

# Sex Comparison of White-Blood-Cell Responses to Acute Vigorous-Intensity Aerobic Exercise

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**Purpose:** It is widely accepted that exercise alters the number and function of circulating white blood cells, and many factors are suggested to influence this phenomenon. Previous study populations have primarily been men, so this study aimed to compare mobilization of white blood cells between sexes after an acute exercise bout. **Methods:** The study included healthy and physically active participants, 10 men (age 26 [SD 7] y) and 10 women (age 33 [SD 8] y). Participants performed a 30-minute cycling ergometer exercise session at 70% maximal oxygen uptake, and blood samples were drawn at rest and at 3 minutes, 1 hour, and 2 hours postexercise. **Results:** Changes in total leukocyte, neutrophil, lymphocyte, monocyte, eosinophil, and basophil counts were analyzed and corrected for plasma volume change. Acute exercise significantly increased the number of total leukocytes and all leukocyte subpopulations excluding eosinophils in both women and men. The mobilization of all leukocyte subpopulations was similar in both sexes, but mobilization of total leukocytes, as a percentage change from baseline, was significantly greater in men (52% [SD 28%] in men, 19% [SD 23%] in women,  $P = .024$ , Cohen  $d = 1.06$ ). **Conclusions:** These findings indicate that the sex difference in white-blood-cell mobilization in response to vigorous acute exercise is small and was only evident in total leukocyte response in the present study.

**Keywords:** gender, immunology, leukocyte, immune cells, mobilization, physical activity

Leukocytosis, the increase in blood leukocyte count, following acute exercise was first reported in 1893 by Schulte.<sup>1</sup> Since then, a variety of studies have proven that exercise induces physiological changes in the immune system.<sup>2,3</sup> During acute exercise white blood cells, especially neutrophils and lymphocytes, migrate from their storage organs and endothelial walls into the circulation, causing an elevation in white blood cell count.<sup>4</sup> The increase in white blood cell count is a transient phenomenon, as the number of all leukocyte subsets usually decreases back to resting values within minutes to 24 hours after acute exercise.<sup>5</sup> This exercise-induced mobilization of white blood cells is considered as part of the “fight or flight” response, where the immune system prepares the body to find infected or malignant cells and combat potential injury by redistributing cells to sites of injury or increased physical

stress. The extent of the effects of immune cell mobilization is unknown, but it has been suggested that it is an important phenomenon in, for example, cancer prevention and control.<sup>6,7</sup>

It is well known that the intensity of exercise influences the mobilization of white blood cells.<sup>8,9</sup> In addition, it has been suggested that several factors, such as age, training status, nutritional status, and history of infections may alter the immune response to acute exercise, although the data have been conflicting.<sup>10,11</sup> The information we know today about the immune responses to exercise comes mainly from studies with male participants<sup>2,8,9,12–15</sup> and the data are subsequently generalized to females. To the best of our knowledge, the effect of sex on white blood cell mobilization has been studied relatively little. Some previous studies have observed sex differences in white blood cell responses to acute exercise,<sup>16,17</sup> but there are also studies reporting no change between sexes.<sup>18,19</sup> Furthermore, the observed sex differences have not been similar between the studies, as some report greater immune responses in men, and some in women.<sup>16,17</sup> For example, Morgado et al<sup>16</sup> reported greater lymphocyte response in men compared with women at any menstrual cycle phase after a maximal incremental swimming exercise, and Timmons et al<sup>17</sup> reported similar neutrophil, lymphocyte, and monocyte responses between men and women at any menstrual cycle phase who did not use oral contraceptives after a 90-minute cycling exercise. Furthermore, Timmons et al<sup>17</sup> found that immune cell responses were greater in women using oral contraceptives during luteal phase than in men.

In general, for many years, biomedical research has been conducted only with men, but recently that has been changing.<sup>20</sup> Inclusion of women and sex-difference studies are crucial to a deeper understanding of human health, and immunological aspects are important especially in the prevention of infections and many

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diseases. For example, it has been suggested that possible sex differences in immune system responses to exercise could partly influence exercise benefits on brain and neurodegenerative diseases through neuroinflammation.<sup>21</sup> Furthermore, knowing the biological sex differences in physiological responses such as exercise stimuli gives the sexes a better understanding of how their bodies adapt and recover from exercise. Consequently, the aim of the present study was to narrow the gender gap in research and to examine if the white blood cell mobilization following a single bout of acute exercise varies between sexes. Seventy percent of  $\text{VO}_2\text{max}$  was selected for exercise intensity to induce substantial physiological responses. We hypothesized that the exercise bout transiently changes individuals' white blood cell counts. Furthermore, we hypothesized that these changes may show some differences between the sexes, but we were unsure which sex would have greater immunological responses to exercise due to conflicting results in previous literature.<sup>16–19</sup>

## Materials and Methods

This study was conducted as an investigator initiated academic study at the Turku University Hospital and Turku PET Center, Turku, Finland between November 2020, and July 2021. Informed consent was obtained from all participants and good clinical practice was followed according to the principles of the Declaration of Helsinki. The study was approved by the Ethics Committee of the Hospital District of Southwestern Finland. This study is a part of a larger trial (Clinicaltrials.gov ID NCT04255758).

### Participants

A total of 20 (10 men, 10 women) healthy and physically active participants were recruited to the study. Inclusion criteria were the following: age 18–45 years, body mass index 18 to 30  $\text{kg}/\text{m}^2$ , and resting blood pressure under 140/90 mm Hg. The exclusion criteria were the following: history of a cardiac event, medically treated diabetes, abundant use of alcohol, use of narcotics, smoking of tobacco or consuming of snuff tobacco, diagnosed depressive or bipolar disorder, abnormalities in resting ECG, pregnancy, and any chronic disease or condition that could create a hazard to the participant's safety, endanger the study procedures, or interfere with the interpretation of study results. Three of the female participants reported using hormonal contraceptives, but the type of the contraceptives were not documented. Women's menstrual cycle phase during the study was not documented and therefore not controlled for.

### Study Design

During participant screening, body mass index and waist circumference were measured, and body fat percentage estimated using Durmin and Womersley<sup>22</sup> 4-site skinfold method. Fat-free mass was determined by subtracting fat mass from total body weight. Participants also filled out questionnaires about their use of medication and regular weekly exercise training (Table 1). Maximal rate of oxygen consumption ( $\text{VO}_2\text{max}$ ) was measured with Vyntus CPX (Vyair Medical Inc) in an incremental cycle ergometry exercise test starting at 50 W and followed by 25 W increments every 2 minutes. The criteria for  $\text{VO}_2\text{max}$  were a respiratory exchange ratio  $>1.1$ , lactate concentration over 10 mmol/L, and a heart rate  $\leq 10$  beats per minutes of the age-predicted maximal heart rate, as calculated by the Tanaka formula.<sup>23</sup> Within the following 2 weeks, the participants

performed a personalized acute exercise bout on cycle ergometer with a total duration of 30 minutes, consisting of 5-minute warm-up at power equivalent to 50%  $\text{VO}_2\text{max}$  and 25-minute work at 70%  $\text{VO}_2\text{max}$ , intensity which would be categorized as "vigorous."<sup>24</sup> The chosen intensity level was designed to be challenging, promoting significant physiological responses, but not so high as to lead to exhaustion over the 30-minute period. The study was always conducted in the afternoon and participants were asked to refrain from strenuous exercise, and abstain from alcohol, as well as from food and caffeine for 24 hours prior to the study. The participants were all used to bicycling exercise. Heart rate (PalmSats 2500) and blood pressure (Apteq AE701f, Rossmax Swiss GmbH) were measured after short supine rest before and after the exercise, and during the exercise on ergometer. Before exercise, an intravenous catheter was inserted at the antecubital area of the arm for repeated blood sampling. A continuous, slow saline infusion (30 drops/min) was used to keep the catheter from clotting during the study. 10 mL of blood was collected to EDTA tubes (BD Biosciences) at 4 different time points: at rest, 3 minutes, 1 hour, and 2 hours after the exercise. The 2-hour time after the acute exercise was spent in supine rest. Lactate concentration was measured at each time point with Lactate Scout 4 (EKF Diagnostics). One participant could not finish the complete 30-minute exercise but was included in the analyses because the other measured parameters were not significantly different from the others, and he could still cycle until the 25-minute time point.

### Blood-Sample Analysis

Blood samples were analyzed at the Turku University Hospital Laboratory. Total white blood cell count and the number of neutrophils, lymphocytes, monocytes, eosinophils, and basophils were analyzed using flow cytometry method (Sysmex XN analyser, Sysmex Corporation).

Change in plasma volume (PV) after exercise bout was taken into consideration and determined from hematocrit (Hct) and hemoglobin (Hb) with the following formula:  $\Delta PV = (\text{Hb}_{\text{pre}} \times [1 - \text{Hct}_{\text{post}}]) / (\text{Hb}_{\text{post}} \times [1 - \text{Hct}_{\text{pre}}]) - 1$ .<sup>25</sup> To correct the total amount of circulating white blood cells (WBC) for plasma volume change, the following formula was used:  $\text{WBC}_{\text{corrected}} = \text{WBC}_{\text{uncorrected}} \times (1 + \Delta PV)$ .<sup>25</sup>

### Statistical Methods

Two-tailed *t* test was used to analyze sex-differences in baseline characteristics and in baseline blood variables (results in Table 1). Repeated-measures 2-way analysis of variance was used to test the effect of exercise on white blood cells counts and on plasma volume in men and in women. The main effects were sex, time, and sex  $\times$  time interaction (Results in Figures 1–3). When the main effect(s) was significant at less than  $P < .05$ , statistical differences were considered by Tukey post hoc test. Statistical analyses were performed with SAS (version 9.4), Graphpad prism (version 8.0), and SPSS Statistics (version 27).

## Results

### Baseline Characteristics

Baseline characteristics of study participants are presented in Table 1. Mean age was 26 (SD 7) years in men and 33 (SD 8) years in women, mean body mass index was 25.5 (SD 2.1)  $\text{kg}/\text{m}^2$  in men and 24.6 (SD 2.4)  $\text{kg}/\text{m}^2$  in women. As anticipated, men

**Table 1 Baseline Characteristics of the Study Participants by Sex**

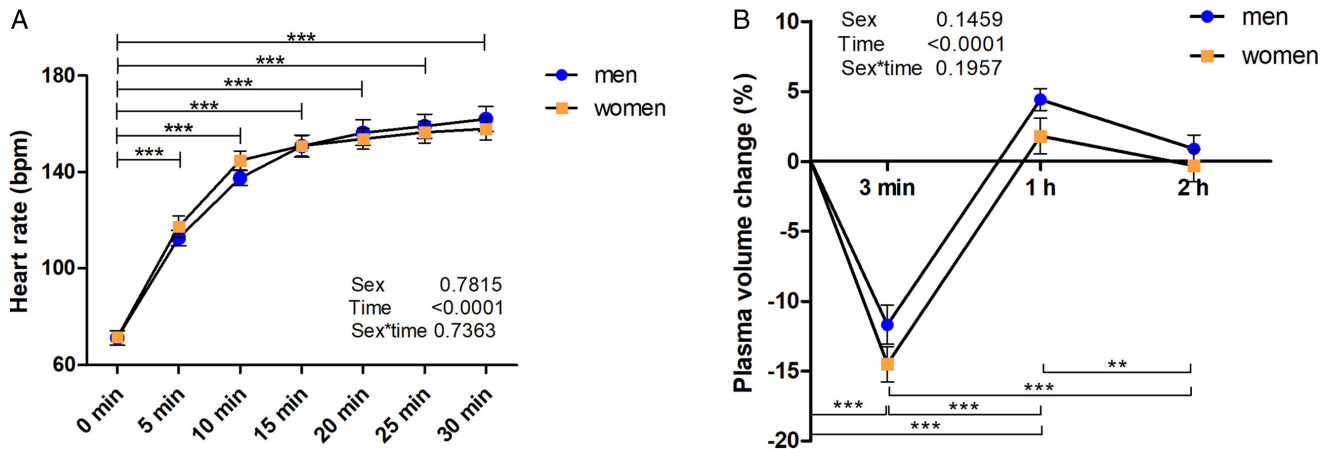
	Men	Women	P
n	10	10	
Age, y	26 (7)	33 (8)	.064
Anthropometrics			
Height, cm	179.5 (4.5)	165.4 (6.0)	<.001
BM, kg	82.4 (10.1)	67.4 (7.3)	.002
Body fat, %	15.7 (3.7)	27.6 (3.2)	<.001
FFM, kg	69.3 (6.9)	48.7 (4.7)	<.001
BMI, kg/m <sup>2</sup>	25.5 (2.1)	24.6 (2.4)	.393
Waist circumference, cm	84.8 (11.3)	79.7 (10.8)	.320
Health measurements			
Systolic blood pressure, mm Hg	117 (6)	112 (6)	.109
Diastolic blood pressure, mm Hg	60 (9)	67 (5)	.046
Baseline heart rate, beats/min	71 (9)	72 (5)	.931
Baseline blood analytes			
Total leukocytes, 10 <sup>9</sup> /L	5.53 (0.79)	6.99 (2.0)	.044
Neutrophils, 10 <sup>9</sup> /L	2.93 (0.84)	3.88 (1.87)	.162
Lymphocytes, 10 <sup>9</sup> /L	1.88 (0.31)	2.44 (0.51)	.009
Monocytes, 10 <sup>9</sup> /L	0.49 (0.13)	0.51 (0.13)	.716
Eosinophils, 10 <sup>9</sup> /L	0.20 (0.16)	0.12 (0.04)	.167
Basophils, 10 <sup>9</sup> /L	0.04 (0.02)	0.04 (0.02)	.351
Erythrocytes, 10 <sup>12</sup> /L	4.70 (0.22)	4.09 (0.24)	<.001
Hemoglobin, g/L	145.1 (6.2)	124.5 (6.5)	<.001
Hematocrit, %	0.42 (0.02)	0.37 (0.02)	<.001
Mean corpuscular volume, fL	89.6 (2.1)	91.1 (2.9)	.897
Mean corpuscular hemoglobin, pg	30.8 (1.1)	30.3 (1.3)	.181
Thrombocytes, 10 <sup>9</sup> /L	256.6 (25.7)	269.9 (48.5)	.772
Lactate, mmol/L	0.91	0.94	.825
Regular weekly exercise training			
Times/wk	4 (2)	3 (3)	.472
Hours/wk	6.3 (3.0)	4.6 (3.8)	.281
Fitness-test measurements			
VO <sub>2</sub> max, mL/kg BM/min	45.1 (4.5)	40.1 (4.0)	.022
VO <sub>2</sub> max, mL/kg FFM/min	53.5 (4.7)	55.4 (4.8)	.405
Maximal heart rate, beats/min	197 (9)	187 (8)	.031
Maximal lactate, mmol/L	12.8 (1.9)	12.0 (1.8)	.277
Maximal power, W	280 (41)	209 (34)	<.001
Maximal power, W/kg	3.6 (0.4)	3.3 (0.4)	.124

Abbreviations: BM, body mass; BMI, BM index; FFM, fat-free mass; VO<sub>2</sub>max, maximal oxygen consumption. Note: Data are presented as mean (SD). Sex difference in 2-tailed *t* test.

were significantly taller and had higher body mass and fat-free mass ( $P < .001$ ,  $P = .002$ ,  $P < .001$ , respectively). Women had significantly higher body fat percentage ( $P < .001$ ) and diastolic blood pressure ( $P = .046$ ) at baseline compared with men. Men had higher VO<sub>2</sub>max relative to total body mass ( $P = .022$ ), and higher maximal heart rate and maximal pedaling power in the fitness test ( $P = .031$ ,  $P < .001$ , respectively). When normalized by fat-free mass, VO<sub>2</sub>max was similar between sexes. Moreover, the absolute number of leukocytes and lymphocytes at baseline was significantly higher in women compared with men ( $P = .044$ ,  $P = .009$ , respectively). There was no sex difference in other baseline characteristics (Table 1).

### Lactate and Circulatory Responses to Acute Exercise

Lactate and heart rate responses to exercise were similar in men and women. Lactate concentration at 3-minute postexercise was on average 3.8 (SD 3.1) mmol/L in men and 3.1 (SD 2.2) mmol/L in women, while the resting values were 0.9 (SD 0.4) mmol/L and 0.9 (SD 0.2) mmol/L, respectively. Heart rates during the 30-minute exercise are presented in Figure 1A. Mean heart rate was 146 (SD 12) in men and 146 (SD 13) in women. Mean heart rate percentage of maximal heart rate during the exercise was 73.0% (SD 4.3%) and 74.4% (SD 12.7%), in men and women, respectively, and at



**Figure 1** — (A) Heart rate during 30-minute exercise at 70% of maximal oxygen uptake and (B) mean plasma volume change after the exercise. \*\* $P < .01$ . \*\*\* $P < .001$ .

the end of the exercise, the heart rate percentage of maximal heart rate was 82.3% (SD 6.6%) and 84.3% (SD 6.0%), in men and women, respectively. The reduction in plasma volume at 3-minute postexercise compared with baseline was 11.7% (SD 4.4%) in men and 14.5% (SD 3.9%) in women ( $P < .001$ ). The plasma volume was 4.4% (SD 2.5%) above baseline in men ( $P < .001$ ) and 1.8% (SD 4.0%) above baseline in women at 1 hour postexercise, and at 2 hours postexercise the plasma volume was 0.9% (SD 3.1%) above baseline in men and 0.3% (SD 3.7%) below baseline in women (Figure 1B). There was no sex difference in plasma volume change.

### Immune-Cell Response to Acute Exercise

Total leukocyte count increased significantly at 3 minutes postexercise ( $P < .001$ , Cohen  $d = 0.85$ ) and remained elevated at 1 hour and 2 hours postexercise similarly in both sexes ( $P < .05$ , Cohen  $d = 0.44$ ,  $P < .001$ , Cohen  $d = 0.94$ , respectively). There were, however, 2 peaks as the total leukocyte count decreased significantly at 1 hour postexercise compared to 3 minutes postexercise ( $P < .01$ , Cohen  $d = 0.45$ ) and then again increased significantly at 2 hours postexercise compared with 1 hour postexercise in both sexes ( $P < .001$ , Cohen  $d = 0.48$ ; Figure 2A). The percentage changes in total leukocytes were similar to the changes in absolute cell counts. There was significant sex  $\times$  time interaction in the percentage change of total leukocytes ( $P = .0237$ , Cohen  $d = 1.06$ ) as men had higher percentage increase at 2 hours postexercise from baseline compared to women (Figure 2D). However, there was no other significant sex  $\times$  time interaction in any of the leukocyte subsets studied. Neutrophil count increased significantly at 3 minutes postexercise ( $P < .01$ , Cohen  $d = 0.51$ ), remained elevated at 1 hour postexercise ( $P < .001$ , Cohen  $d = 0.74$ ), and reached maximal values at 2 hours postexercise in both sexes ( $P < .001$ , Cohen  $d = 1.07$ ; Figure 2B). Lymphocyte count increased significantly at 3 minutes postexercise compared to baseline ( $P < .001$ , Cohen  $d = 1.11$ ) and decreased below baseline at 1 hour postexercise in both sexes ( $P < .001$ , Cohen  $d = 0.87$ ; Figure 2C). The percentage of neutrophils of total leukocytes increased significantly at 1 hour and 2 hours compared with baseline ( $P < .001$ , Cohen  $d = 1.07$ ,  $P < .001$ , Cohen  $d = 1.14$ , respectively; Figure 2E), whereas the percentage of lymphocytes decreased at 1 hour and 2 hours postexercise compared to baseline ( $P < .001$ , Cohen  $d = 1.02$ ,  $P < .001$ , Cohen  $d = 1.07$ , respectively; Figure 2F).

Monocytes showed 2 increases in both sexes. First, monocyte count increased at 3 minutes postexercise compared to baseline ( $P < .001$ , Cohen  $d = 0.76$ ) and after returning to baseline at 1 hour postexercise, monocyte count increased at 2 hours postexercise compared to 1 hour postexercise ( $P < .01$ , Cohen  $d = 0.32$ ; Figure 3A). Eosinophil count did not change significantly at 3 minutes postexercise compared with baseline but decreased significantly at 1 hour and 2 hours postexercise compared with 3 minutes postexercise in both sexes ( $P < .01$ , Cohen  $d = 0.25$ ,  $P < .01$ , Cohen  $d = 0.27$ , respectively; Figure 3B). Basophil count increased at 3 minutes postexercise ( $P < .001$ , Cohen  $d = 0.78$ ) and decreased back to baseline at 1 hour postexercise ( $P < .001$ , Cohen  $d = 0.22$ ; Figure 3C). Percentages of monocytes, eosinophils, and basophils of total leukocytes decreased significantly after exercise compared to baseline, and were lowest at 2 hours postexercise ( $P < .001$ , Cohen  $d = 0.89$ ,  $P < .05$ , Cohen  $d = 0.54$ ,  $P < .01$ , Cohen  $d = 0.52$ , respectively; Figure 3D–3F).

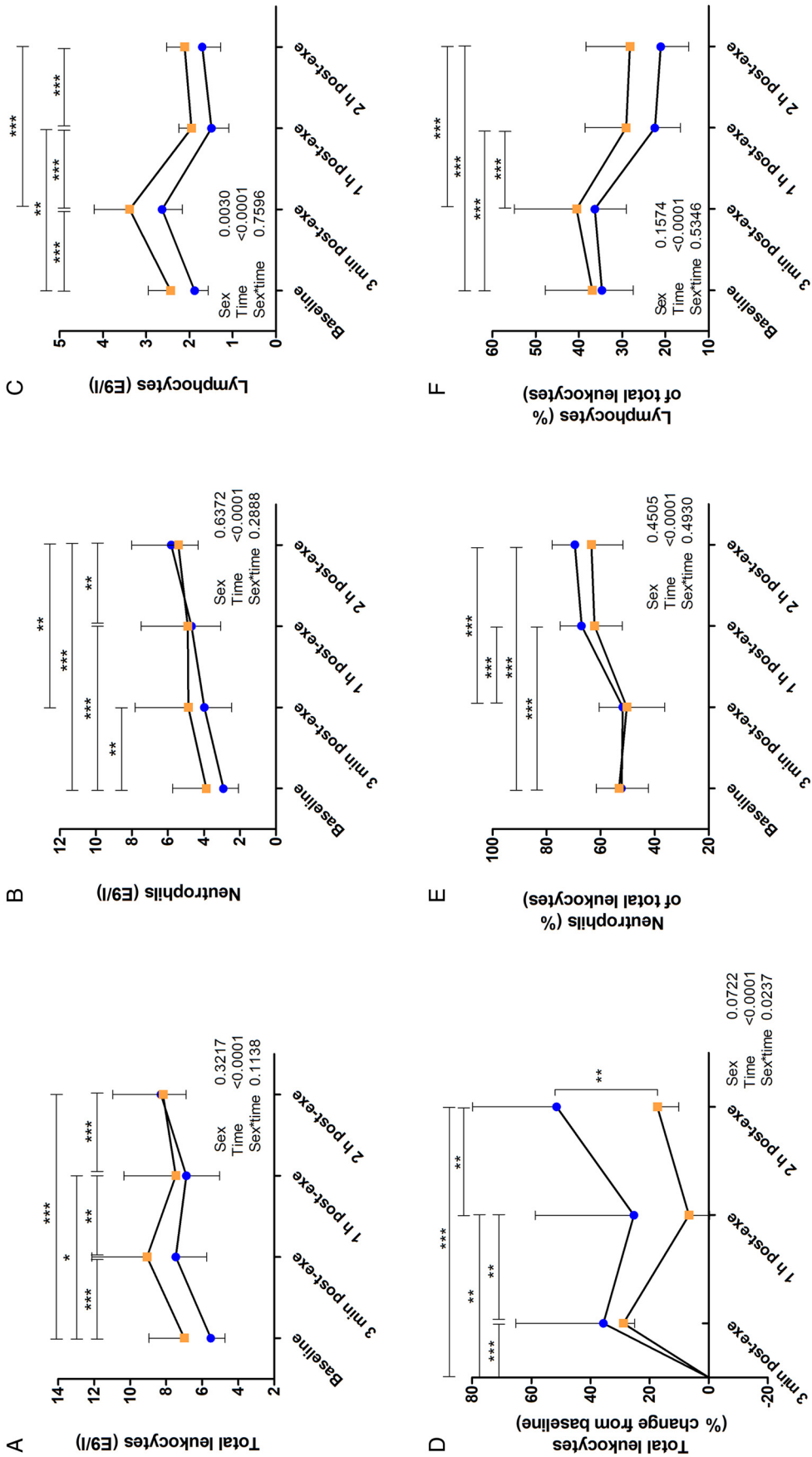
Figure 4 represents individual changes in immune cells between baseline and postexercise peak in men and in women. The postexercise peak represents any postexercise time point (3 min, 1 h, 2 h) in which an individual had the highest immune cell number. Both men and women had significant increases in total leukocytes ( $P < .0001$  and  $P = .0006$ , respectively), neutrophils ( $P < .0001$  and  $P = .0039$ , respectively), lymphocytes ( $P = .0008$  and  $P = .0002$ , respectively), monocytes ( $P = .0122$  and  $P = .0040$ , respectively), and basophils ( $P = .0026$  and  $P = .0019$ , respectively; Figure 4A–4D, 4F). Moreover, eosinophil count did not change in either men or women (Figure 4E).

The percentage change of neutrophils, lymphocytes, monocytes, eosinophils, and basophils from baseline are presented in [Supplementary Material](#) (available online).

## Discussion

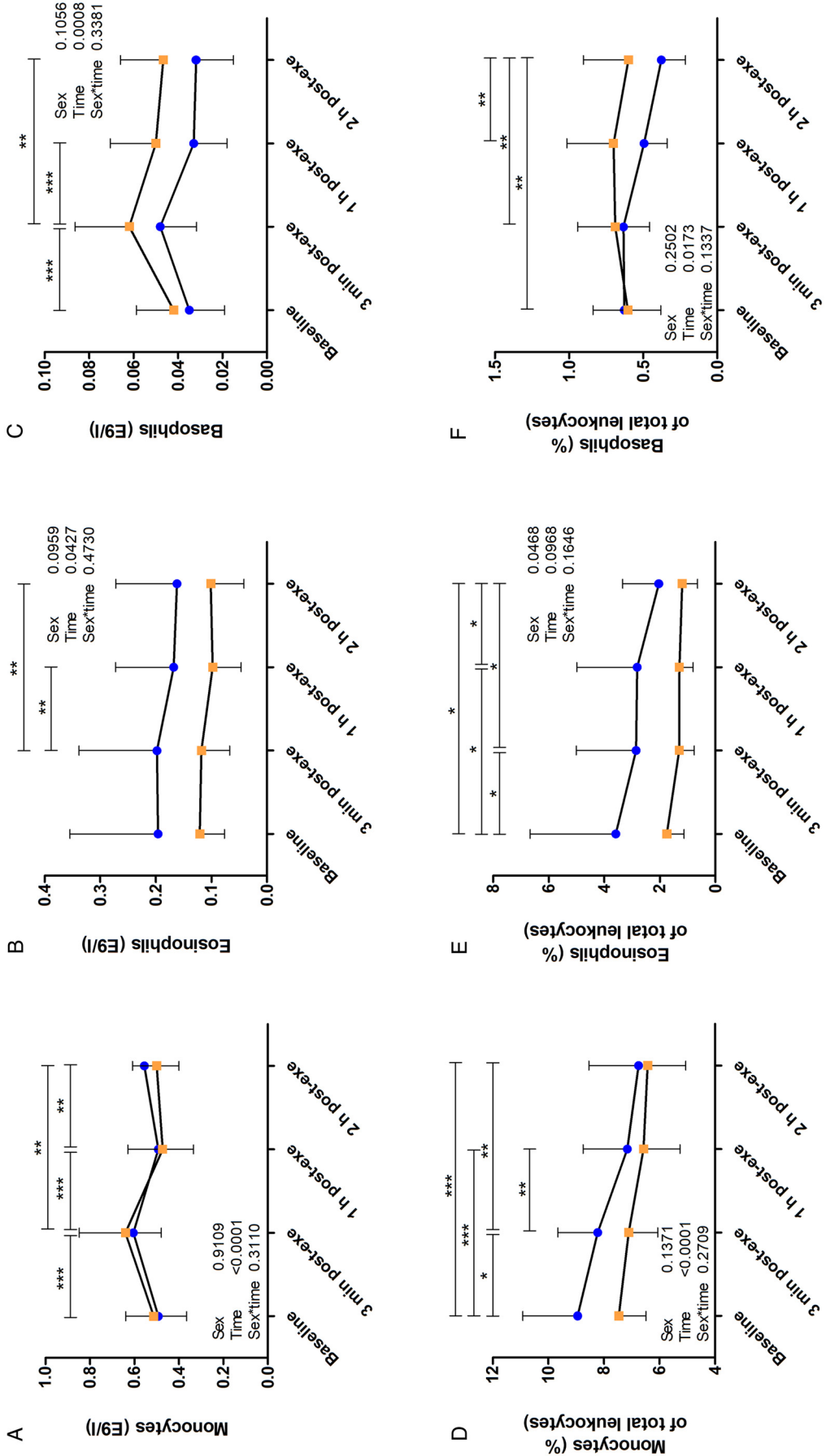
The present study aimed to examine whether white blood cell mobilization after acute exercise differs between sexes. We found that acute exercise increased the number of all leukocyte subsets excluding eosinophils in both sexes. Moreover, the percentage of neutrophils of total leukocytes increased significantly after exercise, while the percentages of lymphocytes, monocytes, eosinophils, and basophils decreased significantly after exercise. Men had a greater increase in the percentage of total leukocytes compared to

—●— Men  
—■— Women

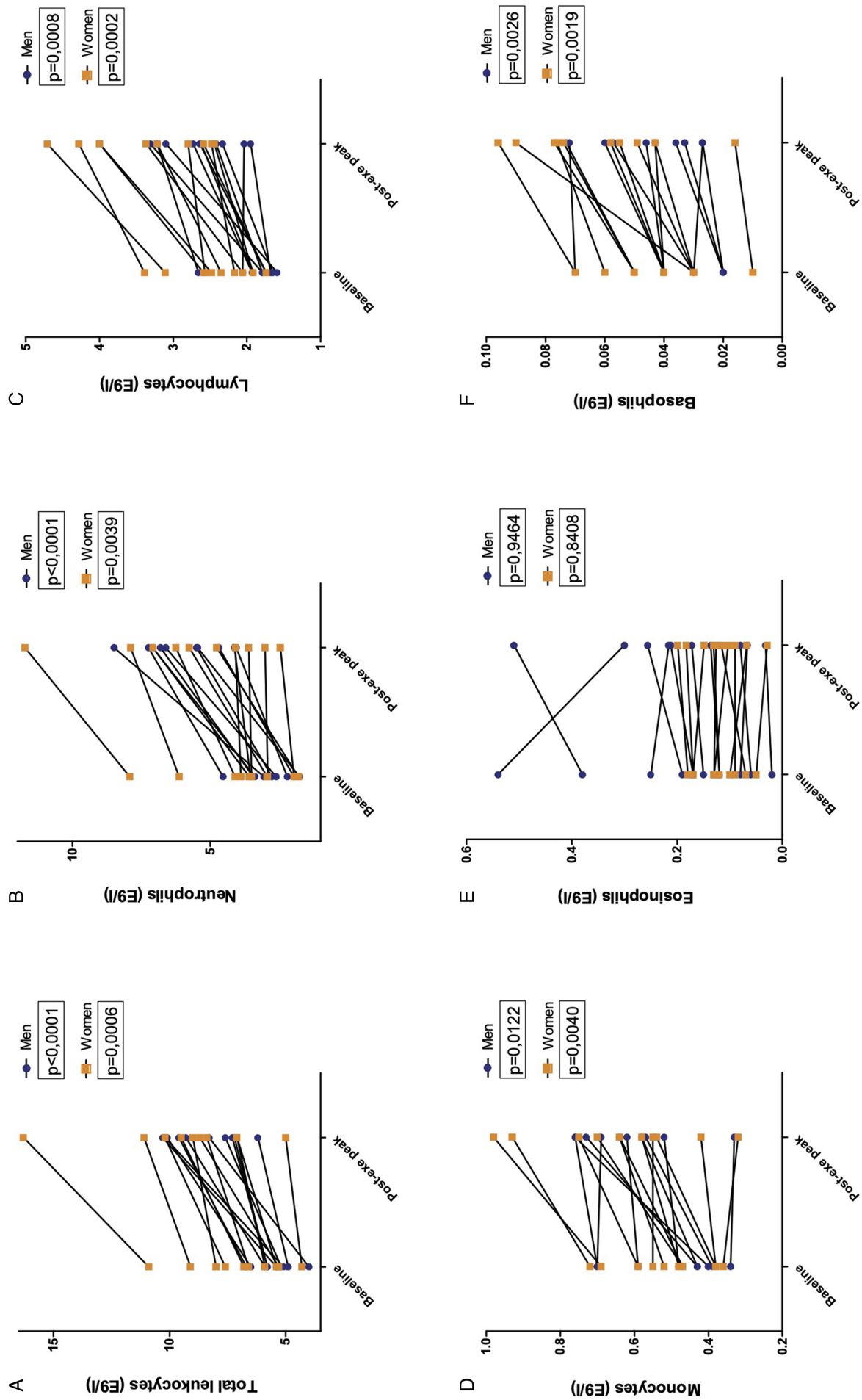


**Figure 2** — Changes in the number of (A) total leukocytes, (B) neutrophils, and (C) lymphocytes; in (D) leukocyte percentage increase from baseline; and in the percentages of (E) neutrophils and (F) lymphocytes of total leukocytes. \* $P < .05$ . \*\* $P < .01$ . \*\*\* $P < .001$ .

● Men  
■ Women



**Figure 3** — Changes in the number of (A) monocytes, (B) eosinophils, and (C) basophils and in the percentages of (D) monocytes, (E) eosinophils, and (F) basophils of total leukocytes. \*  $P < .05$ . \*\*  $P < .01$ . \*\*\*  $P < .001$ .



**Figure 4** — Changes in the number of (A) total leukocytes, (B) neutrophils, (C) lymphocytes, (D) monocytes, (E) eosinophils, and (F) basophils from baseline to peak values postexercise.

women. However, mobilization of different leukocyte subsets was similar in both sexes.

To this date, there are relatively few studies that compare exercise-induced immune cell responses between sexes and some of the previous studies suggest there are no postexercise sex differences in immune cell count or function,<sup>18,26</sup> however the protocols have varied. Barriga et al<sup>18</sup> did not find any sex-differences in leukocyte, neutrophil, lymphocyte, or monocyte counts after 60-minute submaximal exercise at 58% of  $\text{VO}_2\text{max}$  in women and 55% of  $\text{VO}_2\text{max}$  in men. However, they did not find any significant changes in the number of immune cells after the exercise either. Furthermore, Moyna et al<sup>27</sup> reported that lymphocyte function, documented as mitogenic function and cytokine production, following 18-minute continuous incremental cycling exercise at 55%, 70%, and 85% of  $\text{VO}_2\text{max}$  was independent of sex. In contrast to studies reporting no differences, De Lanne et al<sup>28</sup> reported that acute exercise with bicycle ergometer consisting of a 30-minute pedaling at submaximal workload followed by 4-minute period at maximal workload induced leukocytosis that was greater in women compared with men. In the present study, we found that although different leukocyte subsets were affected by exercise, the response was similar in all leukocyte subsets in both sexes. A significant difference between sexes was only observed in percentage change of total leukocytes from rest to 2 hours postexercise. In men the number of leukocytes was 52% above baseline at 2 hours postexercise, whereas in women it was only 19% above baseline. However, our study did not find sex differences in absolute cell counts, which is likely due to women having higher total leukocyte count at baseline compared with men. The fact that any sex differences were not observed in absolute cell counts but were found in percentage change of total leukocytes can also be related to sex hormones and menstrual cycle discussed more below. In fact, menstrual cycle affects baseline values for leukocytes, with significantly higher concentrations observed during the luteal than during the follicular phase,<sup>29</sup> and therefore can contribute to women having higher leukocyte count at rest as compared with men. The current study, however, did not document the menstrual phase of the participants.

The onset of leukocytosis after an acute exercise bout is supported by several previous studies and it is recognized that different subtypes mobilize slightly differently.<sup>30–32</sup> Most of the previous studies, however, were conducted in only male participants.<sup>8,9,11,12,32,33</sup> In the present study, that included both male and females total leukocyte levels, measured as absolute cell count and as percentages increased significantly after 30-minute exercise. Furthermore, the number of neutrophils, lymphocytes, and monocytes increased significantly, having their peaks at 2 hours, 3 minutes, and 3 minutes postexercise, respectively. These results are in line with previous investigations.<sup>30,33</sup> The mobilization of eosinophils and basophils has been studied considerably less. Simonson and Jackson<sup>33</sup> did not find any changes in eosinophil or basophil levels after resistance exercise. However, Gabriel et al<sup>31</sup> found a 36% increase in eosinophil count after exhaustive aerobic exercise at anaerobic threshold and Harries et al<sup>34</sup> found elevated basophil count after interval training. In the present study, we found that basophil count increased significantly at 3 minutes postexercise in both sexes. We also found that in both sexes, eosinophil count was significantly lower at 1 hour and at 2 hours postexercise compared to 3 minutes postexercise, but not compared with baseline.

Current knowledge indicates that menstrual cycle and oral contraception may have an effect on exercise-induced immune responses,<sup>29,35</sup> but based on only few studies and conflicting

results, no firm conclusions can yet be drawn. Timmons et al<sup>17</sup> examined immune cell changes controlled for menstrual phase, oral contraception, and fitness and found that neutrophil, lymphocyte, and monocyte responses to a 90-minute exercise at 65% of  $\text{VO}_2\text{max}$  were greater during luteal phase than during the follicular phase in women using oral contraceptives and also greater than responses in men. Three women in the present study reported hormonal contraceptive use, but because of the low number of the participants and because we did not record menstrual cycle phase, we did not adjust for the use of contraceptives or menstrual cycle. However, not controlling for menstrual cycle and use of contraceptives makes the findings more generalizable to everyday life. Overall, debate has been going on whether studies should control for menstrual cycle or not,<sup>36</sup> but the importance of sex-comparison research is widely recognized.<sup>37</sup> Physiological sex differences have been described previously in, for example, metabolism affecting disease vulnerability<sup>38,39</sup> and fitness and cardiovascular regulation,<sup>40,41</sup> and it appears that general physiological responses to exercise are not major and that there is little evidence that supports exercise training induced adaptations differ substantially between men and women.<sup>42</sup>

Moreover, we acknowledge that our study has other limitations. The number of participants is quite low which may have contributed to the results due to limited statistical power, although around 20 participants is fairly typical in similar physiological studies.<sup>16,17,43</sup> There was a baseline sex-difference in fitness level, which may have had an impact on leukocyte responses to exercise, as seen before.<sup>44</sup> However, there are also studies reporting no difference in exercise-induced leukocyte responses between trained and sedentary people.<sup>45</sup> The sex-difference in our study in  $\text{VO}_2\text{max}$  was only obtained when it was calculated by normalization with whole-body mass.  $\text{VO}_2\text{max}$  calculated by normalization with fat-free mass and maximal power measured as watts per kilogram was similar in both sexes, indicating that the fitness level was not actually that different between sexes, and thus, it is possible, but unlikely, that it affected leukocyte mobilization. The fitness test protocol we used led to 23- and 18-minute tests in average in men and women, respectively, which is considerably longer than recommended for  $\text{VO}_2\text{max}$  testing. This longer duration might have caused fatigue and therefore prevented some participant from reaching a true  $\text{VO}_2\text{max}$ . Furthermore, it is considered that infection history, particularly cytomegalovirus, can determine immune responses to exercise, as people with cytomegalovirus infection have greater T cell responses than those who are not infected.<sup>11,46</sup> To our knowledge, to date, there are no studies evaluating the effects of other infections on exercise-induced immune cell mobilization, but it has been suggested that the effects of exercise on respiratory infections may be gender-dependent.<sup>47</sup> We, however, did not include infection history assessment in our study protocol, which is a limitation and can contribute to the results. Last, we did not examine lymphocyte subsets, such as T cells, and NK cells<sup>44,48–50</sup> which could have given us further valuable information.

One strength in the present study is that we corrected the concentrations of all leukocyte subtypes for plasma volume changes to obtain more accurate results. Exercise can cause acute change in plasma volume, especially if the exercise is of high intensity and long duration.<sup>51,52</sup> This change is thought to result from fluid shift from the plasma into the intracellular space and from loss of fluid in the form of sweat.<sup>53</sup> In the present study, plasma volume decreased 11.7% in men and 14.5% in women at 3 minutes postexercise. Plasma volume was significantly above baseline at 1 hour in men and slightly above baseline in women,

and at 2 hours postexercise plasma volume was close to baseline in both sexes. Participants were allowed to drink water during and after the exercise session, which may partly explain the expansion in plasma volume after exercise. Because of these changes, plasma volume affects the concentrations of blood components including white blood cell<sup>54</sup> and therefore ignoring the change would have led to false results.

Based on the aforementioned strengths and limitations, and current results in relation to the previous literature in the field, we suggest the following points for the future research to advance the topic further: (1) larger sample size studies with different age groups and fitness levels, (2) controlling for menstrual cycle phases and hormonal contraceptive use in female participants, (3) exploring long-term immune adaptations to regular exercise in both sexes, and (4) investigation of other immune markers (eg, cytokines) for a more comprehensive understanding of sex differences in exercise immunology.

## Conclusions

In the present study, we examined sex differences in immune-cell responses to 30-minute individually standardized vigorous-intensity aerobic exercise sessions. We found that acute exercise increased the number of total leukocytes, neutrophils, lymphocytes, monocytes, and basophils in both women and men. Mobilization of all leukocyte subpopulations was similar in both sexes, but the percentage increase in total leukocytes after exercise was greater in men than in women. Our findings suggest that the role of sex in immune-cell mobilization is minor.

## Practical Applications

The findings of this study indicate that the immune system of both sexes adapts and recovers from exercise similarly. The study did not control for menstrual cycle phase, which is a limitation to this study, but also increases the generalizability of the findings. If female athletes want to focus on training depending on their menstrual cycle, future research needs to investigate the effect of the menstrual phase on the exercise-induced responses of the immune system.

## Acknowledgments

This study was supported by Academy of Finland and State Research Funding from the Expert Responsibility Area of Turku University Hospital. We thank Jani Saunavaara, personnel of Turku University Hospital roentgen department, and participants for volunteering their time. **Author Contributions:** Rissanen and I. Heinonen are shared last authorship. *Conceiving and designing research:* I. Heinonen, Rissanen. *Performing experiments:* Neuvonen, Koivula, Sucksdorff, Kapanen. *Analyzing data:* Koivula, Neuvonen, I. Heinonen. *Interpreting results of experiments:* Koivula, I. Heinonen, Neuvonen. *Preparing figures:* Koivula. *Drafting the manuscript:* Koivula. *Editing and revising the manuscript:* All authors. **Data Availability:** The data that support the findings of this study are available from the corresponding author upon reasonable request.

## References

- Shephard RJ. Development of the discipline of exercise immunology. *Exerc Immunol Rev.* 2010;16:194–222. PubMed ID: [20839500](#)
- Shinkai S, Shore S, Shek PN, Shephard RJ. Acute exercise and immune function. *Int J Sports Med.* 1992;13(6):452–461. PubMed ID: [1428375](#) doi:[10.1055/s-2007-1021297](#)
- Graff RM, Kunz HE, Agha NH, et al.  $\beta$  2-adrenergic receptor signaling mediates the preferential mobilization of differentiated subsets of CD8+ T-cells, NK-cells and non-classical monocytes in response to acute exercise in humans. *Brain Behav Immun.* 2018;74: 143–153. PubMed ID: [30172948](#) doi:[10.1016/j.bbi.2018.08.017](#)
- Pedersen BK, Hoffman-Goetz L. Exercise and the immune system: regulation, integration and adaptation. *Physiol Rev.* 2000;80(3): 1055–1081. PubMed ID: [10893431](#) doi:[10.1152/physrev.2000.80.3.1055](#)
- Peake JM, Neubauer O, Walsh NP, Simpson RJ. Recovery of the immune system after exercise. *J Appl Physiol.* 2017;122(5): 1077–1087. PubMed ID: [27909225](#) doi:[10.1152/jappphysiol.006.22.2016](#)
- Pedersen L, Idorn M, Olofsson GH, et al. Voluntary running suppresses tumor growth through epinephrine- and IL-6-dependent NK cell mobilization and redistribution. *Cell Metab.* 2016;23(3): 554–562. PubMed ID: [26895752](#) doi:[10.1016/j.cmet.2016.01.011](#)
- Dethlefsen C, Lillielund C, Midtgaard J, et al. Exercise regulates breast cancer cell viability: systemic training adaptations versus acute exercise responses. *Breast Cancer Res Treat.* 2016;159(3):469–479. PubMed ID: [27601139](#) doi:[10.1007/s10549-016-3970-1](#)
- Schlagheck ML, Walzik D, Joisten N, et al. Cellular immune response to acute exercise: comparison of endurance and resistance exercise. *Eur J Haematol.* 2020;105(1):75–84. PubMed ID: [32221992](#) doi:[10.1111/ejh.13412](#)
- Da Silva Neves PR, Dos Santos Tenório TR, Lins TA, et al. Acute effects of high- and low-intensity exercise bouts on leukocyte counts. *J Exerc Sci Fit.* 2015;13(1):24–28. PubMed ID: [32221992](#) doi:[10.1016/j.jesf.2014.11.003](#)
- Gleeson M, Bishop N, Walsh N. *The Effect of Exercise on Blood Leukocyte Number in Book Exercise Immunology.* Routledge; 2013.
- Turner JE, Aldred S, Witard OC, Drayson MT, Moss PM, Bosch JA. Latent cytomegalovirus infection amplifies CD8 T-lymphocyte mobilisation and egress in response to exercise. *Brain Behav Immun.* 2010;24(8):1362–1370. PubMed ID: [20638470](#) doi:[10.1016/j.bbi.2010.07.239](#)
- Steppich B, Dayyani F, Gruber R, Lorenz R, Mack M, Ziegler-Heitbrock HWL. Selective mobilization of CD14+ CD16+ monocytes by exercise. *Am J Physiol Cell Physiol.* 2000;279(3):C578–C586. PubMed ID: [10942707](#) doi:[10.1152/ajpcell.2000.279.3.C578](#)
- Ndon JA, Snyder AC, Foster C, Wehrenberg WB. Effects of chronic intense exercise training on the leukocyte response to acute exercise. *Int J Sports Med.* 1992;13(2):176–182. PubMed ID: [1555910](#) doi:[10.1055/s-2007-1021252](#)
- Robson PJ, Blain AK, Walsh NP, Castell LM, Gleeson M. Effects of exercise intensity, duration and recovery on in vitro neutrophil function in male athletes. *Int J Sports Med.* 1999;20(2):128–130. PubMed ID: [10190775](#) doi:[10.1055/s-2007-971106](#)
- Simpson RJ, Cosgrove C, Chee MM, et al. Senescent phenotypes and telomere lengths of peripheral blood T-cells mobilized by acute exercise in humans. *Exerc Immunol Rev.* 2010;16:40–55. PubMed ID: [20839490](#)
- Morgado JP, Monteiro CP, Matias CN, et al. Sex-based effects on immune changes induced by a maximal incremental exercise test in well-trained swimmers. *J Sport Sci Med.* 2014;13(3): 708–714.
- Timmons BW, Hamadeh MJ, Devries MC, Tarnopolsky MA. Influence of gender, menstrual phase, and oral contraceptive use on

- immunological changes in response to prolonged cycling. *J Appl Physiol*. 2005;99(3):979–985. PubMed ID: [15879167](#) doi:[10.1152/jappphysiol.00171.2005](#)
18. Barriga C, Pedrera MI, Maynar M, Maynar J, Ortega E. Effect of submaximal physical exercise performed by sedentary men and women on some parameters of the immune system. *Rev Esp Fisiol*. 1993;49(2):79–85. PubMed ID: [8378588](#)
  19. Lobo LF, de Moraes MG, Marcucci-Barbosa LS, et al. A single bout of fatiguing aerobic exercise induces similar pronounced immunological responses in both sexes. *Front Physiol*. 2022;12:833580. doi:[10.3389/fphys.2022.833580](#)
  20. Weitowich NC, Beery A, Woodruff T. Meta-research: a 10-year follow-up study of sex inclusion in the biological sciences. *Elife*. 2020;9:56344. doi:[10.7554/eLife.56344](#)
  21. Cortes CJ, De Miguel Z. Precision exercise medicine: sex specific differences in immune and CNS responses to physical activity. *Brain Plast*. 2022;8(1):65–77. PubMed ID: [36448044](#) doi:[10.3233/BPL-220139](#)
  22. Durmin JV, Womersley J. Body fat assessed from total body density and its estimation from skinfold thickness: measurements on 481 men and women aged from 16 to 72 years. *Br J Nutr*. 1974;32(1):77–97. PubMed ID: [4843734](#) doi:[10.1079/BJN19740060](#)
  23. Tanaka H, Monahan KD, Seals DR. Age-predicted maximal heart rate revisited. *J Am Coll Cardiol*. 2001;37(1):153–156. PubMed ID: [11153730](#) doi:[10.1016/S0735-1097\(00\)01054-8](#)
  24. American College of Sports Medicine. *ACSM's Guidelines for Exercise Testing and Prescription*. 6th ed. 2000.
  25. Dill DB, Costill DL. Calculation of percentage changes in volumes of blood, plasma, and red cells in dehydration. *J Appl Physiol*. 1974;37(2):247–248. PubMed ID: [4850854](#) doi:[10.1152/jappl.1974.37.2.247](#)
  26. Meksawan K, Venkatraman JT, Awad AB, Pendergast DR. Effect of dietary fat intake and exercise on inflammatory mediators of the immune system in sedentary men and women. *J Am Coll Nutr*. 2004;23(4):331–340. doi:[10.1080/07315724.2004.10719376](#)
  27. Moyna NM, Acker GR, Fulton JR, et al. Lymphocyte function and cytokine production during incremental exercise in active and sedentary males and females. *Int J Sports Med*. 1996;17(8):585–591. PubMed ID: [8973979](#) doi:[10.1055/s-2007-972899](#)
  28. De Lanne R, Barnes JR, Brouha L. Hematological changes during muscular activity and recovery. *J Appl Physiol*. 1960;15(1):31–36. doi:[10.1152/jappl.1960.15.1.31](#)
  29. Notbohm HL, Moser F, Goh J, Feuerbacher JF, Bloch W, Schumann M. The effects of menstrual cycle phases on immune function and inflammation at rest and after acute exercise: a systematic review and meta-analysis. *Acta Physiol*. 2023;238(4):e14013. doi:[10.1111/apha.14013](#)
  30. Gabriel H, Urhausen A, Kindermann W. Mobilization of circulating leucocyte and lymphocyte subpopulations during and after short, anaerobic exercise. *Eur J Appl Physiol Occup Physiol*. 1992;65(2):164–170. PubMed ID: [1382982](#) doi:[10.1007/BF00705075](#)
  31. Gabriel H, Schwarz L, Born P, Kindermann W. Differential mobilization of leucocyte and lymphocyte subpopulations into the circulation during endurance exercise. *Eur J Appl Physiol*. 1992;65(6):529–534. PubMed ID: [1382982](#) doi:[10.1007/BF00602360](#)
  32. Shek PN, Sabiston BH, Buguet A, Radomski MW. Strenuous exercise and immunological changes: a multiple-time-point analysis of leukocyte subsets, CD4/CD8 ratio, immunoglobulin production and NK cell response. *Int J Sports Med*. 1995;16(7):466–474. PubMed ID: [8550256](#) doi:[10.1055/s-2007-973039](#)
  33. Simonson SR, Jackson CG. Leukocytosis occurs in response to resistance exercise in men. *J Strength Cond Res*. 2004;18(2):266–271. PubMed ID: [15142013](#)
  34. Harries MG, Burge PS, O'Brien I, Cromwell O, Pepys J. Blood histamine levels after exercise testing. *Clin Allergy*. 1979;9(5):437–441. PubMed ID: [115619](#) doi:[10.1111/j.1365-2222.1979.tb02506.x](#)
  35. Gillum TL, Kuennen MR, Schneider S, Moseley P. A review of sex differences in immune function after aerobic exercise. *Exerc Immunol Rev*. 2011;17:104–121. PubMed ID: [21446354](#)
  36. Giersch GEW, Charkoudian N, Pereira T, et al. Commentaries on point: counterpoint: investigators should/should not control for menstrual cycle phase when performing studies of vascular control. *J Appl Physiol*. 2020;129(5):1122–1135. PubMed ID: [33197376](#) doi:[10.1152/jappphysiol.00809.2020](#)
  37. Robinson AT, Wenner MM, Bunsawat K, Watso JC, Giersch GEW, Charkoudian N. When it's time for the sex talk, words matter. *Am J Physiol Heart Circ Physiol*. 2022;322(1):H66–H70. doi:[10.1152/ajpheart.00556.2021](#)
  38. Clegg DJ, Mauvais-Jarvis F. An integrated view of sex differences in metabolic physiology and disease. *Mol Metab*. 2018;15:11. PubMed ID: [30032908](#) doi:[10.1016/j.molmet.2018.06.011](#)
  39. Tramunt B, Smati S, Grandgeorge N, et al. Sex differences in metabolic regulation and diabetes susceptibility. *Diabetologia*. 2020;63(3):453–461. PubMed ID: [31754750](#) doi:[10.1007/s00125-019-05040-3](#)
  40. Harris RA, Tedjasaputra V, Zhao J, Richardson RS. Premenopausal women exhibit an inherent protection of endothelial function following a high-fat meal. *Reprod Sci*. 2012;19(2):221–228. PubMed ID: [22383760](#) doi:[10.1177/1933719111418125](#)
  41. Skinner BD, Weaver SRC, Lucas SJE, Lucas RAI. Menstrual phase influences cerebrovascular responsiveness in females but may not affect sex differences. *Front Physiol*. 2022;13:1035452. doi:[10.3389/fphys.2022.1035452](#)
  42. Barnes JN, Fu Q. Sex-specific ventricular and vascular adaptations to exercise. *Adv Exp Med Biol*. 2018;1065:329–346. PubMed ID: [30051394](#) doi:[10.1007/978-3-319-77932-4\\_21](#)
  43. Ferrer MD, Tauler P, Sureda A, Tur JA, Pons A. Antioxidant regulatory mechanisms in neutrophils and lymphocytes after intense exercise. *J Sports Sci*. 2009;27(1):49–58. PubMed ID: [19031335](#) doi:[10.1080/02640410802409683](#)
  44. Brown FF, Bigley AB, Sherry C, et al. Training status and sex influence on senescent T-lymphocyte redistribution in response to acute maximal exercise. *Brain Behav Immun*. 2014;39:152–159. PubMed ID: [24200513](#) doi:[10.1016/j.bbi.2013.10.031](#)
  45. Brahmi Z, Thomas JE, Park M, Park M, Dowdeswell IR. The effect of acute exercise on natural killer-cell activity of trained and sedentary human subjects. *J Clin Immunol*. 1985;5(5):321–328. PubMed ID: [3932453](#) doi:[10.1007/BF00918251](#)
  46. LaVoy EC, Hussain M, Reed J, et al. T-cell redeployment and intracellular cytokine expression following exercise: effects of exercise intensity and cytomegalovirus infection. *Physiol Rep*. 2017;5(1):e13070. doi:[10.14814/phy2.13070](#)
  47. Bauza Rodriguez DE, Silveyra P. Sex differences in exercise-induced effects on respiratory infection and immune function. *Eur Med J*. 2024;2(1):52–59.
  48. Koivula T, Lempiäinen S, Rinne P, et al. Acute exercise mobilizes CD8+ cytotoxic T cells and NK cells in lymphoma patients. *Front Physiol*. 2023;13:1078512. doi:[10.3389/fphys.2022.1078512](#)
  49. Koivula T, Lempiäinen S, Rinne P, et al. The effect of acute exercise on circulating immune cells in newly diagnosed breast cancer patients. *Sci Rep*. 2023;13(1):6561. doi:[10.1038/s41598-023-33432-4](#)
  50. Moyna NM, Acker GR, Weber KM, et al. Exercise-induced alterations in natural killer cell number and function. *Eur J Appl Physiol Occup Physiol*. 1996;74(3):227–233. PubMed ID: [8897029](#) doi:[10.1007/BF00377445](#)

51. Collins MA, Hill DW, Cureton KJ, DeMello JJ. Plasma volume change during heavy-resistance weight lifting. *Eur J Appl Physiol Occup Physiol*. 1986;55(1):44–48. PubMed ID: [3698986](#) doi:[10.1007/BF00422891](#)
52. Boulay MR, Song TM, Serresse O, Thériault G, Simoneau JA, Bouchard C. Changes in plasma electrolytes and muscle substrates during short-term maximal exercise in humans. *Can J Appl Physiol*. 1995;20(1):89–101. PubMed ID: [7742773](#) doi:[10.1139/h95-007](#)
53. Von Duvillard SP, Braun WA, Markofski M, Beneke R, Leithäuser R. Fluids and hydration in prolonged endurance performance. *Nutrition*. 2004;20(7–8):651–656. PubMed ID: [15212747](#) doi:[10.1016/j.nut.2004.04.011](#)
54. Kargotich S, Goodman C, Keast D, Morton AR. The influence of exercise-induced plasma volume changes on the interpretation of biochemical parameters used for monitoring exercise, training and sport. *Sports Med*. 1998;26(2):101–117. doi:[10.2165/00007256-199826020-00004](#)