by a one-week pause. Prolonged treatment of more than 12 months in stable and responding patients is beneficial. Prolonged IFN- $\alpha$  therapy is beneficial in patients with good and intermediate risk.

II

Changes in receptor expression reflect the inflammatory activation of phagocytes in mRCC compared to a healthy control group. In mRCC patients in neutrophils the expression of CR3 receptors and the proportion of Fc $\gamma$ RI positive neutrophils increases. As CR3 is previously known to serve as an adhesion molecule, which binds neutrophils to other cells, e.g. cancer cells, the observed activation may reflect a response of neutrophils to cancer cells, and Fc $\gamma$ RI is previously known to be important in cell-mediated cytoxocity. The observed receptor expression in mRCC before and during IFN- $\alpha$  treatment differs from previous observations of patients with infectious disease, or after the induction of IFN- $\gamma$ . IFN- $\alpha$ , both in vivo and in vitro, modulates the expression of phagocytic receptors. Although investigation of the impact of IFN- $\alpha$  on phagocyte receptors is not sensitive enough for cancer diagnosis, it may add information about specific treatment effects and immunomodulation.

#### Ш

p53 associates weakly with tumor grade. Ki-67 associates with T-stage and metastatic development, indicating that Ki-67 is a marker for aggressive disease in RCC with an increased risk of early metastases development. The proportion of COX-2 positive tumors is highest in RCC with the ability to develop later metastases, when compared to both RCC without metastatic potential, and RCC with primary metastases. Metastases-free survival is longer in patients with COX-2 positive tumors compared to COX-2 negative tumors. These data show that COX-2 negativity associates with an aggressive phenotype in mRCC disease. COX-2 and Ki-67 alone are stronger biomarkers than p53 for the development of metastases in RCC.

p53 or Ki-67 alone are not valuable prognostic markers in locally confined RCC, but they can predict poor survival in mRCC. Therefore, p53 and Ki-67 can help in determining metastatic patients with a poor prognosis and, e.g. those who would benefit from high-dose IL-2 or temsirolimus. COX-2 positivity predicts improved overall survival in patients with mRCC treated with IFN-α. p53, Ki-67, and COX-2 cannot predict response to IFN-α. Investigating the ability of p53, Ki-67, and COX-2 protein expression to predict overall survival in a prospective clinical trial would answer the question of whether these biomarkers can be reliably used in mRCC.

Combining the results of COX-2 and Ki-67 expression, may predict overall survival in mRCC. In predicting the development of metastases in nephrectomized RCC patients, COX-2 alone or a covariation of p53 and Ki-67 seem to have prognostic value. Combining p53 or COX-2 with Ki-67 may result in more specific prognosis staging in RCC than observing a single marker. In future,

using the tissue microarray technique, the protein expression profile with several biomarkers can be determined quickly.

These findings increase our understanding of the molecular biology of locally confined RCC patients and metastatic RCC patients with IFN- $\alpha$  therapy. These markers might be useful as a part of the factors in prognostic models, and warrant further studies. The findings of such a study might be translated into prognostic tools that could be used in clinical work.

## ACKNOWLEDGEMENTS

This work was carried out at the Departments of Oncology and Radiotherapy, and Pathology and Clinical Chemistry at the University of Turku, Turku University Hospital. The research plan for my doctoral examination and academic thesis was accepted in the Faculty of Medicine, University of Turku, in February, 2001. I completed my specialization in Oncology and Radiotherapy at the University of Turku, in March, 2003. I started to specialize as a specialist in Clinical Genetics at University of Turku, in February, 2008. Preparing the academic thesis has not only provided me with a wealth of knowledge, but also taught me new ways to gather and process novel information, that has resulted in better care for patients. These years have also changed my way of thinking on medical questions, and my way of looking at the world.

First I wish to express my sincerest gratitude and thanks to my Supervisors, Professor Seppo Pyrhönen, Professor Eeva Salminen, and Docent Tarja-Terttu Pelliniemi for their guidance and support in familiarizing me with the exciting world of science. Their wide knowledge of the different areas of the subject has been a great asset, while their enthusiasm and scientific thinking have been a great source of inspiration. Additionally, I express many thanks to Docent Karl-Ove Söderström for instruction in pathology when co-analyzing the samples after incubation by biomarkers immunohistochemically. My special thanks to Docent Vesa Kataja in the University of Kuopio and Docent Anne Räisänen-Sokolowski in the University of Helsinki for reviewing the manuscript. I thank my co-workers, Professor Esa-Matti Lilius and Jari Nuutila PhD from the Department of Biochemistry and Food Chemistry at the University of Turku. They familiarized me with flow cytometer analysis of phagocytes. I also thank Väinämö Nikkanen MD, PhD, for sharing his clinical knowledge when analyzing the results of the phase II study of interferon-α in 2001. Additionally, I thank Hans Helenius MSc, Tero Vahlberg MSc, and Susanna Hinkka-Yli-Salomäki PhLic in the Department of Biostatistics at the University of Turku for their invaluable help with the statistical methods. I also thank the staff of the Pathological laboratories of Turku University Hospital, especially Ms Tuula Manninen, and the staff of Satakunta Central Hospital. Many thanks to Ms Tuula Raitanen in the patient file archives and Jacqueline Välimäki MA for assistance on the subtleties of English.

I wish to give my special thanks to the staff of the Department of the Clinical Genetics, who have introduced me to the fascinating world of hereditary disorders that span over generations and given a new angle to oncology. Especially to Maila Penttinen MD and Docent Marja Hietala for their keen clinical eye, Professor Jaakko Ignatius for his vast genetic knowledge, and Hannele Koillinen MD, PhD for her inspiring discussions on preparing a thesis and genetics. Your knowledge has been the source of my inspiration.

I wish to thank all my colleagues and friends in the Society of Oncology and Radiotherapy as well as in the Society of Medical Genetics in Finland for scientific and social get-togethers in different forums. Especially, I thank Docent Liisa Pylkkänen, the person who first familiarized me with the methods of cancer research for my advanced studies to qualify as a licentiate. Since those days, we have had many interesting discussions about oncology and different areas of life. I also want to give my special thanks to Maija Peitsalmi, my teacher in biology in Hämeenlinna Lyceum, the person who first introduced me to the exciting world of the cell. Those lessons in Lyceum were one of the sources of inspiration for me to later start research on this subject.

I express my deepest appreciation and heartfelt thanks to my dear parents, Maija-Liisa and Kalevi Kankuri. Their love, encouragement, and wisdom have guided my life, and were resources in preparing this academic dissertation. To my husband and soul-mate Tomi, I express my heartfelt thanks for sharing inspired conversations on the interesting phenomena of life, science, and family life. I want to thank you for your love, and for sharing this process with me, and for being there for me during these years. Our daughter Aurora, born in June, 2006, has brought so much joy into our life. Maija-Liisa, Kalevi, and Tomi, you have all supported me in my efforts to reach this goal. Additionally, I want to thank my relatives and friends for their interest in my research work.

I am grateful for the financial support received from the following: Turku University Hospital (EVO), The Cancer Society of Finland, The Cancer Society of South-Western Finland, The Finnish Medical Society Duodecim, The Finnish Cultural Foundation of Häme, the Foundation of Outpatient Care, the Ida Montin Foundation, and the Finnish Registry for Kidney Diseases.

Turku, December 2009

Minna Kankuri-Tammilehto

A GTan Clelik

## REFERENCES

Aaltomaa S, Lipponen P, Ala-Opas M, Eskelinen M and Syrjänen K (1997)

Prognostic value of Ki-67 expression in renal cell carcinomas. Eur Urol 31(3): 350-355.

Aass N, De Mulder PH, Mickisch GH, Mulders P, van Oosterom AT, van Poppel H, Fossa SD, de Prijck L and Sylvester RJ (2005) Randomized phase II/III trial of interferon Alfa-2a with and without 13-cis-retinoic acid in patients with progressive metastatic renal cell Carcinoma: the European Organisation for Research and Treatment of Cancer Genito-Urinary Tract Cancer Group (EORTC 30951). J Clin Oncol 23 (18): 4172-4178.

Abubakr YA, Chou TH and Redman BG (1994)

Spontaneous remission of renal cell carcinoma: a case report and immunological correlates. J Urol 152 (1): 156-157.

ACS (American Cancer Society) (2004)

Cancer facts and figures.

Alimov A, Sundelin B, Bergerheim U, Pavlenko M, Pisa P, Zetterberg A, Larsson C and Lagercrantz S (2004)

Molecular cytogenetic characterization shows higher genetic homogeneity in conventional renal cell carcinoma compared to other kidney cancers. Int J Oncol 25 (4): 955-960.

Amato RJ, Jac J, Giessinger S, Saxena S and Willis JP (2009)

A phase 2 study with a daily regimen of the oral mTOR inhibitor RAD001 (everolimus) in patients with metastatic clear cell renal cell cancer. Cancer 115 (11): 2438-2446.

Amin MB, Amin MB, Tamboli P, Javidan J, Stricker H, de-Peralta Venturina M, Deshpande A and Menon M (2002) Prognostic impact of histologic subtyping of adult renal epithelial neoplasms: an experience of 405 cases. Am J Surg Pathol 26 (3): 281-291

ANCR (Assocication of the Nordic Cancer Registries) (2009)

Version 3.4 April 2009.

Antonelli A, Portesi E, Cozzoli A, Zanotelli T, Tardanico R, Balzarini P, Grigolato PG and Cosciani Cunico S (2003) The collecting duct carcinoma of the kidney: a cytogenetical study. Eur Urol 43 (6): 680-685.

Argani P, Olgac S, Tickoo SK, Goldfischer M, Moch H, Chan DY, Eble JN, Bonsib SM, Jimeno M, Lloreta J, Billis A, Hicks J, De Marzo AM, Reuter VE and Ladanyi M (2007).

Xp11 translocation renal cell carcinoma in adults: expanded clinical, pathologic, and genetic spectrum. Am. J. Surg. Pathol. 31 (8):1149-1160.

Arteaga CL, Chinratanalab W and Carter MB (2001)

Inhibitors of HER2/neu (erbB-2) signal transduction. Semin Oncol 28 (6 Suppl 18): 30-35.

Atkins M, Regan M, McDermott D, Mier J, Stanbridge E, Youmans A, Febbo P, Upton M, Lechpammer M and Signoretti S (2005) Carbonic anhydrase IX expression predicts outcome of interleukin 2 therapy for renal cancer. Clin Cancer Res 11: 3714-3721.

Atzpodien J, Hoffmann R, Franzke M, Stief C, Wandert T and Reitz M (2002)

Thirteen-year, long-term efficacy of interferon 2alpha and interleukin 2-based home therapy in patients with advanced renal cell carcinoma. Cancer 95 (5): 1045-1050.

Atzpodien J, Royston P, Wandert T, Reitz M; DGCIN -- German Cooperative Renal Carcinoma Chemo-Immunotherapy Trials Group (2003)

Metastatic renal carcinoma comprehensive prognostic system. Br J Cancer 88 (3): 348-353.

Atzpodien J, Schmitt E, Gertenbach U, Fornara P, Heynemann H, Maskow A, Ecke M, Wöltjen HH, Jentsch H, Wieland W, Wandert T and Reitz M; German Cooperative Renal Carcinoma Chemo-Immunotherapy Trials Group (DGCIN) (2005)

Adjuvant treatment with interleukin-2- and interferon-alpha2a-based chemoimmunotherapy in renal cell carcinoma post tumour nephrectomy: results of a prospectively randomised trial of the German Cooperative Renal Carcinoma Chemoimmunotherapy Group (DGCIN). Br J Cancer 92 (5): 843-846.

Bassil B, Dosoretz DE and Prout Jr GR (1985)

Validation of the tumor, nodes and metastasis classification of renal cell carcinoma. J Urol 134: 450-454.

Belldegrun A, Tsui KH, deKernion JB and Smith RB (1999)

Efficacy of nephron-sparing surgery for renal cell carcinoma: analysis based on the new 1997 tumor-node- metastasis staging system. J Clin Oncol 17: 2868-2875.

Bellmunt J, Szczylik C, Feingold J, Strahs A, Berkenblit A (2008)

Temsirolimus safety profile and management of toxic effects in patients with advanced renal cell carcinoma and poor prognostic features. Ann Oncol 19 (8): 1387-1392.

Bennett RT, Lerner SE, Taub HC, Dutcher JP and Fleischmann J (1995)

Cytoreductive surgery for stage IV renal cell carcinoma. J Urol 154: 32-34.

Bergström A, Moradi T, Lindblad P, Nyren O, Adami HO and Wolk A (1999)

Occupational physical activity and renal cell cancer: a nationwide cohort study in Sweden. Int J Cancer 83: 186-191.

Bergström A, Pisani P, Tenet V, Wolk A and Adami HO (2001)

Overweight as an avoidable cause of cancer in Europe. Int J Cancer 91: 421-430. Erratum in: Int J Cancer 2001 92:927.

Bex A, van der Veldt AA, Blank C, van den Eertwegh AJ, Boven E, Horenblas S and Haanen J (2009)

Neoadjuvant sunitinib for surgically complex advanced renal cell cancer of doubtful resectability: initial experience with downsizing to reconsider cytoreductive surgery. World J Urol 27 (4): 533-539.

Blay JY, Rossi JF, Wijdenes J, Menetrier-Caux C, Schemann S, Négrier S, Philip T and Favrot (1997)

Role of interleukin-6 in the paraneoplastic inflammatory syndrome associated with renal-cell carcinoma. Int J Cancer 72 (3): 424-430.

Bleumer I, Oosterwijk E, Oosterwijk-Wakka JC, Voller MC, Melchior S, Warnaar SO, Mala C, Beck J and Mulders PF (2006) A clinical trial with chimeric monoclonal antibody WX-G250 and low dose interleukin-2 pulsing scheme for advanced renal cell carcinoma. J Urol. 175 (1): 57-62. Comment in: J Urol. 2006 Oct;176(4 Pt 1):1687.

Blom JH, van Poppel H, Marechal JM, Jacomin D, Sylvester R, Schröder FH and de Priick L (1999)

Radical nephrectomy with and without lymph node dissection: preliminary results of the EORTC randomized phase III protocol 30881. EORTC Genitourinary Group. Eur Urol 36 (6): 570-575.

Bodmer D, van den Hurk W, van Groningen JJ, Eleveld MJ, Martens GJ, Weterman MA and van Kessel AG (2002) Understanding familial and non-familial renal cell cancer. Hum Mol Genet. 11 (20): 2489-98.

Bonné AC, Bodmer D, Schoenmakers EF, van Ravenswaaij CM, Hoogerbrugge N and van Kessel AG (2004) Chromosome 3 translocations and familial renal cell cancer. Curr Mol Med 4 (8): 849-854.

Bonsib SM (2005)

T2 clear cell renal cell carcinoma is a rare entity: a study of 120 clear cell renal cell carcinomas. J. Urol 174 (4 Pt 1): 1199-1202; discussion 1202.

Bosniak MA, Birnbaum BA, Krinsky GA and Waisman J (1995)

Small renal parenchymal neoplasms: further observations on growth. Radiology 197: 589-597.

Bot FJ, Godschalk JC, Krishnadath KK, van der Kwast TH and Bosman FT (1994)

Prognostic factors in renal-cell carcinoma: immunohistochemical detection of p53 protein versus clinico-pathological parameters. Int J Cancer 57 (5): 634-637.

Boxer RJ, Waisman J, Lieber MM, Mampaso FM and Skinner DG (1978)

Non-metastatic hepatic dysfunction associated with renal carcinoma. J Urol 119 (4): 468-471.

Bracarda S, Porta C, Boni C, Santoro A, Artioli F, Di Bartolomeo C, Contu A, Gasparro D, De Angelis V, Caserta C and GOIRC Study Group (2007)

Randomized prospective phase II trial of two schedules of sorafenib daily and interferon-α2a (IFN) in metastatic renal cell carcinoma (RAPSODY): GOIRC Study 0681. J Clin Oncol 2007 ASCO Annual Meeting Proceedings Part I. 25 (18S): 5100.

Brenner H and Hakulinen T (2001)

Long-term cancer patient survival achieved by the end of the 20<sup>th</sup> century: most up-to-date estimates from the nationwide Finnish cancer registry. Br J Cancer 85: 367-371.

Bretheau D, Lechevallier E, de Fromont M, Sault MC, Rampal M and Coulange C (1995)

Prognostic value of nuclear grade of renal cell carcinoma. Cancer 76 (12): 2543-2549.

Brunelli M, Eble JN, Zhang S, Martignoni G and Cheng L (2003).

Gains of chromosomes 7, 17, 12, 16, and 20 and loss of Y occur early in the evolution of papillary renal cell neoplasia: a fluorescent in situ hybridization study. Mod. Pathol 16 (10): 1053-1059.

Brüning T, Pesch B, Wiesenhütter B, Rabstein S, Lammert M, Baumüller A and Bolt HM (2003)

Renal cell cancer risk and occupational exposure to trichloroethylene: results of a consecutive case-control study in Arnsberg, Germany. Am J Ind Med 43 (3): 274-285.

Brüning T, Weirich G, Hornauer MA, Höfler H and Brauch H (1997)

Renal cell carcinomas in trichloroethene (TRI) exposed persons are associated with somatic mutations in the von Hippel-Lindau (VHL) tumor suppressor gene. Arch Toxicol. 71(5):332-335.

Buckle A-M and Hogg N (1989)

The effect of INF-gamma and colony-stimulating factors on the expression of neutrophil cell membrane receptors. J Immunol 143 (7): 2295-2301.

Bui MH, Seligson D, Han KR, Pantuck AJ, Dorey FJ, Huang Y, Horvath S, Leibovich BC, Chopra S, Liao SY, Stanbridge E, Lerman MI, Palotie A, Figlin RA and Belldegrun AS (2003)

Carbonic anhydrase IX is an independent predictor of survival in advanced renal clear cell carcinoma: implications for prognosis and therapy. Clin Cancer Res 9: 802-811.

Bui MH, Visapää H, Seligson D, Kim H, Han KR, Huang Y, Horvath S, Stanbridge EJ, Palotie A, Figlin RA and Belldegrun AS (2004)

Prognostic value of carbonic anhydrase IX and KI67 as predictors of survival for renal clear cell carcinoma. J Urol 171 (6 Pt 1): 2461-2466.

Buzaid AC, Robertone A, Kisala C and Salmon SE (1987)

Phase II study of interferon alfa-2a, recombinant (Roferon-A) in metastatic renal cell carcinoma. J Clin Oncol 5 (7): 1083-1089.

Canfield SE, Kamat AM, Sanchez-Ortiz RF, Detry M, Swanson DA and Wood CG (2006)

Renal cell carcinoma with nodal metastases in the absence of distant metastatic disease (clinical stage TxN1-2M0): the impact of aggressive surgical resection on patient outcome. J Urol 175 (3 Pt 1): 864-869.

Cangiano T, Liao J, Naitoh J, Dorey F, Figlin R and Belldegrun A (1999)

Sarcomatoid renal cell carcinoma: biologic behaviour, prognosis, and response to combined surgical resection and immunotherapy. J Clin Oncol 17: 523-528.

Cantell K and Hirvonen S (1977)

Preparation of human leukocyte interferon for clinical use. Tex Rep Biol Med 35:138-144.

Chao D, Zisman A, Pantuck AJ, Gitlitz BJ, Freedland SJ, Said JW, Figlin RA and Belldegrun AS (2002) Collecting duct renal cell carcinoma: clinical study of a rare tumor. J Urol 167: 71-74.

Cheville JC, Lohse CM, Zincke H, Weaver AL and Blute ML (2003)

Comparisons of outcome and prognostic features among histologic subtypes of renal cell carcinoma. Am J Surg Pathol 27 (5): 612-624

Childs R, Chernoff A, Contentin N, Bahceci E, Schrump D, Leitman S, Read EJ, Tisdale J, Dunbar C, Linehan WM, Young NS and Barrett AJ (2000)

Regression of metastatic renal-cell carcinoma after nonmyeloablative allogeneic peripheral-blood stem-cell transplantation. N Engl J Med 343 (11): 750-758. [Comment in: N Engl J Med 343 (11):802-803]

Cho DS, Joo HJ, Oh DK, Kang JH, Kim YS, Lee KB and Kim SJ (2005)

Cyclooxygenase-2 and p53 expression as prognostic indicators in conventional renal cell carcinoma. Yonsei Med J 46 (1): 133-140.

Cho KS, Choi YD, Kim SJ, Kim CI, Chung BH, Seong do H, Lee DH, Cho JS, Cho IR and Hong SJ (2008)

A comprehensive prognostic stratification for patients with metastatic renal clear cell carcinoma. Yonsei Med J 49 (3): 451-458.

Choi JS, Kim MK, Seo JW, Choi YL, Kim DH, Chun YK and Ko YH (2006)

MET expression in sporadic renal cell carcinomas. J Korean Med Sci 21(4): 672-677.

Choisy-Rossi C and Yonish-Rouach E (1998)

Apoptosis and the cell cycle: the p53 connection. Cell Death Differ 5: 129-131.

Choueiri TK, Plantade A, Elson P, Negrier S, Ravaud A, Oudard S, Zhou M, Rini BI, Bukowski RM and Escudier B (2008) Efficacy of sunitinib and sorafenib in metastatic papillary and chromophobe renal cell carcinoma. J Clin Oncol 26 (1): 127-131. [Comments in Eur Urol. 2008 May;53(5):1085-1086; Eur Urol. 2008 Nov;54(5):1200-1; J Clin Oncol. 2008 Jul 10;26(20):3469-71; author reply 2471.]

Choueiri TK, Rini B, Garcia JA, Baz RC, Abou-Jawde RM, Thakkar SG, Elson P, Mekhail TM, Zhou M and Bukowski RM. (2007) Prognostic factors associated with long-term survival in previously untreated metastatic renal cell carcinoma. Ann Oncol 18 (2): 249-255.

Chow WH, Gridley G, Fraumeni JF Jr and Jarvholm B (2000)

Obesity, hypertension, and the risk of kidney cancer in men. N Engl J Med 343: 1305-1311.

Christophersen AO, Lie AK and Fosså SD (2006)

Unexpected 10 years complete remission after cortisone mono-therapy in metastatic renal cell carcinoma. Acta Oncol 45 (2): 226-228.

Clark JI, Atkins MB, Urba WJ, Creech S, Figlin RA, Dutcher JP, Flaherty L, Sosman JA, Logan TF, White R, Weiss GR, Redman BG, Tretter CP, McDermott D, Smith JW, Gordon MS and Margolin KA (2003)

Adjuvant high-dose bolus interleukin-2 for patients with high-risk renal cell carcinoma: a cytokine working group randomized trial. J Clin Oncol 21(16):3133-3140.

Clark PE, Schover LR, Uzzo RG, Hafez KS, Rybicki LA and Novick AC (2001)

Quality of life and sychological adaptation after surgical treatment for localized renal cell carcinoma: impact of the amount of remaining renal tissue. Urology 57: 252-256.

Coppin C, Porzsolt F, Autenrieth M, Kumpf J, Coldman A and Wilt T (2007)

Immunotherapy for advanced renal cell cancer. The Cochrane Database of Systematic Reviews, Issue 3: CD001425.

Corless CL, Kibel AS, Iliopoulos O and Kaelin WG Jr (1997)

Immunostaining of the von Hippel-Lindau gene product in normal and neoplastic human tissues. Hum Pathol. 28 (4): 459-464.

Creagan ET, Twito DI, Johansson SL, Schaid DJ, Johnson PS, Flaum MA, Buroker TR, Geeraerts LH, Veeder MH, Gesme DH Jr, et al. (1991)

A randomized prospective assessment of recombinant leukocyte A human interferon with or without aspirin in advanced renal adenocarcinoma. J Clin Oncol 9 (12): 2104-2109.

Cumurciuc R, Martinez-Almoyna L, Henry C, Husson H and de Broucker T (2008)

Posterior reversible encephalopathy syndrome during sunitinib therapy. Rev Neurol (Paris) 164 (6-7): 605-607.

Cunningham J (1938)

The kidney: tumors. 1938 Year book of Urology 167-192.

de Forges A, Rey A, Klink M, Ghosn M, Kramar A and Droz JP (1988)

Prognostic factors of adult metastatic renal carcinoma: a multivariate analysis. Semin Surg Oncol 4(3): 149-154.

Decatris M, Santhanam S and O'Byrne K (2002)

Potential of interferon-alpha in solid tumours: part 1.BioDrugs 16(4): 261-281.

Decker DA, Decker VL, Herskovic A and Cummings GD (1984).

Brain metastases in patients with renal cell carcinoma: prognosis and treatment, J Clin Oncol 2 (3): 169-173.

deKernion JB, Sarna G, Figlin R, Lindner A and Smith RB (1983)

The treatment of renal cell carcinoma with human leukocyte alpha-interferon. J Urol 130 (6): 1063-1066.

Delahunt B and Eble JN (1997).

Papillary renal cell carcinoma: a clinicopathologic and immunohistochemical study of 105 tumors. Mod. Pathol. 10 (6): 537-544.

Delahunt B, Becker RL, Bethwaite PB and Ribas JL (1994)

Computerized nuclear morphometry and survival in renal cell carcinoma: comparison with other prognostic indicators. Pathology 26: 353-358.

Delahunt B, Kittelson JM, McCredie MR, Reeve AE, Stewart JH and Bilous AM (2002)

Prognostic importance of tumor size for localized conventional (clear cell) renal cell carcinoma: assessment of TNM T1 and T2 tumor categories and comparison with other prognostic parameters. Cancer 94: 658-664.

Derweesh IH and Novick AC (2003)

Nephron-sparing surgery for renal celll carcinoma. Cancer Treat Res 116: 93-97.

Dinney CP, Awad SA, Gajewski JB, Belitsky P, Lannon SG, Mack FG and Millard OH (1992)

Analysis of imaging modalities, staging systems, and prognostic indicators for renal cell carcinoma. Urology 39 (2): 122-129.

Divgi CR, Pandit-Taskar N, Jungbluth AA, Reuter VE, Gönen M, Ruan S, Pierre C, Nagel A, Pryma DA, Humm J, Larson SM, Old LJ and Russo P (2007)

Preoperative characterisation of clear-cell renal carcinoma using iodine-124-labelled antibody chimeric G250 (124I-cG250) and PET in patients with renal masses: a phase I trial. Lancet Oncol 8 (4): 304-310.

Djordjevic G, Mozetic V, Mozetic DV, Licul V, Ilijas KM, Mustac E, Oguic R, Fuckar Z and Jonjic N (2007)

Prognostic significance of vascular endothelial growth factor expression in clear cell renal cell carcinoma. Pathol Res Pract 203 (2): 99-106.

Donskov F, Marcussen N, Hokland M, Fisker R, Madsen HH and von der Maase H (2004)

In vivo assessment of the antiproliferative properties of interferon-alpha during immunotherapy: Ki-67 (MIB-1) in patients with metastatic renal cell carcinoma. Br J Cancer 90 (3): 626-631.

Donskov F, Middleton M, Fode K, Meldgaard P, Mansoor W, Lawrance J, Thatcher N, Nellemann H and von der Maase H (2005) Two randomised phase II trials of subcutaneous interleukin-2 and histamine dihydrochloride in patients with metastatic renal cell carcinoma. Br J Cancer 93 (7): 757-762.

du Manoir S, Guillaud P, Camus E, Seigneurin D and Brugal G (1991)

Ki-67 labeling in postmitotic cells defines different Ki-67 pathways within the 2c compartment. Cytometry 12:455-463.

Dudderidge TJ, Stoeber K, Loddo M, Atkinson G, Fanshawe T, Griffiths DF and Williams GH (2005)

Mcm2, Geminin, and KI67 define proliferative state and are prognostic markers in renal cell carcinoma. Clin Cancer Res 11: 2510-2517.

Dutcher JP, de Souza P, McDermott D, Figlin RA, Berkenblit A, Thiele A, Krygowski M, Strahs A, Feingold J and Hudes G (2009) Effect of temsirolimus versus interferon-alpha on outcome of patients with advanced renal cell carcinoma of different tumor histologies. Med Oncol 26(2):202-209.

Eble JN (2003)

Mucinous tubular and spindle cell carcinoma and post-neuroblastoma carcinoma: newly recognised entities in the renal cell carcinoma family. Pathology 35(6):499-504.

Eble JN, Sauter G, Epstein JI and Sesterhenn IA, eds. (2004)

World Health Organization Classification of Tumors. Pathology and Genetics of Tumors of the Urinary System and Male Genital Organs. IARC Press, Lyon, p360.

Edwards BK, Brown ML, Wingo PA, Howe HL, Ward E, Ries LA, Schrag D, Jamison PM, Jemal A, Wu XC, Friedman C, Harlan L, Warren J, Anderson RN and Pickle LW (2005).

Annual report to the nation on the status of cancer, 1975-2002, featuring population-based trends in cancer treatment. J Natl Cancer Inst. 97 (19):1407-1427.

Elinq TE, Thompson DC, Foureman GL, Curtis JF and Hughes MF (1990)

Prostaglandin H synthase and xenobiotic oxidation. Ann Rev Pharmacol Toxicol 30: 1-45.

Elmore JM, Kadesky KT, Koeneman KS and Sagalowsky AI (2003)

Reassessment of the 1997 TNM classification system for renal cell carcinoma. Cancer 98 (11): 2329-2334.

Elson PJ, Witte RS and Trump DL (1988)

Prognostic factors for survival in patients with recurrent or metastatic renal cell carcinoma. Cancer Res 48: 7310-7313.

Ergen FB, Hussain HK, Caoili EM, Korobkin M, Carlos RC, Weadock WJ, Johnson TD, Shah R, Hayasaka S and Francis IR (2004) MRI for preoperative staging of renal cell carcinoma using the 1997 TNM classification: comparison with surgical and pathologic staging. AJR Am J Roentgenol 182 (1): 217-225.

Ernstoff MS, Fusi S and Kirkwood JM (1983)

Parameters of interferon action: I. Immunological effects of whole cell leukocyte interferon (IFN-alpha) in phase I-II trials. J Biol Response Mod 2 (6): 528-539.

Ernstoff MS, Nair S, Bahnson RR, Miketic LM, Banner B, Gooding W, Day R, Whiteside T, Hakala T and Kirkwood JM (1990) A phase IA trial of sequential administration recombinant DNA-produced interferons: combination recombinant interferon gamma and recombinant interferon alfa in patients with metastatic renal cell carcinoma. J Clin Oncol 8 (10): 1637-1649.

Ertl CW and Darras FS (1999)

Solitary metachronous contralateral adrenal metastasis from renal cell carcinoma. Urology 54 (1): 162.

Escudier B, Chevreau C, Lasset C, Douillard JY, Ravaud A, Fabbro M, Caty A, Rossi JF, Viens P, Bergerat JP, Savary J and Négrier S (1999)

Cytokines in metastatic renal cell carcinoma: is it useful to switch to interleukin-2 or interferon after failure of a first treatment? Groupe Français d'Immunothérape. J Clin Oncol 17 (7): 2039-2043.

Escudier B, Eisen T, Stadler WM, Szczylik C, Oudard S, Siebels M, Negrier S, Chevreau C, Solska E, Desai AA, Rolland F, Demkow T, Hutson TE, Gore M, Freeman S, Schwartz B, Shan M, Simantov R and Bukowski RM; TARGET Study Group (2007a) Sorafenib in advanced clear-cell renal-cell carcinoma. N Engl J Med 356(2):125-134. [Erratum in: N Engl J Med. 2007 357 (2): 203.]

Escudier B, Kataja V; ESMO Guidelines Working Group (2009)

Renal cell carcinoma: ESMO clinical recommendations for diagnosis, treatment and follow-up. Ann Oncol 20: 81-82.

Escudier B, Pluzanska A, Koralewski P, Ravaud A, Bracarda S, Szczylik C, Chevreau C, Filipek M, Melichar B, Bajetta E, Gorbunova V, Bay JO, Bodrogi I, Jagiello-Gruszfeld A and Moore N; AVOREN Trial investigators (2007b) Bevacizumab plus interferon alfa-2a for treatment of metastatic renal cell carcinoma: a randomised, double-blind phase III trial. Lancet 370 (9605): 2103-2111.

FCR (Finnish Cancer Registry) (2007)

Institute for Statistical and Epidemiological Cancer Research: Cancer in Finland 2004 and 2005. Cancer Statistics of the National Research and Development Centre for Welfare and Health (STAKES), publication No. 72. Cancer Society of Finland, Helsinki.

Feldman DR, Kondagunta GV, Schwartz L, Patil S, Ishill N, DeLuca J, Russo P and Motzer RJ (2008)

Phase II trial of pegylated interferon-alpha 2b in patients with advanced renal cell carcinoma. Clin Genitourin Cancer 6 (1): 25-30.

Fergany AF, Hafez KS and Novick AC (2000)

Long-term results of nephron sparing surgery for localized renal cell carcinoma: 10-year followup. J Urol 163: 442-445.

Ferlay J, Bray P, Pisani P and Parkin DM (2001)

GLOBOCAN 2000: Cancer incidence, mortality and prevalence worldwide, version 1.0. IARC CancerBase No. 5. IARCPress, Lyon.

Ficarra V, Guille F, Schips L, de la Taille A, Prayer Galetti T, Tostain J, Cindolo L, Novara G, Zigeuner R, Bratti E, Li G, Altieri V, Abbou CC, Zanolla L, Artibani W and Patard JJ (2005)

Proposal for revision of the TNM classification system for renal cell carcinoma. Cancer 104 (10): 2116-2123.

Ficarra V, Novara G, Iafrate M, Cappellaro L, Bratti E, Zattoni F and Artibani W (2007)

Proposal for Reclassification of the TNM Staging System in Patients with Locally Advanced (pT3-4) Renal Cell Carcinoma According to the Cancer-Related Outcome. Eur Urol. 51 (3): 722-729.

Ficarra V, Righetti R, Martignoni G, D'Amico A, Pilloni S, Rubilotta E, Malossini G and Mobilio G (2001) Prognostic value of renal cell carcinoma nuclear grading: multivariate analysis of 333 cases. Urol Int 67 (2): 130-134.

Figlin RA, deKernion JB, Mukamel E, Palleroni AV, Itri LM and Sarna GP (1988)

Recombinant interferon alfa-2a in metastatic renal cell carcinoma: assessment of antitumor activity and anti-interferon antibody formation. J Clin Oncol 6 (10): 1604-1610.

Figlin RA, Thompson JA, Bukowski RM, Vogelzang NJ, Novick AC, Lange P, Steinberg GD and Belldegrun AS (1999) Multicenter, randomized, phase III trial of CD8(+) tumor-infiltrating lymphocytes in combination with recombinant interleukin-2 in metastatic renal cell carcinoma. J Clin Oncol 17 (8): 2521-2529.

Finlay C, Hinds PW, Tan TH, Eliyahu D, Oren M and Levine AJ (1998)

Activating mutations for transformation by p53 produce a gene product that forms an hsc70-p53 complex with an altered half-life. Mol Cell Biol 8: 531-539.

Finney R (1973)

The value of radiotherapy in the treatment of hypernephroma--a clinical trial. Br J Urol 45 (3): 258-269.

Fisher RI, Rosenberg SA and Fyfe G (2000)

Long-term survival update for high-dose recombinant interleukin-2 in patients with renal cell carcinoma. Cancer J Sci Am 6 Suppl 1: S55-57.

Flanigan RC, Salmon SE, Blumenstein BA, Bearman SI, Roy V, McGrath PC, Caton JR Jr, Munshi N and Crawford ED (2001) Nephrectomy followed by interferon alfa-2b compared with interferon alfa-2b alone for metastatic renal-cell cancer. N Engl J Med 345: 1655-1659.

Flocks RH and Kadesky MC (1958)

Malignant neoplasms of the kidney: an analysis of 353 patients followed five years or more. J Urol 79: 196-201.

Fosså SD (1988)

Is interferon with or without vinblastine the "treatment of choice" in metastatic renal cell carcinoma? The Norwegian Radium Hospital's experience 1983-1986. Semin Surg Oncol 4 (3): 178-183.

Fosså SD, de Garis ST, Heier MS, Flokkmann A, Lien HH, Salveson A and Moe B (1986)

Recombinant interferon alfa-2a with or without vinblastine in metastatic renal cell arcinoma. Cancer 57 (8 Suppl): 1700-1704.

Fosså SD, Droz JP, Pavone-Macaluso MM, Debruyne FJ, Vermeylen K and Sylvester R (1992a)

Vinblastine in metastatic renal cell carcinoma: EORTC phase II trial 30882. The EORTC Genitourinary Group. Eur J Cancer. 28A(4-5): 878-880.

Fosså SD, Kramar A and Droz JP (1994)

Prognostic factors and survival in patients with metastatic renal cell carcinoma treated with chemotherapy or interferon-alpha. Eur J Cancer 30(9): 1310-1314.

Fosså SD, Lehne G, Gunderson R, Hjelmaas U and Holdener EE (1992b)

Recombinant interferon alpha-2A combined with prednisone in metastatic renal-cell carcinoma: treatment results, serum interferon levels and the development of antibodies. Int J Cancer 50 (6): 868-870.

Fosså SD, Martinelli G, Otto U, Schneider G, Wander H, Oberling F, Bauer HW, Achtnicht U and Holdener EE (1992c) Recombinant interferon alfa-2a with or without vinblastine in metastatic renal cell carcinoma: results of a European multi-center phase III study. Ann Oncol 3 (4): 301-305.

Frank I, Blute ML, Cheville JC, Lohse CM, Weaver AL and Zincke H (2002)

An outcome prediction model for patients with clear cell renal cell carcinoma treated with radical nephrectomy based on tumor stage, size, grade and necrosis: the SSIGN score. J Urol 168 (6): 2395-2400.

Freed SZ, Halperin JP and Gordon M (1977)

Idiopathic regression of metastases from renal cell carcinoma. J Urol 118 (4): 538-542.

Fuhrman SA, Lasky LC and Limas C (1982)

Prognostic significance of morphologic parameters in renal cell carcinoma. Am J Surg Pathol 6: 655-663.

Fyfe G, Fisher RI, Rosenberg SA, Sznol M, Parkinson DR and Louie AC (1995)

Results of treatment of 255 patients with metastatic renal cell carcinoma who received high-dose recombinant interleukin-2 therapy. J Clin Oncol 13 (3): 688-696.

Galban S, Martindale JL, Mazan-Mamczarz K, Lopez de Silanes I, Fan J, Wang W, Decker J and Gorospe M (2003) Influence of the RNA-binding protein HuR in pVHL-regulated p53 expression in renal carcinoma cells. Mol Cell Biol 23 (20): 7083-7095.

George CM, Vogelzang NJ, Rini BI, Geoffroy FJ, Kollipara P and Stadler WM (2002)

A phase II trial of weekly intravenous gemcitabine and cisplatin with continuous infusion fluorouracil in patients with metastatic renal cell carcinoma. Ann Oncol 13 (1): 116-120.

George DJ and Kaelin WG Jr (2003)

The von Hippel-Lindau protein, vascular endothelial growth factor, and kidney cancer. N Engl J Med 349 (5): 419-421.

Gerdes J, Lemke H, Baisch H, Wacker HH, Schwab U and Stein H (1984)

Cell cycle analysis of a cell proliferation-associated human nuclear antigen defined by the monoclonal antibody Ki-67. J Immunol 133: 1710-1715.

Gershanovich ML, Moiseenko VM, Vorob'ev AV and Kiapiulia K (1996)

Hormone therapy of advanced renal cancer with high-dose toremifene (Fareston). Vopr Onkol 42 (5): 105-109.

Ghanema N, Uhla M, Brinkb I, Schäfera O, Kellyb T, Moserb E and Langera M (2005)

Diagnostic value of MRI in comparison to scintigraphy, PET, MS-CT and PET/CT for the detection of metastases of bone. Eur J Radiol 55: 41-55.

Gilbert SM, Murphy AM, Katz AE, Goluboff ET, Sawczuk IS, Olsson CA, Benson MC and McKiernan JM (2006) Reevaluation of TNM staging of renal cortical tumors: recurrence and survival for T1N0M0 and T3aN0M0 tumors are equivalent. Urology 68 (2): 287-291.

Gill IS, Desai MM, Kaouk JH, Meraney AM, Murphy DP, Sung GT and Novick AC (2002)

Laparoscopic partial nephrectomy for renal tumor: duplicating open surgical techniques. J Urol 167: 469-467.

Gillenwater J and Howards S (1981)

The kidney, hypertension and vascular disease: tumor. 1981 Year Book Urol 116-133.

Gitlitz BJ, Belldegrun AS, Zisman A, Chao DH, Pantuck AJ, Hinkel A, Mulders P, Moldawer N, Tso CL and Figlin RA (2003) A pilot trial of tumor lysate-loaded dendritic cells for the treatment of metastatic renal cell carcinoma. J Immunother 26 (5): 412-419.

Giuliani L, Giberti C, Martorana G and Rovida S (1990)

Radical extensive surgery for renal cell carcinoma: long-term results and prognostic factors. J Urol 143: 468-473.

Gleave ME, Elhilali M, Fradet Y, Davis I, Venner P, Saad F, Klotz LH, Moore MJ, Paton V and Bajamonde A (1998) Interferon gamma-1b compared with placebo in metastatic renal-cell carcinoma. Canadian Urologic Oncology Group. N Engl J Med 338 (18): 1265-1271.

Gnarra JR, Tory K, Weng Y, Schmidt L, Wei MH, Li H, Latif F, Liu S, Chen F, Duh FM, *et al.* (1994) Mutations of the VHL tumor suppressor gene in renal carcinoma. Nat Genet 7: 85-90.

Gobbo S, Eble JN, Grignon DJ, Martignoni G, MacLennan GT, Shah RB, Zhang S, Brunelli M and Cheng L (2008) Clear cell papillary renal cell carcinoma: a distinct histopathologic and molecular genetic entity. Am J Surg Pathol 32 (8): 1239-1245.

Goldstein NS (1997)

The current state of renal cell carcinoma grading. Union Internationale Contre le Cancer (UICC) and the American Joint Committee on Cancer (AJCC) Cancer 80 (5): 977-980.

Golimbu M, Joshi P, Sperber A, Tessler A, Al-Askari S and Morales P (1986)

The anatomy of the intrarenal arteries and its application to segmental resection of the kidney. Br J Surg 42: 132-139.

Gore ME. Porta C. Oudard S et al (2007)

Sunitinib in metastatic renal cell carcinoma: preliminary assessment of toxicity in an expanded access trial with subpopulation analysis. ASCO Annual Meeting Proceedigs. J Clin Oncol 25 (18 Suppl.) 5010.

Gore ME, Szczylik C, Porta C, Bracarda S, Bjarnason GA, Oudard S, Hariharan S, Lee SH, Haanen J, Castellano D, Vrdoljak E, Schöffski P, Mainwaring P, Nieto A, Yuan J, Bukowski R (2009) Safety and efficacy of sunitinib for metastatic renal-cell carcinoma: an expanded-access trial. Lancet Oncol 10 (8): 757-763.

Greenlee RT, Murray T, Bolden S and Wingo PA (2000) Cancer statistics, 2000. CA Cancer J Clin 50 (1): 7-33.

Gucalp R, Ritch P, Wiernik PH, Sarma PR, Keller A, Richman SP, Tauer K, Neidhart J, Mallette LE, Siegel R, *et al.* (1992) Comparative study of pamidronate disodium and etidronate disodium in the treatment of cancer-related hypercalcemia. J Clin Oncol. 10 (1): 134-142.

Gudbjartsson T, Hardarson S, Petursdottir V, Thoroddsen A, Magnusson J and Einarsson GV (2005)

Histological subtyping and nuclear grading of renal cell carcinoma and their implications for survival: a retrospective nation-wide study of 629 patients. Eur Urol 48 (4): 593-600.

Guevremont C, Jeldres C, Perrotte P and Karakiewicz PI (2009)

Sorafenib in the management of metastatic renal cell carcinoma. Curr Oncol 16 Suppl 1: S27-32.

Guinan P, Sobin LH, Algaba F, Badellino F, Kameyama S, MacLennan G and Novick A (1997)

TNM staging of renal cell carcinoma: Workgroup No. 3. Union International Contre le Cancer (UICC) and the American Joint Committee on Cancer (AJCC). Cancer. 80: 992-993.

Guinan PD, Vogelzang NJ, Fremgen AM, Chmiel JS, Sylvester JL, Sener SF and Imperato JP (1995)

Renal cell carcinoma: tumor size, stage and survival. Members of the Cancer Incidence and End Results Committee. J Urol 153: 901-903.

Guse K, Diaconu I, Rajecki M, Sloniecka M, Hakkarainen T, Ristimäki A, Kanerva A, Pesonen S and Hemminki A (2009) Ad5/3-9HIF-Delta24-VEGFR-1-Ig, an infectivity enhanced, dual-targeted and antiangiogenic oncolytic adenovirus for kidney cancer treatment. Gene Ther 16 (8): 1009-1020.

Hafez KS, Fergany AF and Novick AC (1999)

Nephron sparing surgery for localized renal cell carcinoma: impact of tumor size on patient survival, tumor recurrence and TNM staging. J Urol 162: 1930-1933.

Hafez KS, Novick AC and Butler BP (1998)

Management of small solitary unilateral renal cell carcinomas: impact of central versus peripheral tumor location. J Urol 159: 1156-1160.

Hafez KS, Novick AC and Campbell SC (1997)

Patterns of tumor recurrence and guidelines for followup after nephron sparing surgery for sporadic renal cell carcinoma. J Urol 157: 2067-2070.

Haitel A, Wiener HG, Baethge U, Marberger M and Susani M (2000)

mdm2 expression as a prognostic indicator in clear cell renal cell carcinoma: comparison with p53 overexpression and clinicopathological parameters. Clin Cancer Res 6: 1840-1844.

Halperin EC and Harisiadis L (1983)

The role of radiation therapy in the management of metastatic renal cell carcinoma. Cancer 51 (4): 614-617.

Han KR, Janzen NK, McWhorter VC, Kim HL, Pantuck AJ, Zisman A, Figlin RA, Dorey FJ, Said JW and Belldegrun AS (2004) Cystic renal cell carcinoma: biology and clinical behavior. Urol Oncol 22 (5): 410-414.

Hanahan D and Weinberg RA (2000)

The hallmarks of cancer. Cell 100: 57-70.

Hand J and Broders A (1932)

Cacinoma of the kidney: the degree of malignancy in relation to factors bearing on prognosis. J Urol 28: 199.

Hara S, Kondo Y, Matsuzawa I, Hashimoto Y, Kimura G, Akimoto M and Imura N (2002)

Expression of cyclooxygenase-2 in human bladder and renal cell carcinoma. Adv Exp Med Biol 507: 123-126.

Harmen PE (1978)

TNM classification of malignant tumors. Union Internationale Contre le Cancer, Geneva.

Harshman LC, Li M and Srinivas S (2008)

The combination of thalidomide and capecitabine in metastatic renal cell carcinoma -- is not the answer. Am J Clin Oncol 31 (5): 417-423.

Hashimoto Y, Kondo Y, Kimura G, Matsuzawa I, Sato S, Ishizaki M, Imura N, Akimoto M and Hara S (2004)

Cyclooxygenase-2 expression and relationship to tumour progression in human renal cell carcinoma. Histopathology 44 (4): 353-359.

Hatcher PA, Anderson EE, Paulson DF, Carson CC and Robertson JE (1991)

Surgical management and prognosis of renal cell carcinoma invading the vena cava. J Urol 145 (1): 20-23; discussion 23-24.

Haven CJ, Wong FK, van Dam EW, van der Juijt R, van Asperen C, Jansen J, Rosenberg C, de Wit M, Roijers J, Hoppener J, Lips CJ, Larsson C, Teh BT and Morreau H (2000)

A genotypic and histopathological study of a large Dutch kindred with hyperparathyroidism-jaw tumor syndrome. J Clin Endocrinol Metab 85 (4): 1449-1454.

Hawkins RE, Macdermott C, Shablak A, Hamer C, Thistlethwaite F, Drury NL, Chikoti P, Shingler W, Naylor S and Harrop R (2009) Vaccination of patients with metastatic renal cancer with modified vaccinia Ankara encoding the tumor antigen 5T4 (TroVax) given alongside interferon-alpha. J Immunother 32 (4): 424-429.

Hellström PA, Bloigu R, Ruokonen AO, Vainionpää VA, Nuutinen LS and Kontturi MJ (1997)

Is routine ipsilateral adrenalectomy during radical nephrectomy harmful for the patient? Scand J Urol Nephrol 31: 19-25.

Hemminki K and Li X (2004)

Familial renal cell cancer appears to have a recessive component. J Med Genet 41 (5): 58.

Herman JG, Latif F, Weng Y, Lerman MI, Zbar B, Liu S, Samid D, Duan DS, Gnarra JR, Linehan WM, et al. (1994)

Silencing of the VHL tumor-suppressor gene by DNA methylation in renal carcinoma. Proc Natl Acad Sci U S A. 91 (21): 9700-9704.

Hermanek P and Sobin L, eds. (1987)

TNM classification of malignant tumors. 4 ed. UICC international union against cancer, UICC monograph series. Springer-Verlag, Berlin, p197.

Herschman HR (1996)

Prostaglandin synthase 2. Biochim. Biophys Act 1299: 125-140.

Hipp MM, Hilf N, Walter S et al (2007)

Sorafenib but not sunitinib affects the induction of immune responses. J Clin Oncol 25 (18 Suppl.), A3504.

Hofmockel G, Riess S, Bassukas ID and Dammrich J (1997)

Epidermal growth factor family and renal cell carcinoma: expression and prognostic impact. Eur Urol 31 (4): 478-484.

Hofmockel G, Wittmann A, Dammrich J and Bassukas ID (1996)

Related Articles, Expression of p53 and bcl-2 in primary locally confined renal cell carcinomas: no evidence for prognostic significance. Anticancer Res 16 (6B): 3807-3811.

Holan V, Kohno K and Minowada J (1991)

Natural human interferon-alpha augments interleukin-2 production by a direct action on the activated IL-2-producing T cells. J Interferon Res 11 (6): 319-325.

Hollingsworth JM, Miller DC, Daignault S and Hollenbeck BK (2006)

Rising incidence of small renal masses: a need to reassess treatment effect. J Natl Cancer Inst. 98(18):1331-1334

Homma Y, Kawabe K, Kitamura T, Nishimura Y, Shinohara M, Kondo Y, Saito I, Minowada S and Asakage Y (1995) Increased incidental detection and reduced mortality in renal cell cancer: recent retrospective analysis at eight institutions. Int J Urol 2: 77-80.

Hosmer DW and Lemeshow S. Applied Logistic Regression. (2nd ed), New York, John Wiley & Sons Inc, pp. 1-375, 2000.

Hricak H, Thoeni R, Carroll PR, Demas BE, Marotti M and Tanagho E (1988)

Detection and staging of renal neoplasm: a reassessment of MR imaging. Radiology 166: 643-649.

Hudes G, Carducci M, Tomczak P, Dutcher J, Figlin R, Kapoor A, Staroslawska E, Sosman J, McDermott D, Bodrogi I, Kovacevic Z, Lesovoy V, Schmidt-Wolf IG, Barbarash O, Gokmen E, O'Toole T, Lustgarten S, Moore L and Motzer RJ; Global ARCC Trial. (2007)

Temsirolimus, interferon alfa, or both for advanced renal-cell carcinoma. N Engl J Med 356 (22): 2271-2281.

Hunt JD, van der Hel OL, McMillan GP, Boffetta P and Brennan P (2005)

Renal cell carcinoma in relation to cigarette smoking: meta-analysis of 24 studies. Int J Cancer. 114 (1): 101-108.

IARCC (2004) In: IARC monographs on the evaluation of carcinogenic risks to humans. World Health rganization. Vol 83. IARCPress, Lyon.

Igel'nik AM, Grishin MA, Mazanov GP and Shchurandina NV (1991)

The conservative treatment of kidney cancer in middle-aged and elderly patients. Urol Nefrol (Mosk) (1): 29-34.

Iliopoulos O, Kibel A, Gray S and Kaelin WG Jr (1995)

Tumor suppression by the human von Hippel-Lindau gene product. Nat Med 1 (8): 822-826.

Isola J, Tanner M, Forsyth A, Cooke TG, Watters AD and Bartlett JM (2004)

Interlaboratory comparison of HER-2 oncogene amplification as detected by chromogenic and fluorescence in situ hybridization. Clin Cancer Res 10 (14): 4793-4798.

Itoi T, Yamana K, Bilim V, Takahashi K and Tomita F (2004).

Impact of frequent Bcl-2 expression on better prognosis in renal cell carcinoma patients. Br J Cancer 90 (1): 200-205.

Jacqmin D, Champy MF, Offner M and Bollack C (1988)

A trial of stimulation with tamoxifen of progesterone nuclear receptors in patients with kidney cancer. J Urol (Paris) 94 (9-10): 435-436.

Janzen NK, Kim HL, Figlin RA and Belldegrun AS (2003)

Surveillance after radical or partial nephrectomy for localized renal cell carcinoma and management of recurrent disease. Urol Clin North Am 30: 843-852.

Javidan J, Stricker HJ, Tampoli P, Amin MB, Peabody JO, Deshpande A, Menon M and Amin MB (1999)

Prognostic significance of the 1997 TNM classification of renal cell carcinoma. J Urol 162:1277-81.

Jayson M and Sanders H (1998)

Increased incidence of serendipitously discovered renal cell carcinoma. Urology 51: 203-205.

Johnsen JA and Hellsten S (1997)

Lymphatogenous spread of renal cell carcinoma: an autopsy study. J Urol 157: 450-453.

Juusela H, Malmio K, Alfthan O and Oravisto KJ (1977)

Preoperative irradiation in the treatment of renal adenocarcinoma. Scand J Urol Nephrol 11 (3): 277-281.

Kanamaru H, Akino H, Suzuki Y, Noriki S and Okada K (2001)

Prognostic value of nuclear area index in combination with the World Health Organization grading system for patients with renal cell carcinoma. Urology 57: 257-261.

Kanamaru H, Li B, Miwa Y, Akino H and Okada K (1999)

Immunohistochemical expression of p53 and bcl-2 proteins is not associated with sarcomatoid change in renal cell carcinoma. Urol Res 27: 169-173.

Kao GD, Malkowicz SB, Whittington R, D'Amico AV and Wein AJ (1994)

Locally advanced renal cell carcinoma: low complication rate and efficacy of postnephrectomy radiation therapy planned with CT. Radiology 193 (3): 725-730.

Karami S, Brennan P, Hung RJ, Boffetta P, Toro J, Wilson RT, Zaridze D, Navratilova M, Chatterjee N, Mates D, Janout V, Kollarova H, Bencko V, Szeszenia-Dabrowska N, Holcatova I, Moukeria A, Welch R, Chanock S, Rothman N, Chow WH and Moore LE (2008)

Vitamin D receptor polymorphisms and renal cancer risk in Central and Eastern Europe. J Toxicol Environ Health A 71 (6): 367-372.

Kasuya Y, Hosaka Y, Matsushima H, Goto T, Kitamura T (2001)

Differences in cell kinetic changes among renal cancer cell lines treated with interferon-alpha. Int J Urol 8 (8): 449-454.

Kattan MW, Reuter V, Motzer RJ, Katz J and Russo P (2001)

A postoperative prognostic nomogram for renal cell carcinoma. J Urol 166(1): 63-67.

Kavolius JP, Mastorakos DP, Pavlovich C, Russo P, Burt ME and Brady MS (1998)

Resection of metastatic renal cell carcinoma. J Clin Oncol 16: 2261-2266.

Kellokumpu-Lehtinen P and Nordman E (1990)

Recombinant interferon-alpha 2a and vinblastine in advanced renal cell cancer: a clinical phase I-II study. J Biol Response Mod 9 (4): 439-444.

Khan KNM, Stanfield KM, Trajkovic D and Knapp DW (2001)

Expression of cyclooxygenase-2 in canine renal cell carcinoma. Vet Pathol 38: 116-119.

Kim B and Louie AC (1992)

Surgical resection following interleukin 2 therapy for metastatic renal cell carcinoma prolongs remission. Arch Surg 127 (11): 1343-1349.

Kim HL, Belldegrun AS, Freitas DG, Bui MH, Han KR, Dorey FJ and Figlin RA (2003)

Paraneoplastic signs and symptoms of renal cell carcinoma: implications for prognosis. J Urol 170(5):1742-1746.

Kim HL, Seligson D, Liu X, Janzen N, Bui MH, Yu H, Shi T, Figlin RA, Horvath S and Belldegrun AS (2004b) Using protein expressions to predict survival in clear cell renal carcinoma. Clin Cancer Res 10 (16): 5464-5471.

Kim HL, Zisman A, Han KR, Figlin RA and Belldegrun AS (2004a)

Prognostic significance of venous thrombus in renal cell carcinoma. Are renal vein and inferior vena cava involvement different? J Urol 171 (2 Pt 1): 588-591.

Kirkali Z and Van Poppel H (2007b)

A critical analysis of surgery for kidney cancer with vena cava invasion. Eur Urol 52 (3): 658-662.

Kirkali Z, Algaba F, Scarpelli M, Trias I, Selvaggi FP and Van Poppel H (2007a)

What does the urologist expect from the pathologist (and what can the pathologists give) in reporting on adult kidney tumour specimens? Eur Urol 51 (5): 1194-1201.

Kirkali Z, Yorukoglu K, Ozkara E, Kazimoglu H and Mungan U (2001)

Proliferative activity, angiogenesis and nuclear morphometry n renal cell carcinoma. Int J Urol 8: 697-703.

Kirkwood JM, Harris JE, Vera R, Sandler S, Fischer DS, Khandekar J, Ernstoff MS, Gordon L, Lutes R, Bonomi P, et al. (1985) A randomized study of low and high doses of leukocyte alpha-interferon in metastatic renal cell carcinoma: the American Cancer Society collaborative trial. Cancer Res 45 (2): 863-871.

Kjaer M, Frederiksen PL and Engelholm SA (1987)

Postoperative radiotherapy in stage II and III renal adenocarcinoma. A randomized trial by the Copenhagen Renal Cancer Study Group. Int J Radiat Oncol Biol Phys 13 (5): 665-672.

Koski TA, Lehtonen HJ, Jee KJ, Ninomiya S, Joosse SA, Vahteristo P, Kiuru M, Karhu A, Sammalkorpi H, Vanharanta S, Lehtonen R, Edgren H, Nederlof PM, Hietala M, Aittomäki K, Herva R, Knuutila S, Aaltonen LA, Launonen V (2009)

Array comparative genomic hybridization identifies a distinct DNA copy number profile in renal cell cancer associated with hereditary leiomyomatosis and renal cell cancer. Genes Chromosomes Cancer 48 (7): 544-551.

Kovacs G, Akhtar M, Beckwith BJ, Bugert P, Cooper CS, Delahunt B, Eble JN, Fleming S, Ljungberg B, Medeiros LJ, Moch H, Reuter VE, Ritz E, Roos G, Schmidt D, Srigley JR, Störkel S, van den Berg E and Zbar B (1997) The Heidelberg classification of renal cell tumors. J Pathol 183: 131-133.

Krown SE (1987)

Interferon treatment of renal cell carcinoma. Current status and future prospects. Cancer 59 (3 Suppl): 647-651.

Kuebler JP, Hogan TF, Trump DL and Bryan GT (1984)

Phase II study of continuous 5-day vinblastine infusion in renal adenocarcinoma. Cancer Treat Rep 68 (6): 925-926.

Kuebler JP, Oberley TD, Meisner LF, Sidky YA, Reznikoff CA, Borden EC, Cummings KB and Bryan GT (1987) Effect of interferon alpha, interferon beta, and interferon gamma on the in vitro growth of human renal adenocarcinoma cells. Invest New Drugs 5 (1): 21-29.

Kurth KH, Debruyne FM, Hall RR, Denis L, Verbaes A, Bollack C, de Voogt HJ, de Pauw M and Silvester R (1987) Embolization and postinfarction nephrectomy in patients with primary metastatic renal adenocarcinoma. Eur Urol 13 (4): 251-255.

Lam JS, Shvarts O, Leppert JT, Pantuck AJ, Figlin RA ans Belldegrun AS (2005)

Postoperative surveillance protocol for patients with localized and locally advanced renal cell carcinoma based on a validated prognostic nomogram and risk group stratification system. J Urol 174: 466-472.

Lanigan D, Conroy R, Barry-Walsh C, Loftus B, Royston D and Leader M (1994)

A comparative analysis of grading systems in renal adenocarcinoma. Histopathology 24: 473-476.

Latif Z. Watters AD. Bartlett JM. Underwood MA and Aitchison M (2002)

Gene amplification and overexpression of HER2 in renal cell carcinoma. BJU Int 89 (1): 5-9.

Lau WK, Blute ML, Weaver AL, Torres VE and Zincke H (2000)

Matched comparison of radical nephrectomy vs. nephron-sparing surgery in patients with unilateral renal cell carcinoma and a normal contralateral kidney. Mayo Clin Proc 75: 1236-1242.

Lebeau A, Deimling D, Kaltz C, Sendelhofert A, Iff A, Luthardt B, Untch M and Lohrs U (2001)

Her-2/neu analysis in archival tissue samples of human breast cancer: comparison of immunohistochemistry and fluorescence in situ hybridization. J Clin Oncol 19 (2): 354-363.

Lee CT, Katz J, Shi W, Thaler HT, Reuter VE and Russo P (2000)

Surgical management of renal tumors 4 cm or less in a contemporary cohort. J Urol 163: 730-736.

Lehtonen HJ, Kiuru M, Ylisaukko-Oja SK, Salovaara R, Herva R, Koivisto PA, Vierimaa O, Aittomäki K, Pukkala E, Launonen V and Aaltonen LA (2006)

Increased risk of cancer in patients with fumarate hydratase germline mutation. J Med Genet. 43 (6): 523-526.

Leibovich BC, Blute ML, Cheville JC, Lohse CM, Weaver AL and Zincke H (2004)

Nephron sparing surgery for appropriately selected renal cell carcinoma between 4 and 7 cm results in outcome similar to radical nephrectomy. J Urol 171: 1066-1070.

Leino L, Sorvajärvi K, Katajisto J, Laine M, Lilius EM, Pelliniemi TT, Rajamäki A, Silvoniemi P and Nikoskelainen J (1997) Febrile infection changes the expression of IgG Fc receptors and complement receptors in human neutrophils *in vivo*. Clin Exp Immunol 107 (1): 37-43.

Lendvay TS and Marshall FF (2003)

The tuberous sclerosis complex and its highly variable manifestations. J Urol 169 (5): 1635-1642.

Levi F, Lucchini F, Negri E and La Vecchia C (2004)

Declining mortality from kidney cancer in Europe. Ann Oncol 15(7): 1130-1135.

Li GR, Soulie M, Escourrou G, Plante P and Pontonnier F (1996)

Micrometastatic adrenal invasion by renal carcinoma in patients undergoing nephrectomy. Br J Urol 78: 826-828.

Licht MR and Novick AC (1993)

Nephron sparing surgery for renal cell carcinoma. J Urol 149: 1-7.

Lindblad P, Chow WH, Chan J, Bergström A, Wolk A, Gridley G, McLaughlin JK, Nyrén O and Adami HO (1999) The role of diabetes mellitus in the aetiology of renal cell cancer. Diabetologia 42 (1): 107-112.

Lindblad P, Mellemgaard A, Schlehofer B, Adami HO, McCredie M, McLaughlin JK and Mandel JS (1995)

International renal-cell cancer study. V. Reproductive factors, gynecologic operations and exogenous hormones. Int J Cancer 61 (2): 192-198.

Linehan WM, Walther MM and Zbar B (2003)

The genetic basis of cancer of the kidney. J Urol 170: 2163-2172.

Lipponen P, Eskelinen M, Hietala K, Syrjänen K and Gambetta RA (1994)

Expression of proliferating cell nuclear antigen (PC10), p53 protein and c-erbB-2 in renal adenocarcinoma. Int J Cancer 57 (2): 275-280

Lissoni P, Mandalà M and Brivio F (2000)

Abrogation of the negative influence of opioids on IL-2 immunotherapy of renal cell cancer by melatonin. Eur Urol 38 (1): 115-118.

Ljungberg B, Alamdari FI, Rasmuson T and Roos G (1999)

Follow-up guidelines for nonmetastatic renal cell carcinoma based on the occurrence of metastases after radical nephrectomy. BJU Int 84 (4): 405-411.

Ljungberg B, Hanbury DC, Kuczyk MA, Merseburger AS, Mulders PF, Patard JJ, Sinescu IC; European Association of Urology Guideline Group for renal cell carcinoma (2007)

Renal cell carcinoma guideline. Eur Urol 51 (6): 1502-1510.

Ljungberg B, Stenling R, Osterdahl B, Farrelly E, Aberg T and Roos G (1995)

Vein invasion in renal cell carcinoma: impact on metastatic behavior and survival. J Urol 154 (5): 1681-164.

Lohse CM, Blute ML, Zincke H, Weaver AL and Cheville JC (2002)

Comparison of standardized and nonstandardized nuclear grade of renal cell carcinoma to predict outcome among 2,042 patients. Am J Clin Pathol 118 (6): 877-886.

Lokich J (1997)

Spontaneous regression of metastatic renal cancer. Case report and literature review. Am J Clin Oncol 20 (4): 416-418.

Lopez Hänninen E, Kirchner H and Atzpodien J (1996)

Interleukin-2 based home therapy of metastatic renal cell carcinoma: risks and benefits in 215 consecutive single institution patients. J Urol 155 (1): 19-25.

Lopez-Beltran A, Scarpelli M, Montironi R and Kirkali Z (2006)

2004 WHO classification of the renal tumors of the adults. Eur Urol 49 (5): 798-805.

Magyarlaki T, Buzogany I, Kaiser L, Sukosd F, Dobronte R, Simon B, Fazekas A and Nagy J (2001).

Prognostic histological and immune markers of renal cell carcinoma, Pathol, Oncol. Res. 7 (2): 118-124.

Maisonneuve P, Agodoa L, Gellert R, Stewart JH, Buccianti G, Lowenfels AB, Wolfe RA, Jones E, Disney AP, Briggs D, McCredie M and Boyle P (1999)

Cancer in patients on dialysis for end-stage renal disease: an international collaborative study. Lancet 354: 93-99.

Maitra A, Ashfaq R, Gunn CR, Rahman A, Yeo CJ, Sohn TA, Cameron JL, Hruban RH and Wilentz RE (2002)

Cyclooxygenase 2 expression in pancreatic adenocarcinoma and pancreatic intraepithelial neoplasia: an immunohistochemical analysis with automated cellular imaging. Am J Clin Pathol 118: 194-201.

Malchoff CD, Sarfarazi M, Tendler B, Forouhar F, Whalen G, Joshi V, Arnold A and Malchoff DM (2000)

Papillary thyroid carcinoma associated with papillary renal neoplasia: genetic linkage analysis of a distinct heritable tumor syndrome. J Clin Endocrinol Metab 85 (5): 1758-1764. [Comment in: J Clin Endocrinol Metab. 2000 May;85(5):1755-1757]

Maldazys JD and deKernion JB (1986)

Prognostic factors in metastatic renal carcinoma. J Urol 136(2): 376-379.

Mani S, Todd MB, Katz K and Poo WJ (1995)

Prognostic factors for survival in patients with metastatic renal cancer treated with biological response modifiers. J Urol 154: 35-40.

Marcus SG, Choyke PL, Reiter R, Jaffe GS, Alexander RB, Linehan WM, Rosenberg SA and Walther MM (1993) Regression of metastatic renal cell cercinoma after cytoreductive nephrectomy. J Urol 150 (2Pt1): 463-466.

Marincola FM, White DE, Wise AP, Rosenberg SA (1995)

Combination therapy with interferon alfa-2a and interleukin-2 for the treatment of metastatic cancer. J Clin Oncol 13 (5): 1110-1122

Marshall G, Ferreccio C, Yuan Y, Bates MN, Steinmaus C, Selvin S, Liaw J, Smith AH (2007)

Fifty-year study of lung and bladder cancer mortality in Chile related to arsenic in drinking water. J Natl Cancer Inst 99 (12): 920-928.

Marur S, Eliason J, Heilbrun LK, Dickow B, Smith DW, Baranowski K, Alhasan S and Vaishampayan U (2008) Phase II trial of capecitabine and weekly docetaxel in metastatic renal cell carcinoma. Urology 72 (4): 898-902.

Masferrer JL, Leahy KM, Koki AT, Zweifel BS, Settle SL, Woerner BM, Edwards DA, Flickinger AG, Moore RJ and Seibert K (2000)

Antiangiogenic and antitumor activities of cyclooxygenase-2 inhibitors. Cancer Res 60:1306-1311.

Masters JR (2007)

Clinical applications of expression profiling and proteomics in prostate cancer. Anticancer Res 27 (3A): 1273-1276.

Matei DV, Rocco B, Varela R, Verweij F, Scardino E, Renne G and De Cobelli O (2005)

Synchronous collecting duct carcinoma and papillary renal cell carcinoma: a case report and review of the literature. Anticancer Res 25 (1B): 579-586.

Mathew A, Devesa SS, Fraumeni JF Jr and Chow WH (2002)

Global increases in kidney cancer incidence, 1973-1992. Eur J Cancer Prev 11 (2): 171-178

Maxwell PH, Wiesener MS, Chang GW, Clifford SC, Vaux EC, Cockman ME, Wykoff CC, Pugh CW, Maher ER and Ratcliffe PJ (1999)

The tumor suppressor protein VHL targets hypoxia-inducible factors for oxygen-dependent proteolysis. Nature 399: 271-275.

May P and May E (1999)

Twenty years of p53 research: structural and functional aspects of the p53 protein. Oncogene 18: 7621-7636.

McDermott DF, Regan MM, Clark JI, Flaherty LE, Weiss GR, Logan TF, Kirkwood JM, Gordon MS, Sosman JA, Ernstoff MS, Tretter CP, Urba WJ, Smith JW, Margolin KA, Mier JW, Gollob JA, Dutcher JP and Atkins MB (2005)

Randomized phase III trial of high-dose interleukin-2 versus subcutaneous interleukin-2 and interferon in patients with metastatic renal cell carcinoma. J Clin Oncol 23 (1): 133-141. Erratum in: J Clin Oncol (2005) 23 (12): 2877.

McLaughlin JK, Chow WH, Mandel JS, Mellemgaard A, McCredie M, Lindblad P, Schlehofer B, Pommer W, Niwa S and Adami HO (1995)

International renal-cell cancer study. VIII. Role of diuretics, other anti-hypertensive medications and hypertension. Int J Cancer 63: 216-221.

McLaughlin JK, Mandel JS, Blot WJ, Schuman LM, Mehl ES and Fraumeni JF Jr (1984)

A population-based case-control study of renal cell carcinoma. J Natl Cancer Inst 72: 275-284.

McNichols DW, Segura JW and DeWeerd JH (1981)

Renal cell carcinoma: long-term survival and late recurrence. J Urol 126: 17-23.

Medeiros LJ, Jones EC, Aizawa S, Aldape HC, Cheville JC, Goldstein NS, Lubensky IA, Ro J, Shanks J, Pacelli A and Jung SH (1997)

Grading of renal cell carcinoma: Workgroup No. 2. Union Internationale Contre le Cancer and the American Joint Committee on Cancer (AJCC). Cancer 80 (5): 990-991.

Mejean A, Oudard S and Thiounn N (2003)

Prognostic factors of renal cell carcinoma. J Urol 169: 821-827.

Mekhail TM, Abou-Jawde RM, Boumerhi G, Malhi S, Wood L, Elson P and Bukowski R (2005)

Validation and extension of the Memorial Sloan-Kettering prognostic factors model for survival in patients with previously untreated metastatic renal cell carcinoma. J Clin Oncol 23: 832-841.

Menezes RJ, Tomlinson G and Kreiger N (2003)

Physical activity and risk of renal cell carcinoma. Int J Cancer 107: 642-646.

Mensing T, Welge P, Voss B, Fels LM, Fricke HH, Brüning T and Wilhelm M (2002)

Renal toxicity after chronic inhalation exposure of rats to trichloroethylene. Toxicol Lett. Mar 10;128(1-3):243-247.

Messing EM, Manola J, Wilding G, Propert K, Fleischmann J, Crawford ED, Pontes JE, Hahn R and Trump D; Eastern Cooperative Oncology Group/Intergroup trial (2003)

Phase III study of interferon alfa-NL as adjuvant treatment for resectable renal cell carcinoma: an Eastern Cooperative Oncology Group/Intergroup trial. J Clin Oncol 21 (7): 1214-1222.

Mickisch GH, Garin A, van Poppel H, de Prijck L, Sylvester R, European Organisation for Research and Treatment of Cancer (EORTC) Genitourinary Group (2001)

Radical nephrectomy plus interferon-alfa-based immunotherapy compared with interferon alfa alone in metastatic renal-cell carcinoma: a randomised trial. Lancet 358 (9286): 966-970.

Mickish GH (1999)

Lymphatic metastases in renal cell carcinoma. What is the value of operation and adjuvant therapy? Urologe A 38: 326-331.

Miller AB, Hoogstraten B, Staquet M and Winkler A (1981)

Reporting results of cancer treatment. Cancer 47: 207-214.

Milowsky MI, Rosmarin A, Tickoo SK, Papanicolaou N and Nanus DM (2002)

Active chemotherapy for collecting duct carcinoma of the kidney: a case report and review of the literature. Cancer 94 (1): 111-116.

Minasian LM, Motzer RJ, Gluck L, Mazumdar M, Vlamis V and Krown SE (1993)

Interferon alfa-2a in advanced renal cell carcinoma: treatment results and survival in 159 patients with long-term follow-up. J Clin Oncol 11: 1368-1375.

Mitsuyama Y, Hashiguchi H, Murayama T, Koono M and Nishi S (1992)

An autopsied case of interferon encephalopathy. Jpn J Psychiatry Neurol 46 (3): 741-748.

Miyata Y, Koga S, Kanda S, Nishikido M, Hayashi T and Kanetake H (2003)

Expression of cyclooxygenase-2 in renal cell carcinoma: correlation with tumor cell proliferation, apoptosis, angiogenesis, expression of matrix metalloproteinase-2, and survival. Clin Cancer Res 9 (5): 1741-1749.

Moch H, Gasser T, Amin MB, Torhorst J, Sauter G and Mihatsch MJ (2000)

Prognostic utility of the recently recommended histologic classification and revised TNM staging system of renal cell carcinoma: a Swiss experience with 588 tumors. Cancer 89: 604-614.

Moch H, Sauter G, Gasser TC, Buchholz N, Bubendorf L, Richter J, Jiang F, Dellas A and Mihatsch MJ (1997)

p53 protein expression but not mdm-2 protein expression is associated with rapid tumor cell proliferation and prognosis in renal cell carcinoma. Urol Res 25 Suppl 1: S25-30.

Montemurro F, Choa G, Faggiuolo R, Sperti E, Capaldi A, Donadio M, Minischetti M, Salomone A, Vietti-Ramus G, Alabiso O and Aglietta M (2003)

Safety and activity of docetaxel and trastuzumab in HER2 overexpressing metastatic breast cancer: a pilot phase II study. Am J Clin Oncol 26 (1): 95-97.

Mori Y, Kondziolka D, Flickinger JC, Logan T and Lunsford LD (1998)

Stereotactic radiosurgery for brain metastasis from renal cell carcinoma. Cancer 83 (2): 344-353.

Mostofi FK (1981)

Histological Typing of Kidney Tumors. World Health Organization, 1981.

Mostofi FK and Davis CJ in Collaboration with Sobin LH and Pathologists in 6 Countries. World Health Organization (1998) Histological Typing of Kidney Tumors; in (2<sup>nd</sup> ed): International Histological Classification of Tumors. Springer 1998, Berlin Heidelberg, pp. 1-117.

Motzer RJ, Agarwal N, Beard C, Bolger GB, Boston B, Carducci MA, Choueiri TK, Figlin RA, Fishman M, Hancock SL, Hudes GR, Jonasch E, Kessinger A, Kuzel TM, Lange PH, Levine EG, Margolin KA, Michaelson MD, Olencki T, Pili R, Redman BG, Robertson CN, Schwartz LH, Sheinfeld J, Wang J (2009)

NCCN clinical practice guidelines in oncology: kidney cancer. J Natl Compr Canc Netw 7 (6): 618-630.

Motzer RJ, Bacik J, Mariani T, Russo P, Mazumdar M and Reuter V (2002a)

Treatment outcome and survival associated with metastatic renal cell carcinoma of non-clear-cell histology. J Clin Oncol 20 (9): 2376-2381.

Motzer RJ, Bacik J, Murphy BA, Russo P and Mazumdar M (2002b)

Interferon-alfa as a comparative treatment for clinical trials of new therapies against advanced renal cell carcinoma. J Clin Oncol 20 (1): 289-296.

Motzer RJ, Bacik J, Schwartz LH, Reuter V, Russo P, Marion S and Mazumdar M (2004)

Prognostic factors for survival in previously treated patients with metastatic renal cell carcinoma. J Clin Oncol 22: 454-463.

Motzer RJ, Bander NH and Nanus DM (1996)

Renal-cell carcinoma. N Engl J Med 335: 865-875.

Motzer RJ, Berg W, Ginsberg M, Russo P, Vuky J, Yu R, Bacik J and Mazumdar M (2002c)

Phase II trial of thalidomide for patients with advanced renal cell carcinoma. J Clin Oncol 20 (1): 302-306.

Motzer RJ, Bukowski RM, Figlin RA, Hutson TE, Michaelson MD, Kim ST, Baum CM and Kattan MW (2008a) Prognostic nomogram for sunitinib in patients with metastatic renal cell carcinoma. Cancer 113 (7): 1552-1558.

Motzer RJ, Escudier B, Oudard S, Hutson TE, Porta C, Bracarda S, Grünwald V, Thompson JA, Figlin RA, Hollaender N, Urbanowitz G, Berg WJ, Kay A, Lebwohl D and Ravaud A; RECORD-1 Study Group (2008b)

Efficacy of everolimus in advanced renal cell carcinoma: a double-blind, randomised, placebo-controlled phase III trial. Lancet 372 (9637): 449-456.

Motzer RJ, Hutson TE, Tomczak P, Michaelson MD, Bukowski RM, Rixe O, Oudard S, Négrier S, Szczylik C, Kim ST, Chen I, Bycott PW, Baum CM and Figlin RA (2007)

Sunitinib versus interferon alfa in metastatic renal-cell carcinoma. N Engl J Med 356 (2): 115-124.

Motzer RJ, Mazumdar M, Bacik J, Berg W, Amsterdam A and Ferrara J (1999)

Survival and prognostic stratification of 670 patients with advanced renal cell carcinoma. J Clin Oncol 17: 2530-2540.

Motzer RJ, Mazumdar M, Bacik J, Russo P, Berg WJ and Metz EM (2000a)

Effect of cytokine therapy on survival for patients with advanced renal cell carcinoma. J Clin Oncol 18 (9): 1928-1935.

Motzer RJ, Murphy BA, Bacik J, Schwartz LH, Nanus DM, Mariani T, Loehrer P, Wilding G, Fairclough DL, Cella D and Mazumdar M (2000b)

Phase III trial of interferon alfa-2a with or without 13-cis-retinoic acid for patients with advanced renal cell carcinoma. J Clin Oncol 18 (16): 2972-2980.

Motzer RJ, Rakhit A, Ginsberg M, Rittweger K, Vuky J, Yu R, Fettner S and Hooftman L (2001a)

Phase I trial of 40-kd branched pegylated interferon alfa-2a for patients with advanced renal cell carcinoma. J Clin Oncol 19 (5): 1312-1319.

Motzer RJ, Rakhit A, Thompson JA, Nemunaitis J, Murphy BA, Ellerhorst J, Schwartz LH, Berg WJ and Bukowski RM (2001b) Randomized multicenter phase II trial of subcutaneous recombinant human interleukin-12 versus interferon-alpha 2a for patients with advanced renal cell carcinoma. J Interferon Cytokine Res 21 (4): 257-263.

MRCRCC (Medical Research Council Renal Cancer Collaborators) (1999)

Interferon-α and survival in metastatic renal carcinoma; early results of a randomised controlled trial. Lancet 353: 14-17.

Mulders P, Tso CL, Pang S, Kaboo R, McBride WH, Hinkel A, Gitlitz B, Dannull J, Figlin R and Belldegrun A (1998) Adenovirus-mediated interleukin-2 production by tumors induces growth of cytotoxic tumor-infiltrating lymphocytes against human renal cell carcinoma. J Immunother 21 (3): 170-180.

Mungan MU, Gurel D, Canda AE, Tuna B, Yorukoglu K and Kirkali Z (2006)

Expression of COX-2 in normal and pyelonephritic kidney, renal intraepithelial neoplasia, and renal cell carcinoma. Eur Urol 50 (1): 92-97.

Muss HB, Costanzi JJ, Leavitt R, Williams RD, Kempf RA, Pollard R, Ozer H, Zekan PJ, Grunberg SM, Mitchell MS, et al. (1987) Recombinant alfa interferon in renal cell carcinoma: a randomized trial of two routes of administration. J Clin Oncol 5 (2): 286-291.

Nakajima T, Suzuki M, Ando S, Iida T, Araki A, Fujisawa T and Kimura H (2006)

Spontaneous regression of bone metastasis from renal cell carcinoma; a case report. BMC Cancer 6:11.

Nanus DM, Garino A, Milowsky MI, Larkin M and Dutcher JP (2004)

Active chemotherapy for sarcomatoid and rapidly progressing renal cell carcinoma. Cancer 101 (7): 1545-1551.

Nanus DM, Pfeffer LM, Bander NH, Bahri S and Albino AP (1990)

Antiproliferative and antitumor effects of alpha-interferon in renal cell carcinomas: correlation with the expression of a kidney-associated differentiation glycoprotein. Cancer Res 50 (14): 4190-4194.

Négrier S, Caty A, Lesimple T, Douillard JY, Escudier B, Rossi JF, Viens P and Gomez F (2000)

Treatment of patients with metastatic renal carcinoma with a combination of subcutaneous interleukin-2 and interferon alfa with or without fluorouracil. Groupe Français d'Immunothérapie, Fédération Nationale des Centres de Lutte Contre le Cancer. J Clin Oncol 18 (24): 4009-4015.

Négrier S, Escudier B, Gomez F, Douillard JY, Ravaud A, Chevreau C, Buclon M, Pérol D and Lasset C (2002)

Prognostic factors of survival and rapid progression in 782 patients with metastatic renal carcinomas treated by cytokines: a report from the Groupe Français d'Immunothérapie. Ann Oncol. 2002 13 (9): 1460-1468.

Négrier S, Escudier B, Lasset C, Douillard JY, Savary J, Chevreau C, Ravaud A, Mercatello A, Peny J, Mousseau M, Philip T and Tursz T (1998)

Recombinant human interleukin-2, recombinant human interferon alfa-2a, or both in metastatic renal-cell carcinoma. Groupe Français d'Immunothérapie. N Engl J Med 1998 338 (18): 1272-1278.

Négrier S, Gomez F, Douillard JY, Ravaud A, Chevreau C, Buclon M, Perol D, Lasset C and Escudier B; Groupe Français d'Immunothérapie (2005)

Prognostic factors of response or failure of treatment in patients with metastatic renal carcinomas treated by cytokines: a report from the Groupe Français d'Immunothérapie. World J Urol 23 (3): 161-165.

Négrier S, Perol D, Ravaud A, Bay JO, Oudard S, Chabaud S, Fargeot P, Delva R, Deplanque G, Gravis G and Escudier B; French Immunotherapy Group (2008)

Randomized study of intravenous versus subcutaneous interleukin-2, and IFNalpha in patients with good prognosis metastatic renal cancer Clin Cancer Res 14 (18):5907-5912.

Négrier S, Perol D, Ravaud A, Chevreau C, Bay JO, Delva R, Sevin E, Caty A and Escudier B; For The French Immunotherapy Intergroup (2007)

Medroxyprogesterone, interferon alfa-2a, interleukin 2, or combination of both cytokines in patients with metastatic renal carcinoma of intermediate prognosis: results of a randomized controlled trial. Cancer 110 (11): 2468-2477.

Neidhart JA, Gagen MM, Young D, Tuttle R, Melink TJ, Ziccarrelli A and Kisner D (1984) Interferon-alpha therapy of renal cancer. Cancer Res 44 (9): 4140-4143.

Neves RJ, Zincke H and Taylor WF (1988)

Metastatic renal cell cancer and radical nephrectomy: identification of prognostic factors and patient survival. J Urol 139 (6): 1173-1176.

Novick AC (1995)

Partial nephrectomy for renal cell carcinoma. Urology 46: 149-152.

Nurmi MJ (1984)

Prognostic factors in renal carcinoma. An evaluation of operative findings. Br J Urol 56: 270-275.

Oda H, Nakatsuru Y and Ishikawa T (1995)

Mutations of the p53 gene and p53 protein overexpression are associated with sarcomatoid transformation in renal cell carcinomas. Cancer Res 55: 658-662.

Oh WK, Manola J, George DJ, Fierman A, Fontaine-Rothe P, Morrissey S, Prisby J, Kaufman DS, Shapiro CL, Kantoff PW, Smith MR (2002)

A phase II trial of interferon-alpha and toremifene in advanced renal cell cancer patients. Cancer Invest 20 (2): 186-191.

Oken MM, Creech RH, Tormey DC, Horton J, Davis TE, McFadden ET and Carbone PP (1982)

Toxicity and response criteria of the Eastern Cooperative Oncology Group. Am J Clin Oncol 5: 649-655.

Olayioye MA, Neve RM, Lane HA and Hynes NE (2000).

The ErbB signaling network: receptor heterodimerization in development and cancer. EMBO J 19 (13): 3159-3167.

Olencki T, Peereboom D, Wood L, Budd GT, Novick A, Finke J, McLain D, Elson P and Bukowski RM (2001)

Phase I and II trials of subcutaneously administered rIL-2, interferon alfa-2a, and fluorouracil in patients with metastatic renal carcinoma. J Cancer Res Clin Oncol 127 (5): 319-324.

Oliver RT, Nethersell AB and Bottomley JM (1989)

Unexplained spontaneous regression and alpha-interferon as treatment for metastatic renal carcinoma. Br J Urol 63: 128-131.

Olivier M, Eeles R, Hollstein M, Khan MA, Harris CC and Hainaut P (2002)

The IARC TP53 database: new online mutation analysis and recommendations to users. Hum Mutat 19: 607-614.

Olumi AF, Weidner N and Presti JC (2001)

p53 immunoreactivity correlates with Ki-67 and bcl-2 expression in renal cell carcinoma. Urol Oncol 6:63-67.

Onishi T, Machida T, Masuda F, Kurauchi H, Mori Y, Suzuki M, Iizuka N, Kondo I, Furuta N and Shirakawa H (1989) Nephrectomy in renal carcinoma with distant metastasis. Br J Urol 63 (6): 600-604.

Oosterwijk E, Debruyne FM (1995)

Radiolabeled monoclonal antibody G250 in renal-cell carcinoma. World J Urol 13: 186-190.

Palapattu GS, Pantuck AJ, Dorey F, Said JW, Figlin RA and Belldegrun AS (2003)

Collecting system invasion in renal cell carcinoma: impact on prognosis and future staging strategies. J Urol 170 (3): 768-772.

Palmer PA, Vinke J, Philip T, Négrier S, Atzpodien J, Kirchner H, Oskam R and Franks CR (1992)

Prognostic factors for survival in patients with advanced renal cell carcinoma treated with recombinant interleukin-2. Ann Oncol 3 (6): 475-480.

Paner GP, Srigley JR, Radhakrishnan A, Cohen C, Skinnider BF, Tickoo SK, Young AN and Amin MB (2006) Immunohistochemical analysis of mucinous tubular and spindle cell carcinoma and papillary renal cell carcinoma of the kidney: significant immunophenotypic overlap warrants diagnostic caution. Am J Surg Pathol 30 (1): 13-19.

Pantuck AJ, Trinh Q, Karakiewicz PI, Fergelot P, Rioux-Leclercq N, Figlin R, Said J, Belldegrun A and Patard J (2007) Use of carbonic anhydrase IX (CAIX) expression and Von Hippel Lindau (VHL) gene mutation status to predict survival in renal cell carcinoma. Journal of Clinical Oncology, ASCO Annual Meeting Proceedings Part I. Vol 25, No. 18S: 5042

Parekh DJ, Cookson MS, Chapman W, Harrell F Jr, Wells N, Chang SS and Smith JA Jr (2005)

Renal cell carcinoma with renal vein and inferior vena caval involvement: clinicopathological features, surgical techniques and outcomes. J Urol 173 (6): 1897-1902. [See comment in: J Urol 2005, 174(5):2067; author reply 2067]

Parkin DM, Whelan SL, Ferlay J, Teppo L and Thomas DB (2003)

Cancer Incidence in Five Continents Vol. VIII. The International Agency for Research on Cancer (IARC) Scientific Publication No. 155. Globocan 2002. pp.1-782.

Patard JJ, Kim HL, Lam JS, Dorey FJ, Pantuck AJ, Zisman A, Ficarra V, Han KR, Cindolo L, De La Taille A, Tostain J, Artibani W, Dinney CP, Wood CG, Swanson DA, Abbou CC, Lobel B, Mulders PF, Chopin DK and Palmer JM (1983) Role of partial nephrectomy in solitary or bilateral renal tumors. JAMA 249: 2357-2361.

Patard JJ, Kim HL, Lam JS, Dorey FJ, Pantuck AJ, Zisman A, Ficarra V, Han KR, Cindolo L, De La Taille A, Tostain J, Artibani W, Dinney CP, Wood CG, Swanson DA, Abbou CC, Lobel B, Mulders PF, Chopin DK, Figlin RA and Belldegrun AS (2004) Use of the University of California Los Angeles integrated staging system to predict survival in renal cell carcinoma: an international multicenter study. J Clin Oncol 22 (16): 3316-3322.

Patard JJ, Leray E, Rioux-Leclercq N, Cindolo L, Ficarra V, Zisman A, De La Taille A, Tostain J, Artibani W, Abbou CC, Lobel B, Guille F, Chopin DK, Mulders PF, Wood CG, Swanson DA, Figlin RA, Belldegrun AS and Pantuck AJ (2005)
Prognostic value of histologic subtypes in renal cell carcinoma: a multicenter experience. J Clin Oncol 23 (12): 2763-2771.

Pesch B, Haerting J, Ranft U, Klimpel A, Oelschlägel B, Schill W (2000)

Occupational risk factors for renal cell carcinoma: agent-specific results from a case-control study in Germany. MURC Study Group. Multicenter urothelial and renal cancer study. Int J Epidemiol 29 (6): 1014-1024.

Pestka S (1983)

The human interferons--from protein purification and sequence to cloning and expression in bacteria: before, between, and beyond. Arch Biochem Biophys. 221 (1): 1-37.

Pestka S (1997)

The human interferon-alpha species and hybrid proteins. Semin Oncol 24 (3 Suppl 9): S9-4-S9-17.

Pfeffer LM and Donner DB (1990)

The down-regulation of alpha-interferon receptors in human lymphoblastoid cells: relation to cellular responsiveness to the antiproliferative action of alpha-interferon. Cancer Res 50 (9): 2654-2657.

Phuoc NB, Ehara H, Gotoh T, Nakano M, Yokoi S, Deguchi T and Hirose Y (2007)

Immunohistochemical analysis with multiple antibodies in search of prognostic markers for clear cell renal cell carcinoma. Urology 69 (5): 843-848.

Pizzocaro G, Piva L, Colavita M, Ferri S, Artusi R, Boracchi P, Parmiani G and Marubini E (2001)

Interferon adjuvant to radical nephrectomy in Robson stages II and III renal cell carcinoma: a multicentric randomized study. J Clin Oncol 19 (2): 425-431.

Porzsolt F, Messerer D, Hautmann R, Gottwald A, Sparwasser H, Stockamp K, Aulitzky W, Moormann JG, Schumacher K, Rasche H, et al. (1988)

Treatment of advanced renal cell cancer with recombinant interferon alpha as a single agent and in combination with medroxyprogesterone acetate. A randomized multicenter trial. J Cancer Res Clin Oncol 114 (1): 95-100.

Prümmer O (1993)

Interferon-alpha antibodies in patients with renal cell carcinoma treated with recombinant interferon-alpha-2A in an adjuvant multicenter trial. The Delta-P Study Group. Cancer 71 (5): 1828-1834.

Purmonen T, Martikainen JA, Soini EJ, Kataja V, Vuorinen RL and Kellokumpu-Lehtinen PL (2008)

Economic evaluation of sunitinib malate in second-line treatment of metastatic renal cell carcinoma in Finland. Clin Ther 30 (2): 382-392.

Pyrhönen S, Salminen E, Ruutu M, Lehtonen T, Nurmi M, Tammela T, Juusela H, Rintala E, Hietanen P and Kellokumpu-Lehtinen PL (1999)

Prospective randomized trial of interferon alfa-2a plus vinblastine versus vinblastine alone in patients with advanced renal cell cancer. J Clin Oncol 17 (9): 2859-2867.

Quesada JR, Rios A, Swanson D, Trown P and Gutterman JU (1985a)

Antitumor activity of recombinant-derived interferon alpha in metastatic renal cell carcinoma. J Clin Oncol 3 (11): 1522-1528.

Quesada JR, Swanson DA and Gutterman JU (1985b)

Phase II study of interferon alpha in metastatic renal-cell carcinoma: a progress report. J Clin Oncol 3 (8): 1086-1092.

Quesada JR, Swanson DA, Trindade A and Gutterman JU (1983)

Renal cell carcinoma: antitumor effects of leukocyte interferon Cancer Res 43 (2): 940-947.

Rabinovitch RA, Zelefsky MJ, Gaynor JJ and Fuks Z (1994)

Patterns of failure following surgical resection of renal cell carcinoma: implications for adjuvant local and systemic therapy. J Clin Oncol 12 (1): 206-212.

Reich NC and Levine AJ (1984).

Growth regulation of a cellular tumor antigen, p53, in nontransformed cells. Nature 308: 199-201.

Renault PF, Hoofnagle JH, Park Y, Mullen KD, Peters M, Jones DB, Rustgi V and Jones EA (1987) Psychiatric complications of long-term interferon alfa therapy. Arch Intern Med 147 (9): 1577-1580.

Renshaw AA, Granter SR, Fletcher JA, Kozakewich HP, Corless CL and Perez-Atayde AR (1999)

Renal cell carcinomas in children and young adults: increased incidence of papillary architecture and unique subtypes. Am J Surg Pathol 23 (7): 795-802.

Repmann R. Goldschmidt AJ and Richter A (2003)

Adjuvant therapy of renal cell carcinoma patients with an autologous tumor cell lysate vaccine: a five-year follow-up analysis. Anticancer Res 23: 969-974.

Rini BI, Halabi S, Taylor J, Small EJ, Schilsky RL; Cancer and Leukemia Group B (2004)

Cancer and Leukemia Group B 90206: A randomized phase III trial of interferon-alpha or interferon-alpha plus anti-vascular endothelial growth factor antibody (bevacizumab) in metastatic renal cell carcinoma. Clin Cancer Res 10 (8): 2584-2586.

Rini BI, Weinberg V, Dunlap S, Elchinoff A, Yu N, Bok R, Simko J and Small EJ (2006)

Maximal COX-2 immunostaining and clinical response to celecoxib and interferon alpha therapy in metastatic renal cell carcinoma. Cancer 106 (3): 566-575.

Rini BI, Vogelzang NJ, Dumas MC, Wade JL 3rd, Taber DA and Stadler WM (2000)

Phase II trial of weekly intravenous gemcitabine with continuous infusion fluorouracil in patients with metastatic renal cell cancer. J Clin Oncol 18 (12): 2419-2426.

Rioux-Leclercq N, Karakiewicz PI, Trinh QD, Ficarra V, Cindolo L, de la Taille A, Tostain J, Zigeuner R, Mejean A and Patard JJ (2007)

Prognostic ability of simplified nuclear grading of renal cell carcinoma. Cancer 109 (5): 868-874.

Rioux-Leclercq N, Turlin B, Bansard J, Patard J, Manunta A, Moulinoux JP, Guille F, Ramee MP and Lobel B (2000)

Value of immunohistochemical Ki-67 and p53 determinations as predictive factors of outcome in renal cell carcinoma. Urol 2000: 55: 501-505.

Roberts WW, Bhayani SB, Allaf ME, Chan TY, Kavoussi LR and Jarrett TW (2005)

Pathological stage does not alter the prognosis for renal lesions determined to be stage T1 by computerized tomography. J Urol 173 (3): 713-715.

Robson CJ, Churchill BM and Anderson W (2002)

The results of radical nephrectomy for renal carcinoma. 1969. J Urol 167 (2 Pt 2): 873-875.

Rofsky NM, Weinreb JC, Bosniak MA, Libes RB and Birnbaum BA (1991)

Renal lesion characterization with gadolinium-enhanced MR imaging: efficacy and safety in patients with renal insufficiency. Radiology 180 (1):85-89.

Rosenberg SA, Lotze MT, Yang JC, Topalian SL, Chang AE, Schwartzentruber DJ, Aebersold P, Leitman S, Linehan WM, Seipp CA, et al. (1993)

Prospective randomized trial of high-dose interleukin-2 alone or in conjunction with lymphokine-activated killer cells for the treatment of patients with advanced cancer. J Natl Cancer Inst. Apr 21;85(8):622-32. [Erratum in: J Natl Cancer Inst 1993 85(13):1091.]

Rosenberg SA, Mulé JJ, Spiess PJ, Reichert CM and Schwarz SL (1985)

Regression of established pulmonary metastases and subcutaneous tumor mediated by the systemic administration of high-dose recombinant interleukin 2. J Exp Med 161 (5): 1169-1188.

Rosenblum SL (1987)

Paraneoplastic syndromes associated with renal cell carcinoma. J S C Med Assoc 83 (7): 375-378.

Ruijs MW, Verhoef S, Wigbout G, Pruntel R, Floore AN, de Jong D, van T Veer LJ, Menko FH (2006) Late-onset common cancers in a kindred with an Arg213Gln TP53 germline mutation. Familial Cancer 5 (2): 169-174.

Ruszczak Z and Schwartz RA (1997)

Interferons in dermatology: biology, pharmacology, and clinical applications. Adv Dermatol 13:235-288.

Sagalowsky AI, Kadesky KT, Ewalt DM and Kennedy TJ (1994)

Factors influencing adrenal metastasis in renal cell carcinoma. J Urol 151: 1181-1184.

Sagaster P, Micksche M, Flamm J and Ludwig H (1995)

Randomised study using IFN-alpha versus IFN-alpha plus coumarin and cimetidine for treatment of advanced renal cell cancer. Ann Oncol 6 (10): 999-1003.

Saitoh H (1981)

Distant metastasis of renal adenocarcinoma. Cancer 48 (6): 1487-1491.

Saitoh H, Hida M, Nakamura K, Shimbo T, Shiramizu T and Satoh T (1982a)

Metastatic processes and a potential indication of treatment for metastatic lesions of renal adenocarcinoma. J Urol 128 (5): 916-918.

Saitoh H, Nakayama M, Nakamura K and Satoh T (1982b)

Distant metastasis of renal adenocarcinoma in nephrectomized cases. J Urol 127 (6): 1092-1095.

Sali D and Boffetta P (2000)

Kidney cancer and occupational exposure to asbestos: a meta-analysis of occupational cohort studies. Cancer Causes Control 11 (1): 37-47.

Samanic C, Chow WH, Gridley G, Jarvholm B and Fraumeni JF Jr (2006)

Relation of body mass index to cancer risk in 362,552 Swedish men. Cancer Causes Control 17 (7): 901-909.

Samellas W (1963)

Adenocarcinoma of the kidney: a 10-year apparent cure following the resection of solitary pulmonary metastasis. J Urol 90: 250-252.

Sanchez-Ortiz RF, Rosser CJ, Madsen LT, Swanson DA and Wood CG (2004)

Young age is an independent prognostic factor for survival of sporadic renal cell carcinoma. J Urol 171 (6 Pt 1): 2160-2165.

Schafhauser W, Ebert A, Brod J, Petsch S and Schrott KM (1999)

Lymph node involvement in renal cell carcinoma and survival chance by systematic lymphadenectomy. Anticancer Res 19: 1573-1578.

Schmidt L, Duh FM, Chen F, Kishida T, Glenn G, Choyke P, Scherer SW, Zhuang Z, Lubensky I, Dean M, Allikmets R,

Chidambaram A, Bergerheim UR, Feltis JT, Casadevall C, Zamarron A, Bernues M, Richard S, Lips CJ, Walther MM, Tsui LC, Geil L, Orcutt ML, Stackhouse T, Lipan J, Slife L, Brauch H, Decker J, Niehans G, Hughson MD, Moch H, Storkel S, Lerman MI, Linehan WM and Zbar B (1997).

Germline and somatic mutations in the tyrosine kinase domain of the MET proto-oncogene in papillary renal carcinomas. Nat Genet 16 (1): 68-73.

Schmidt LS, Warren MB, Nickerson ML, Weirich G, Matrosova V, Toro JR, Turner ML, Duray P, Merino M, Hewitt S, Pavlovich CP, Glenn G, Greenberg CR, Linehan WM and Zbar B (2001)

Birt-Hogg-Dubé syndrome, a genodermatosis associated with spontaneous pneumothorax and kidney neoplasia, maps to chromosome 17p11.2. Am J Hum Genet 69 (4): 876-882.

Schwarz A, Vatandaslar S, Merkel S and Haller H (2007)

Renal cell carcinoma in transplant recipients with acquired cystic kidney disease. Clin J Am Soc Nephrol. 2(4):750-756.

Seliger B, Rongcun Y, Atkins D, Hammers S, Huber C, Storkel S and Kiessler R (2000).

HER-2/neu is expressed in human renal cell carcinoma at heterogenous levels independently of tumor grading and staging and can be recognized by HLA-A2.1 restricted cytotoxic T lymphocytes. Int J Cancer 87 (3): 345-359.

Selli C, Amorosi A, Vona G, Sestini R, Travaglini F, Bartoletti R and Orlando C (1997)

Retrospective evaluation of c-erbB-2 oncogene amplification using competitive PCR in collecting duct carcinoma of the kidney. J Urol 158 (1): 245-247.

Selli C, Hinshaw WM, Woodard BH and Paulson DF (1983)

Stratification of risk factors in renal cell carcinoma. Cancer 52: 899-903.

Shapiro JA, Williams MA, Weiss NS, Stergachis A, LaCroix AZ and Barlow WE (1999)

Hypertension, antihypertensive medication use, and risk of renal cell carcinoma. Am J Epidemiol; 149: 521-530.

Shiao YH, Rice JM, Anderson LM, Diwan BA and Hard GC (1998)

von Hippel-Lindau gene mutations in N-nitrosodimethylamine-induced rat renal epithelial tumors. J Natl Cancer Inst 90: 1720-1723.

Shvarts O, Seligson D, Lam J, Shi T, Horvath S, Figlin R, Belldegrun A and Pantuck AJ (2005)

p53 is an independent predictor of tumor recurrence and progression after nephrectomy in patients with localized renal cell carcinoma. J Urol 173: 725-728.

Signoretti S, Bratslavsky G, Waldman FM, Reuter VE, Haaga J, Merino M, Thomas GV, Pins MR, Libermann T, Gillespie J,

Tomaszewski JE, Compton CC, Hruszkewycz A, Linehan WM and Atkins MB (2008)
Tissue-based research in kidney cancer: current challenges and future directions. Clin Cancer Res. 2008 14 (12): 3699-3705.

Skinner DG, Colvin RB, Vermillion CD, Pfister RC and Leadbetter WF (1971)

Diagnosis and management of renal cell carcinoma. A clinical and pathologic study of 309 cases. Cancer 28: 1165-1177.

Slamon DJ, Godelphin W, Jones LA, et al (1989).

Studies of the HER2/neu protooncogene in human breast and ovarian cancer. Science 244: 707-712.

Smith I, Procter M, Gelber RD, Guillaume S, Feyereislova A, Dowsett M, Goldhirsch A, Untch M, Mariani G, Baselga J, Kaufmann M, Cameron D, Bell R, Bergh J, Coleman R, Wardley A, Harbeck N, Lopez RI, Mallmann P, Gelmon K, Wilcken N, Wist E, Sánchez Rovira P and Piccart-Gebhart MJ; HERA study team (2007)

2-year follow-up of trastuzumab after adjuvant chemotherapy in HER2-positive breast cancer: a randomised controlled trial. Lancet 369 (9555): 29-36.

Smith WL, DeWitt DL and Garavito RM (2000)

Cyclooxygenases: structural, cellular, and molecular biology. Annu Rev Biochem 69: 145-182.

Sobin L and Wittekind C (2002)

TNM Classification of Malignant Tumors, 6<sup>th</sup> edition. John Wiley & Sons, Inc, New York, p239.

Sobin LH and Fleming ID, eds (1997)

TNM classification of malignant tumors, fifth edition. Union Internationale Contre

le Cancer and the American Joint Committee. Cancer 80 (9): 1803-1804.

Soda T, Fujikawa K, Ito T, Sasaki M, Nishio Y and Miyakawa M (1999)

Volume-weighted mean nuclear volume as a prognostic factor in renal cell carcinoma. Lab Invest 79: 859-867.

Sorbellini M, Kattan MW, Snyder ME, Reuter V, Motzer R, Goetzl M, McKiernan J and Russo P (2005)

A postoperative prognostic nomogram predicting recurrence for patients with conventional clear cell renal cell carcinoma. J Urol 173 (1): 48-51.

Spanknebel K, Cheung KY, Stoutenburg J, Hurst-Wicker K, Hesdorffer C, Rn GD and Kaufman AH (2005)

Initial clinical response predicts outcome and is associated with dose schedule in metastatic melanoma and renal cell carcinoma patients treated with high-dose interleukin 2. Ann Surg Oncol 12: 381-390.

Srigley JR, Hutter RV, Gelb AB, Henson DE, Kenney G, King BF, Raziuddin S and Pisansky TM (1997)

Current prognostic factors--renal cell carcinoma: Workgroup No. 4. Union Internationale Contre le Cancer (UICC) and the American Joint Committee on Cancer (AJCC). Cancer 80: 994-996.

Stadler W M, Figlin R A, Ernstoff M S, Curti B, Pendergrass K, Srinivas S, Canfield V, Weissman C, Poulin-Costello M and Bukowski R M, on behalf of the ARCCS investigators (2007).

The Advanced Renal Cell Carcinoma Sorafenib (ARCCS) expanded access trial: Safety and efficacy in patients with non-clear cell renal cell carcinoma. J Clin Oncol 2007 ASCO Annual Meeting Proceedings Part I. 25 (18S): 5036.

#### Stein J (2006)

Incidence of brain metastasis in advanced renal carcinoma among patients randomized in a phase III trial of sorafenib, an oral multi-kinase inhibitor. Abstract 454P in ESMO book.

Stein M, Kuten A, Halpern J, Coachman NM, Cohen Y and Robinson E (1992)

The value of postoperative irradiation in renal cell cancer. Radiother Oncol 24 (1): 41-44.

Steineck G, Strander H, Carbin BE, Borgström E, Wallin L, Achtnich U, Arvidsson A, Söderlund V, Näslund I, Esposti PL and Norell SE (1990)

Recombinant leukocyte interferon alpha-2a and medroxyprogesterone in advanced renal cell carcinoma. A randomized trial. Acta Oncol 29 (2): 155-162.

Stephenson AJ, Hakimi AA, Snyder ME and Russo P (2004)

Complications of radical and partial nephrectomy in a large contemporary cohort. J Urol 171: 130-134.

Stumm G, Eberwein S, Rostock-Wolf S, Stein H, Pomer S, Schlegel J and Waldherr R (1996)

Concomitant overexpression of the EGFR and erbB-2 genes in renal cell carcinoma (RCC) is correlated with dedifferentiation and metastasis. Int J Cancer 20 (69): 17-22.

Störkel S, Eble JN, Adlakha K, Amin M, Blute ML, Bostwick DG, Darson M, Delahunt B and Iczkowski K (1997)

Classification of renal cell carcinoma: Workgroup No. 1. Union Internationale Contre le Cancer (UICC) and the American Joint Committee on Cancer (AJCC). Cancer 80: 987-989.

Störkel S, Thoenes W, Jacobi GH and Lippold R (1989)

Prognostic parameters in renal cell carcinoma - a new approach. Eur Urol 16: 416-422.

Subbaramaiah K, Telang N, Ramonetti JT, Araki R, DeVito B, Weksler BB and Dannenberg AJ (1996)

Transcription of cyclooxygenase-2 is enhanced in transformed mammary epithelial cells.

Cancer Res 56 (19): 4424-4429.

Sunela KL, Koskinen S, and Kellokumpu-Lehtinen PL (2009)

A phase-II study of combination of pegylated interferon alfa-2a and capecitabine in locally advanced or metastatic renal cell cancer. Cancer Chemother Pharmacol.

Syrjänen K and Hjelt L (1978)

Grading of human renal adenocarcinoma. Scand J Urol Nephrol 12 (1): 49-55.

Takahashi S, Shirai T, Ogawa K, Imaida K, Yamazaki C, Ito A, Masuko K and Ito N (1993)

Renal cell adenomas and carcinomas in hemodialysis patients: relationship between hemodialysis period and development of lesions. Acta Pathol Jpn 43 (11): 674-682.

Takahashi S, Tanigawa T, Imagawa M, Mimata H, Nomura Y and Ogata J (1994)

Interferon as adjunctive treatment for non-metastatic renal cell carcinoma. Br J Urol 74 (1): 11-14.

Takahashi Y, Harashima N, Kajigaya S, Yokoyama H, Cherkasova E, McCoy JP, Hanada K, Mena O, Kurlander R, Tawab A, Srinivasan R, Lundqvist A, Malinzak E, Geller N, Lerman MI and Childs RW (2008)

Regression of human kidney cancer following allogeneic stem cell transplantation is associated with recognition of an HERV-E antigen by T cells. J Clin Invest 118 (3): 1099-1109. [Erratum in: J Clin Invest Apr;118(4):1584.]

Takashi M, Takagi Y, Sakata T, Shimoji T and Miyake K (1995)

Prognostic factors in renal cancer. Urol Clin North Am 20: 247-262.

Taketo MM (1998)

Cyclooxygenase-2 inhibitors in tumorigenesis (part I). J Natl Cancer Inst 90: 1529-1536.

Tan X, Zhai Y, Chang W, Hou J, He S, Lin L, Yu Y, Xu D, Xiao J, Ma L, Wang G, Cao T and Cao G (2008)

Global analysis of metastasis-associated gene expression in primary cultures from clinical specimens of clear-cell renal-cell carcinoma. Int J Cancer 123 (5): 1080-1088.

Tannir NM, Thall PF, Ng CS, Wang X, Wooten L, Siefker-Radtke A, Mathew P, Pagliaro L, Wood C and Jonasch E (2008) A phase II trial of gemcitabine plus capecitabine for metastatic renal cell cancer previously treated with immunotherapy and targeted agents. J Urol 180 (3): 867-872; discussion 872.

Thoenes W, Störkel S and Rumpelt HJ (1986)

Histopathology and classification of renal cell tumors (adenomas, oncocytomas and carcinomas). The basic cytological and histomorphological elements and their use for diagnostics. Pathol Res Pract 181: 125-143.

Thomas AA, Rini BI, Lane BR, Garcia J, Dreicer R, Klein EA, Novick AC and Campbell SC (2009)

Response of the primary tumor to neoadjuvant sunitinib in patients with advanced renal cell carcinoma. J Urol 181 (2): 518-523.

Thrasher JB and Paulson DF (1993)

Prognostic factors in renal cancer. Int Urol Nephrol. 27 (1): 1-8.

Thödtmann R, Sauter T, Weinknecht S, Weissbach L, Blatter J, Ohnmacht U and Hanauske A (2003) A phase II trial of pemetrexed in patients with metastatic renal cell cancer. Invest New Drugs 21: 353-358.

Tosaka A, Ohya K, Yamada K, Ohashi H, Kitahara S, Sekine H, Takehara Y and Oka K (1990)

Incidence and properties of renal masses and asymptomatic renal cell carcinoma detected by abdominal ultrasonography. J Urol 144: 1097-1099.

Trinchieri G, Matsumoto-Kobayashi M, Clark SC, Seehra J, London L, Perussia B (1984)

Response of resting human peripheral blood natural killer cells to interleukin 2. J Exp Med 160 (4): 1147-1169.

Trinchieri G. Peritt D and Gerosa F (1996)

Acute induction and priming for cytokine production in lymphocytes. Cytokine Growth Factor Rev 7 (2): 123-132.

Trump DL, Elson PJ, Borden EC, Harris JE, Tuttle RL, Whisnant JK, Oken MM, Carignan JR, Ruckdeschel JC and Davis TE (1987) High-dose lymphoblastoid interferon in advanced renal cell carcinoma: an Eastern Cooperative Oncology Group Study. Cancer Treat Rep 71 (2): 165-169.

Tsui KH, Shvarts O, Smith RB, Figlin R, de Kernion JB and Belldegrun A (2000b)

Renal cell carcinoma: prognostic significance of incidentally detected tumors. J Urol 163 (2): 426-430.

Tsui KH, Shvarts O, Smith RB, Figlin RA, de Kernion JB and Belldegrun A (2000a)

Prognostic indicators for renal cell carcinoma. A multivariate analysis of 643 patients using the revised 1997 TNM staging criteria. J Urol 163: 1090-1095.

Tsujii M and DuBois RN (1995)

Alterations in cellular adehesion and apoptosis in epithelial cells overexpressing prostaglandin endoperoxide synthase 2. Cell 83: 493-501.

Tuna B, Yorukoglu K, Gurel D, Mungan U and Kirkali Z (2004)

Significance of COX-2 expression in human renal cell carcinoma. Urology 2004, 64(6):1116-1120.

Tykkä H, Oravisto KJ, Lehtonen T, Sarna S and Tallberg T (1978)

Active specific immunotherapy of advanced renal-cell carcinoma. Eur Urol 4 (4): 250-258.

Uchida T, Gao JP, Wang C, Jiang SX, Muramoto M, Satoh T, Minei S, Shimura S, Irie A, Kameya T and Baba S (2002) Clinical significance of p53, mdm2, and bcl-2 proteins in renal cell carcinoma. Urology: 59: 615-620.

Uhlman DL, Nguyen PL, Manivel JC, Aeppli D, Resnick JM, Fraley EE, Zhang G and Niehans GA (1994)

Association of immunohistochemical staining for p53 with metastatic progression and poor survival in patients with renal cell carcinoma. J Natl Cancer Inst 86: 1470-1475.

Umeda T and Niijima T (1986)

Phase II study of alpha interferon on renal cell carcinoma. Summary of three collaborative trials. Cancer Sep 58 (6): 1231-1235.

Usubutyn A, Uygur MC, Ayhan A, Toklu C, Sahin A, Ozen H and Ruacan S (1998)

Comparison of grading systems for estimating the prognosis of renal cell carcinoma. Int. Urol. Nephrol. 30:391-397.

Wagner JR, Walther MM, Linehan WM, White DE, Rosenberg SA and Yang JC (1999)

Interleukin-2 based immunotherapy for metastatic renal cell carcinoma with the kidney in place. J Urol 162 (1): 43-45.

Wallace PK, Howell AL, Fanger MW (1994)

Role of Fc gamma receptors in cancer and infectious disease. J Leukoc Biol 55 (6): 816-826.

van der Werf-Messing B (1973)

Proceedings: Carcinoma of the kidney. Cancer 32 (5):1056-1061.

van Houwelingen KP, van Dijk BA, Hulsbergen-van de Kaa CA, Schouten LJ, Gorissen HJ, Schalken JA, van den Brandt PA and Oosterwijk E (2005)

Prevalence of von Hippel-Lindau gene mutations in sporadic renal cell carcinoma: results from The Netherlands cohort study. BMC Cancer 5:57.

van Kessel AG, Wijnhoven H, Bodmer D, Eleveld M, Kiemeney L, Mulders P, Weterman M, Ligtenberg M, Smeets D and Smits A (1999)

Renal cell cancer: chromosome 3 translocations as risk factors. J Natl Cancer Inst 91 (13): 1159-1160.

Van Poppel H, Vandendriessche H, Boel K, Mertens V, Goethuys H, Haustermans K, Van Damme B and Baert L (1997) Microscopic vascular invasion is the most relevant prognosticator after radical nephrectomy for clinically nonmetastatic renal cell carcinoma. J. Urol 158: 45-49.

Washio M, Mori M, Khan M, Sakauchi F, Watanabe Y, Ozasa K, Hayashi K, Miki T, Nakao M, Mikami K, Ito Y, Kubo T, Wakai K and Tamakoshi A; JACC Study Group (2007)

Diabetes mellitus and kidney cancer risk: the results of Japan Collaborative Cohort Study for Evaluation of Cancer Risk (JACC Study). Int J Urol 14 (5): 393-397.

Wenzel C, Locker GJ, Bartsch R, Pluschnig U, Mader R, Hussian D, Kramer G, Marberger M, Lintner C, Rauchenwald M, Zielinski CC and Steger GG (2003)

Capecitabine monotherapy and in combination with immunotherapy in the treatment of metastatic renal cell carcinoma. Anticancer Drugs 14: 779-784.

Whang M, O'Toole K, Bixon R, Brunetti J, Ikeguchi E, Olsson CA, Sawczuk TS and Benson MC (1995)

The incidence of multifocal renal cell carcinoma in patients who are candidates for partial nephrectomy. J Urol 154 (3): 968-970; discussion 970-971.

Viaud S, Terme M, Flament C, Taieb J, André F, Novault S, Escudier B, Robert C, Caillat-Zucman S, Tursz T, Zitvogel L and Chaput N (2009)

Dendritic cell-derived exosomes promote natural killer cell activation and proliferation: a role for NKG2D ligands and IL-15Ralpha. PLoS One 4 (3):e4942.

Wirth MP (1993)

Immunotherapy for metastatic renal cell carcinoma. Urol Clin North Am 20 (2): 283-295.

Witte RS, Leong T, Ernstoff MS, Krigel RL, Oken MM, Harris J, Tormey DC and Trump DL (1995)

A phase II study of interleukin-2 with and without beta-interferon in the treatment of advanced renal cell carcinoma. Invest New Drugs 13 (3): 241-247.

Vogelzang NJ, Lipton A and Figlin RA (1993)

Subcutaneous interleukin-2 plus interferon alfa-2a in metastatic renal cancer: an outpatient multicenter trial. J Clin Oncol 11: 1809-1816

Wolk A, Lindblad P and Adami HO (1996)

Nutrition and renal cell cancer. Cancer Causes Control 7 (1): 5-18.

Wronski M, Maor MH, Davis BJ, Sawaya R and Levin VA (1997)

External radiation of brain metastases from renal carcinoma: a retrospective study of 119 patients from the M. D. Anderson Cancer Center. Int J Radiat Oncol Biol Phys 37 (4): 753-759.

Wunderlich H, Dreihaupt M, Schlichter A, Kosmehl H, Reichelt O and Schubert J (2004)

New cut-off point between T1 and T2 renal cell carcinoma - necessary for a better discriminatory power of the TNM classification. Urol Int. 72 (2): 123-128.

Vuoristo M, Jantunen I, Pyrhönen S, Muhonen T and Kellokumpu-Lehtinen P (1994)

A combination of subcutaneous recombinant interleukin-2 and recombinant interferon-alpha in the treatment of advanced renal cell carcinoma or melanoma. Eur J Cancer 30A (4): 530-532.

Yang JC, Haworth L, Sherry RM, Hwu P, Schwartzentruber DJ, Topalian SL, Steinberg SM, Chen HX and Rosenberg SA (2003a) A randomized trial of bevacizumab, an anti-vascular endothelial growth factor antibody, for metastatic renal cancer. N Engl J Med 349 (5): 427-434.

Yang JC, Sherry RM, Steinberg SM, Topalian SL, Schwartzentruber DJ, Hwu P, Seipp CA, Rogers-Freezer L, Morton KE, White DE, Liewehr DJ, Merino MJ and Rosenberg SA (2003b)

Randomized study of high-dose and low-dose interleukin-2 in patients with metastatic renal cancer. J Clin Oncol 21 (16): 3127-3132.

Yang XJ, Tan MH, Kim HL, Ditlev JA, Betten MW, Png CE, Kort EJ, Futami K, Furge KA, Takahashi M, Kanayama HO, Tan PH, Teh BS, Luan C, Wang K, Pins M, Tretiakova M, Anema J, Kahnoski R, Nicol T, Stadler W, Vogelzang NG, Amato R, Seligson D, Figlin R, Belldegrun A, Rogers CG and Teh BT (2005)

A molecular classification of papillary renal cell carcinoma. Cancer Res 65 (13): 5628-5637.

Yildiz E, Gokce G, Kilicarslan H, Ayan S, Goze OF and Gultekin EY (2004)

Prognostic value of the expression of Ki-67, CD44 and vascular endothelial growth factor, and microvessel invasion, in renal cell carcinoma. BJU Int 93: 1087-1093.

Yoshimura R, Matsuyama M, Kawahito Y, Tsuchida K, Kuratsukuri K, Takemoto Y, Mitsuhashi M, Sano H and Nakatani T (2004) Study of cyclooxygenase-2 in renal cell carcinoma. Int J Mol Med 13 (2): 229-233.

Yuan JM, Castelao JE, Gago-Dominguez M, Ross RK and Yu MC (1998)

Hypertension, obesity and their medications in relation to renal cell carcinoma. Br J Cancer 77: 1508-1513.

Yuan JM, Gago-Dominguez M, Castelao JE, Hankin JH, Ross RK and Yu MC (1998) Cruciferous vegetables in relation to renal cell carcinoma. Int J Cancer 77 (2): 211-216.

Zav'Yalov VP and Zav'Yalova GA (1997)

Interferons alpha/beta and their receptors: place in the hierarchy of cytokines. APMIS. 1997 105 (3): 161-186.

Zbar B, Kaelin W, Maher E and Richard S (1999)

Third International Meeting on von Hippel-Lindau disease. Cancer Res 59 (9): 2251-2253.

Zbar B, Kishida T, Chen F, Schmidt L, Maher ER, Richards FM, Crossey PA, Webster AR, Affara NA, Ferguson-Smith MA, Brauch H, Glavac D, Neumann HP, Tisherman S, Mulvihill JJ, Gross DJ, Shuin T, Whaley J, Seizinger B, Kley N, Olschwang S, Boisson C, Richard S, Lips CH and Lerman M (1996)

Germline mutations in the Von Hippel-Lindau disease (VHL) gene in families from North America, Europe, and Japan. Hum Mutat 8 (4): 348-57.

Zhang D, Okada S, Yu Y, Zheng P, Yamaguchi R and Kasai H (1997)

Vitamin E inhibits apoptosis, DNA modification, and cancer incidence induced by iron-mediated peroxidation in Wistar rat kidney. Cancer Res 57 (12): 2410-2414. Nutrition and renal cell cancer.

Zigeuner R, Ratschek M, Rehak P, Schips L and Langner C (2004)

Value of p53 as a prognostic marker in histologic subtypes of renal cell carcinoma: a sytematic analysis of primary and metastatic tumor tissue. Urology 63: 651-655.

Zisman A, Chao DH, Pantuck AJ, Kim HJ, Wieder JA, Figlin RA, Said JW and Belldegrun AS (2002a) Unclassified renal cell carcinoma: clinical features and prognostic impact of a new histological subtype. J Urol 168: 950-955.

Zisman A, Pantuck AJ, Chao D, Dorey F, Said JW, Gitlitz BJ, de Kernion JB, Figlin RA and Belldegrun AS (2001a) Re-evaluation of the 1997 TNM classification for RCC: T1 and T2 cut-off point at 4.5 cm rather than 7 cm better correlates with clinical outcome. J Urol 166: 54-58.

Zisman A, Pantuck AJ, Dorey F, Chao DH, Gitlitz BJ, Moldawer N, Lazarovici D, deKernion JB, Figlin RA and Belldegrun AS (2002b) Mathematical model to predict individual survival for patients with renal cell carcinoma. J Clin Oncol 20: 1368-1374.

Zisman A, Pantuck AJ, Dorey F, Said JW, Shvarts O, Quintana D, Gitlitz BJ, deKernion JB, Figlin RA and Belldegrun AS (2001b) Improved prognostication of RCC using an integrated staging system. J Clin Oncol 19: 1649-1657.